Review Article

Journal of Advanced Veterinary Research (2023) Volume 13, Issue 1, 150-156

Listeriosis in Rabbits (*Oryctolagus cuniculus*): A Significant Bacterial Disease with an Emerging Zoonosis

Wafaa A. Abd El-Ghany*

Poultry Diseases Department, Faculty of Veterinary Medicine, Cairo University, Giza, 12211, Egypt.

*Correspondence

Wafaa A. Abd El-Ghany Poultry Diseases Department, Faculty of Veterinary Medicine, Cairo University, Giza, 12211, Egypt. E-mail address: wafaa.soliman@cu.edu.eg

Abstract

Rabbit's rearing becomes one of the most important source of income in many countries world-wide. Rabbit's meat has many advantageous over any time of meat and it is recommended especially for sick people. Rabbits are susceptible to some important bacterial diseases such as listeriosis which causes severe economic losses. Listeriosis is a disease caused by many species of Listeria which may be environmental saprophytes or sometimes pathogenic for mammals, birds, and humans. Listeria monocytogenes (L. monocytogenes) is the most common cause of illness either in animals or humans. Infection with this bacterium usually occur in animals via ingestion, while in human via handling or consumption of uncooked or under-processed food. Most of signs in rabbits and humans are the same and including septicaemia especially in young, meningoencephalitis, and metritis and fetal mortality in pregnancy. Additionally, human may show the signs of severe febrile gastroenteritis due to food borne illness as well as skin and joint affections. Diagnosis of the disease depends mainly on the traditional methods of isolation and identification along with the recent molecular techniques for detection. Serological diagnosis is of little value. Prevention and control of listeriosis in rabbits are based on cleaning and disinfection of rabbit's cages, using of specific antibiotic treatment, and application of some natural alternatives such as probiotics and phytobiotics. In human, avoidances of the possible causes of Listeria infection and stressors and administration of a specific drug is the must. Besides, improvement of the quality control measures in processing plants or during handling of food are essential to effectively prevent and control listeriosis associated with food-borne illness. Accordingly, this article focuses on listeriosis in rabbits regarding the pathogenesis and virulence of the causative agent, clinical picture in rabbits, zoonotic importance in human, laboratory diagnosis, and the different strategies used for the prevention and control of the disease.

KEYWORDS

Listeria monocytogenes, Rabbits, Human, Virulence, Control

INTRODUCTION

Rabbit's industry has been developed in many countries due to several advantageous over other animal species including good feed conversion ratio, fast growth rate, high fertility rate, short production cycle, and big productive capability (Cullere and Dalle Zotte, 2018). In addition, rabbit's meat has gained popularity as a very healthy meat source due to ease digestibility, high level of protein, and low levels of fat, cholesterol, and sodium (Dalle Zotte and Szendro, 2011; Wang *et al.*, 2019). However, rabbits are vulnerable to many important bacterial diseases that affect health and productivity such as listeriosis.

Listeriosis, silage disease, circling disease, or meningoencephalitis is caused by *Listeria monocytogenes* (*L. monocytogenes*). The bacterium was first isolated in 1926 from the livers of diseased rabbits (Murray *et al.*, 1926). Later in 1966, *L. monocytogenes* was considered as a major foodborne bacterium and many experimental infections were done on different animal species to prove its pathogenicity (Gray and Killinger, 1966). *Listeria monocytogenes* is one of the most fastidious and opportunistic pathogen associated with life-threating listeriosis in a wide range of animals, birds, fish, and human being (Wesley, 2007; Johansson and Freitag, 2019). This pathogen is a facultative intracellular organism which is widely distributed in the environment. The bacterium may persist for many years in the facilities, grows at a wide temperature range from 0.5°C to 45°C, and tolerates environment with high salts or acids. *Listeria monocytogenes* is a Gram-positive, small, motile, non-sporulating, and bacillary to coccobacilli bacterium. Ingestion of contaminated food is regarded as the main route of the listeriosis transmission (Schlech *et al.*, 1983). Listeriosis is commonly associated with septicaemia, meningitis, meningoencephalitis, brain stem encephalitis, abortion, stillbirth, and gastro-enteric infections in the affected hosts (Barbuddhe and Chakraborty, 2009; OIE, 2014).

Rabbits is regarded as a natural host for *L. monocytogenes* and the meat of affected animals is an important source of human food-borne infection (De Cesare *et al.*, 2017; Zhao *et al.*, 2020). Listeriosis has gained a great concern in rabbits due to its adverse effects on production as well as the food-born zoonosis and public health importance (Dhama *et al.*, 2013; Johansson and

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made. ISSN: 2090-6277/2090-6269/ © 2011-2023 Journal of Advanced Veterinary Research. All rights reserved.

Freitag, 2019). Infection of rabbits with *L. monocytogenes* causes infertility, abortion, and high mortality with severe losses in rabbitaries (Abdel Moteleb *et al.*, 1990; Abd El-Ghaffar and Abd-El-Gwad, 1997). Despite *L. monocytogenes* may show low incidence rate, it often associated with high mortalities (Radoshevich and Cossart, 2018). The pathogen was isolated from diseased farmed rabbits in different localities including Egypt with incidences 20% or more (Abd El-Waneas, 1985; Abd El-Ghaffar and Abd El-Gwad, 1997; Hatab and Abd El-Latif, 2006; Moursi *et al.*, 2006; Ibrahim and Ibrahim, 2016).

Therefore, this article focuses on listeriosis in rabbits regarding the pathogenesis and virulence of the causative bacterium, clinical picture in rabbits, zoonotic importance in human, laboratory diagnosis, and the different strategies used for the prevention and control of the disease.

PATHOGENESIS AND VIRULENCE

The intracellular nature of *L. monocytogenes* and the weak intracellular diffusion of many antimicrobials increase the ability of the pathogen to resist the environmental conditions and create difficulties in the eradication programs (Stratakos *et al.*, 2020). The bacterium can survive at a wider range of temperature (4°C to 37°C), pH, and water activities (Janakiraman, 2008). Moreover, *L. monocytogenes* can grow in the presence or absence of oxygen and multiply at temperature of 0°C.

The soil, decayed tissues, or water are the sources of *L. monocytogenes* infection. The bacterium can enter the body via ingestion, inhalation, broken sin, or eyes. Un-processed or under-cooked food is an important source of human's infection. The predilection sites of *L. monocytogenes* are the intestinal wall, brain (medulla oblongata), and placenta of animals. The pathogen may cause encephalitis via tiny wounds in the buccal mucosa of infected animal.

It has been detected that *L. monocytogenes* is not acid tolerant and a large portion of the bacterium is likely to be killed in the stomach of the infected animal (D'Orazio, 2014). *Listeria monocytogenes* may invade the intestinal barrier through active endocytosis of the endothelial cells. This mechanism occurs via the interaction between the bacterial cell surface protein (internalin) and the epithelial cell surface receptor (E-cadherin). The bacterial cell could be passed from the intestinal cell to another in the host without exposure to the immune system which represented by antibodies, complement, or neutrophils. This could be achieved via pushing of the bacteria against the cell membrane and formation of pseudopods that ingested by some cells such as macrophages, enterocytes, and hepatocytes.

The virulence of *L. monocytogenes* is related to the ability of the pathogen to invade, adhere, and translocate across the barriers of the intestine during the gastrointestinal phase of infection (Radoshevich and Cossart, 2018). Consequently, hampering the bacterium during this phase is the best way to inhibit its spread to the deeper tissues and subsequent lethality. Moreover, *L. monocytogenes* has an intracellular life cycle which allows the pathogen to pass from one cell to another without leaving the cell. Moreover, presence of haemolytic factor (hlyA gene) which is known as listeriolysin O (Khan *et al.*, 2013; Ibrahim and Ibrahim, 2016) and invasion associated protein (iap gene) are associated with the pathogenicity and virulence of *L. monocytogenes* (Wuenscher *et al.*, 1993).

It has been reported that *L. monocytogenes* may proliferate in the liver, spleen, and immune organs (Regan *et al.*, 2014) and also it can cross the placental and blood-brain barriers (Janakiraman, 2008) until attacking by the host's immune defense. During the

early stage of infection, *L. monocytogenes* infected cells stimulate the infiltration of some inflammatory cells such as monocytes resulting in the formation of abscesses, and after the next days, the bacterium could be inactivated mainly by T-cells (Bortolussi *et al.*, 1984).

The central nervous system is also a target for *L. monocytogenes* that causes meningitis, rhombencephalitis, and brain abscesses (Southwick and Purich, 1996, Vazquez-Boland *et al.*, 2001; Orndorff *et al.*, 2006). Infected animals show increased level of lactate dehydrogenase enzyme released from the cerebrospinal fluid causing brain tissue damage and/or increased permeability of the blood-brain barrier (Abate *et al.*, 1998; Pamukcu *et al.*, 2004). Furthermore, presence of inflammatory cells such as neutrophils, macrophages, and lymphocytes indicates the important role of these inflammatory cells in the brain tissue injury (Taha *et al.*, 1991; Kruger *et al.*, 1995; Najdenski and Vesselinova, 2002; Kabakci and Yarim, 2004). Inflammatory cells are related to secretion of pro-inflammatory cytokines and formation of toxic free radicals such as nitric oxides which is essential for the degeneration of the brain tissue (Greenberg *et al.*, 1998; Shin *et al.*, 2000).

Prompting of pro-inflammatory cytokines in rabbits in response to *L. monocytogenes* has been studied (Zhao *et al.*, 2020). It has been reported that *L. monocytogenes* might upregulate pro-inflammatory cytokines genes such as tumor necrosis factor (TNF- α) and interleukin (IL-6), thus lead to increasing the permeability of the intestinal epithelium (Drolia *et al.*, 2018). Moreover, the initial production of interferon gamma (IFN γ) is a very important step for stimulation of the immune response and regulation of *L. monocytogenes* infection (Harty and Bevant, 1995). In this context, increasing the expression levels of TNF- α , IFN γ , IL-6, IL-8, and IL-1 β genes have been reported in *L. monocytogenes* infected rabbits compared with those infected and treated with either enrofloxacin or probiotics (Abd EI-Hamid *et al.*, 2022).

CLINICAL PICTURE

The forms of listeriosis are usually represented by septicaemia in bunnies, meningoencephalitis in adults, and metritis and fetal mortality in pregnant does (Vazquez-Boland *et al.*, 2001; Malik *et al.*, 2002).

Encephalitis is usually occur when *L. monocytogenes* ascends the trigeminal nerve causing localized infection of the brain stem. The severity of nervous signs vary according to the degree of neurons damage and the signs are usually manifested by trigeminal and facial nerve paralysis and circling (Kahn *et al.*, 2006). The encephalitic form of listeriosis in rabbits is known as 'circling or rolling disease' as a result of walking of the animal around in circles in one direction with the head on "sideways" (OIE, 2014). Incoordination, loss of equilibrium, disorientation, immobility, and rolling sideways are another signs of the disease in the affected bunnies. The nervous signs may persist for some days or weeks, but the complete recovery is uncommon. Sometimes, affected bunnies may suddenly die without showing nervous signs.

Pregnant does may show purulent conjunctivitis, loses of weight, and lastly abortion (Hatab and Abd El-Latif, 2006). Intra-uterine death of feti or even death just after birth has been also reported (Okerman, 1999; Patton *et al.*, 2000). Abortion associated listeriosis is due to the ability of the causative bacterium to cause myometrial contraction (Baker, 1998).

Experimental oral inoculation of 2-months-old rabbits with *L. monocytogenes* induced signs 2 weeks post-infection (Ibrahim and Ibrahim, 2016). Infected rabbits showed loss of appetite, depression, ruffled fur, lacrimation, conjunctivitis, urine retention, diarrhea, and finally emaciation followed by death. Moreover,

nervous signs in the form of lateral deviation of the head and neck, convulsions, and death were also observed. Similar nervous signs have been also observed in rabbits following *L. monocytogenes* infection (Hoelzer *et al.*, 2012; Ahmed, 2013). Oral or conjunctival inoculation with *L. monocytogenes* produced abortion in pregnant does (Gray and Killinger, 1966). Besides, abscess formation has been demonstrated in rabbits following *L. monocytogenes* subcutaneous inoculation (Gray and Killinger, 1966).

Significant pathological alterations in the livers, spleen, and brains of rabbits could be observed as a result of septicemia induced by *L. monocytogenes* infection (Watson and Evans, 1985; Abd El-Hamid *et al.*, 2022). Dead rabbits with listeriosis showed severe generalized congestion of liver, spleen, kidneys, and lungs, along with softening of the brain (Okerman, 1999; Vazquez-Boland *et al.*, 2001; Moursi *et al.*, 2006; Ahmed, 2013; Ibrahim and Ibrahim, 2016). Necrosis of the live and spleen, atrophy of spleen, enteritis, cystitis, bloody vaginal discharge, peritonitis, endometritis, meningitis, and hydrocephalus could be also demonstrated in rabbits following such infection (Abd El-Ghaffar and Abd-El-Gwad, 1997). Listeriosis induced severe lesions in the mid brain, less severe lesions in the cerebellum, and rare lesions in the cerebrum (Pamukcu *et al.*, 2004).

ZOONOSIS

Contamination of food with *L. monocytogenes* may occur during processing, handling, and packaging under poor quality control measures (Carpentier and Cerf, 2011). The pathogen is widely distributed in ready-to-eat and raw meat products (Mengesha *et al.*, 2009). Moreover, direct contact with the contaminated materials such as aborted feti may be a possible way of *L. monocytogenes* infection in human. Food-born listeriosis and deaths of workers in retail and food service environments have been reported in many localities (Lianou and Sofos, 2007; Meloni *et al.*, 2009). Human listeriosis may cause severe and life-threatening complications (Dhama *et al.*, 2013). The disease in human is caused by all the 13 serotypes, especially 1/2a, 1/2b, and 4b, and the annual endemic infection rate varies from 2 to 15 cases/ millions of human (Munoz, 2012).

The clinical syndromes of listeriosis in people may appear

as sporadic, endemic, or foodborne outbreak. The disease usually induces fatigue, chills, headache, and febrile gastroenteritis. Severe cases of listeriosis are associated with septicaemia, skin rashes, meningoencephalitis, abortion, infection of other organs, and finally death if the disease is neglected without probable treatment (Koopmans *et al.*, 2014). Young and elderly individuals, pregnant women, and immuno-deficient debilitated people are at higher risk to *L. monocytogenes* serious illness, while healthy persons are usually vulnerable to gastroenteritis and pregnancy losses (Poulsen and Czuprynski, 2013; Popovic *et al.*, 2014). Table 1, shows the different forms of listeriosis in humans.

LABORATORY DIAGNOSIS

Selection of the proper samples from infected rabbits is mainly depend on the prevalent signs in the infected farm. Samples could be collected from the aborted fetus, placenta, uterine discharges, blood, cerebrospinal fluid, brain, conjunctiva, liver, kidneys, spleen, and brain.

Isolates of *L. monocytogenes* grow on selective media such as Fraser broth after incubation at 37°C for 24 hrs followed by sub-culturing on PALCAM and incubation at 37°C for 24-48 hrs (Fraser and Sperber, 1988). Typical colonies appear as grey green with black sunken centers (Holt *et al.*, 1994). The bacterium can grow at temperatures as low as -2°C in laboratory media broth (Bajard *et al.*, 1996). On sheep blood agar, *L. monocytogenes* induces a complete zone of β -hemolysis which appears near to the line of the streak of *Staphylococcus aureus* (*S. aureus*) in CAMP test (Volokhovel *et al.*, 2007).

Microscopic examination of suspected *L. monocytogenes* colonies revealed presence of non-sporulating Gram positive rods that arranged singly, in short chains, in pairs at V shape angle, or in parallel groups (Hass and Kreft, 1988; Warbureton *et al.*, 2003).

The different biochemical reactions of *L. monocytogenes* are shown in Table 2. On semi solid agar media, *L. monocytogenes* strains show umbrella growth pattern with a characteristic tumbling motility using peritrichous flagella (Quinn *et al.*, 2002).

The Anton's test revealed that inoculation of two to three drops of *L. monocytogenes* in the eyes of rabbits induced purulent conjunctivitis and keratitis 24-48 hrs post-inoculation (An-

Form	Signs		
Gastrointestinal	Abdominal pain, vomiting, diarrhea, and flue like signs such as fever, nausea, headache, and myalgia. Most of these cases are usual self-limiting within few days.		
Nervous	Meningitis, meningoencephalitis, rhombencephalitis or brain abscesses.		
	Unilateral or bilateral cranial nerve deficits, ataxia, cerebellar dysfunction, hemiparesis, impaired consciousness, and death.		
	Sometimes death result from cardiac and respiratory failure.		
Reproductive	Pregnant women may remain asymptomatic or shows mild flu-like signs and/or gastroenteritis. However, she may later abort or give birth to a stillborn or premature infant.		
	Neonates usually have septicemia and sometimes granulomas, skin lesions and/or abscesses. Pneumonia and meningitis are less commonly reported syndromes. In the later stage, meningitis is most common.		
Skin	Rashes, papule, vesicle, or pustule which may be localized and non-pruritic, and may sometimes be disseminated especially in im- munosuppressed persons. Some cases showed fever, chills, lymphadenopathy, and general pain.		
Bone and joint	Arthritis and osteomyelitis		
Eye	Conjunctivitis, keratoconjunctivitis, chorioretinitis, and unilateral endophthalmitis.		
Others	Cholecystitis, cholangitis, biliary cyst, hepatitis or liver abscesses.		
	Pyelonephritis.		
	Endocarditis.		
	Peritonitis		
	Pneumonia and pleuritis		

Table 1. Clinical forms of listeriosis in humans

ton, 1934). Moreover, the pathogenicity test in mice showed that intraperitoneal injection of *L. monocytogenes* lead to 100% mortality rate within 24-48 hrs after injection with severe congestion of the internal organs and brain (Seeliger and Jones, 1986; Marco *et al.*, 1992).

Table 2. The biochemical reactions of Listeria monocytogenes.

Reaction	Result
Catalase	+
Vogues Proskaur	+
Oxidase	-
Urease	-
Indole	-
H ₂ S production	-
Sugar fermentation	
Maltose	+
Sucrose	+
Dextrose	+
Sorbitol	+
Xylose	-
lactose	±
Mannitol	±

+: Positive; - = Negative

Immunohistochemistry, immunofluorescence, polymerase chain reaction, or loop-mediated isothermal amplification may provide a more rapid diagnosis of *L. monocytogenes* in the tissues. Moreover, *L. monocytogenes* could be subtyped genetically using pulsed field gel electrophoresis, whole genome sequencing, DNA hybridization, multi-virulence-locus sequence typing, or multilocus sequence typing. These testes are usually used in epidemiological investigations of human listeriosis and detection of out-breaks sources.

Serological tests can detect immunoglobulins (IgG) to listeriolysin. Serological diagnosis of listeriosis using agglutination or enzyme linked immunosorbent assay test is not common because most of cases are caused by few common serotypes, and each of these serotypes can contain more than one Listeria species. In addition, some apparently healthy rabbits show high antibody titers and cross-reactions with Enterococci species or Staphylococcus aureus (S. aureus) in some tests. The serological response against L. monocytogenes in the absence of clinical signs has been detected (Low and Donachie, 1991). Frequent ingestion of low doses of the bacterium induced high titers of anti-Listeria agglutinins in apparent healthy animals and humans (Seeliger, 1987). Anti-L. monocytogenes antibodies could be produced in rabbits to demonstrate the immune response to the bacterium (Larsen et al., 1974). Belen Lopez et al. (1993) hypothesized the interaction between L. monocytogenes and the host immune system in the tonsils after oral and gastric inoculation of rabbits with the bacteria. The IgM and IgG were produced in rabbits in response to oral inoculation of L. monocytogenes and/or S. aureus with a cross immunity (Larsen et al., 1974). Thirteen serovars of this bacterium are highly virulent and common in animals such as serovars 4b, 1/2a, 1/2b and 3, while serovars 4b, 1/2a, 1/2b, and 1/2c are thought to be the cause of more than 95% of the clinical cases in humans.

PREVENTION AND CONTROL

Once the outbreak of listeriosis occurs in certain countries such as United States of America (USA), Veterinarians should re-

port the national and/or local authority. Diseased rabbits should be isolated and sources of contamination, such as the aborted feti, should be hygienically disposed. Thorough cleaning and disinfection of fomites including water tanks or feeders is the must. Eradication of mechanical vectors such as rodents which can shed *L. monocytogenes* in feces. Minimize stressors and concomitant infections which are suspected to increase exposure to listeriosis.

Listeria organism can be inactivated by heating or autoclaving. Many disinfectants such as quaternary ammonium compounds, glutaraldehyde, ethanol, sodium hypochlorite, iodine, and chlorhexidine can effectively dissolve on the organic matters covering *L. monocytogenes*. Sometimes presence of biofilm layer on the bacterium surface makes destroying of bacterium is difficult especially in food processing facilities.

Improved quality control measures are essential to effectively prevent and control listeriosis associated with food-borne illness (Nakari *et al.*, 2014). Measures including good cooking of animal products, well washing of raw vegetables, and avoidance of undercooked food should be considered to reduce the risk of foodborne listeriosis, Monitoring of environment, testing of products, as well as cleaning and sanitation are also crucial. In certain countries, such as USA, detection of *L. monocytogenes* in any concentration in food prompts a recall, and measures are carried-out.

ANTIBIOTIC TREATMENT

Antibiotic treatment is the most important way for controlling L. monocytogenes. The treatment course varies according to the level of infection. Sometimes, treatment may be difficult because the pathogen can invade all types of cells. Moreover, treatment of encephalitic or immunocompromised host is usually not effective. However, high doses of early medication along with supportive treatment are important for rabbits suffering rhombencephalitis. Monitoring of antibiotic sensitivity is very important (Okada et al., 2011; Barbosa et al., 2013). Penicillin's classes are regarded to be the most effective treatment of L. monocytogenes infection particularly for humans, but other drugs have sometimes been used. Despite the highest effectiveness of penicillin against L. monocytogenes infection, resistance has been reported (Rivero et al., 2003). Some strains of L. monocytogenes showed resistance to cephalosporins, especially cefotaxime (Boisivon et al., 1990). Moreover, L. monocytogenes strains from food, environment, animal, and human origins showed resistance to gentamicin, cotrimixazole, and ofloxacin (Rahimi et al., 2012; Soni et al., 2013; Ndahi et al., 2014). It is important to mention that haphazard using of penicillin for the treatment of listeriosis in farmed animals has been suggested as one of the causes of resistance in Listeria-infected human via animal derived foods (Tiwari et al., 2013).

Though some *L. monocytogenes* strains are sensitive to several antibiotics *In-vitro*, some of these antimicrobials may not be effective *in-vivo*. An early *in-vitro* study revealed that *L. monocytogenes* strains from rabbits-origin were highly sensitive to gentamycin, tetracycline, and spictinomycin, while resistant to chloramphenicol, lincomycine, and streptomycin (Abd El-Ghaffar and Abd-El-Gwad, 1997). Additionally, *L. monocytogenes* isolated from diseased rabbits were highly susceptible to ampicillin, amoxicillin, penicillin, tetracycline, doxycycllin, trimethoprim, neomycin, and gentamycin, but less susceptible to streptomycin, cefotaxime, and nalidixic acid *In-vitro* (Charpentier and Courvalin, 1999; Moursi *et al.*, 2006).

Prophylactic medication with sulphonamides, penicillin, or tetracycline may be effective in prevention of listeriosis (Radostits *et al.*, 2008), while ampicillin and erythromycin are the drugs of choice for treatment.

PROBIOTICS

Antibiotics treatment can harm the gut microbiota and the host immune response, enhances the development of resistant bacteria, and accumulates in the tissues as residues. Accordingly, new antibiotics alternatives have been developed to control L. monocytogenes infection (Suez et al., 2019). Such alternatives include using of probiotics (Amalaradjou and Bhunia, 2012). They may compete with the pathogenic bacteria for adhesion sites, prevent the disruption of intestinal integrity, and enhance the host's immune system, thus conferring resistance to L. monocytogenes (Amalaradjou and Bhunia, 2012). It has been proposed that probiotics block the bacterial attachment and consequently its invasion to the intestinal epithelial cells that resulting in improved growth performance of rabbits and provide protection against L. monocytogenes infection. Drolia et al. (2020) demonstrated that that probiotics contain Lactobacilli species inhibited L. monocytogenes-induced intestinal permeability via the preservation of the cell junctional configuration of claudin-1, occludin, and E-cadherin biomarkers. Besides, this species of probiotic stimulated the secretion of tight junction protein gene (MUC), hence fortified the mucus barrier and decreased L. monocytogenes-induced MUC-2 loss and apoptotic responses (Drolia et al., 2020).

In-vitro studies showed that Lactobacillus (L) producing bacteria such as L. acidophilus, L. plantarum, and Enterococcus faecium (E. faecium), or even their metabolites inhibited the formation of L. monocytogenes biofilm (Rocha et al., 2019). Addition of L. acidophilus at concentration of 10⁹ or 10¹² colony forming unit (CFU)/mL to the milk revealed a significant inhibitory effect L. monocytogenes growth (Ehsani et al., 2019). Moreover, a strain of L. plantarum or its cell-free supernatants significantly prevented the growth of L. monocytogenes, Salmonella enteritidis, Escherichia coli, and S. aureus (Arena et al., 2016). The proliferation of spoilage bacteria such as L. monocytogenes and Salmonella species has been inactivated following inoculation of L. plantarum at a level of 10⁸ CFU/g of raw minced beef meat (Trabelsi et al., 2019). In milk, the multiplication of L. monocytogenes was significantly inhibited by incorporation of E. faecium and its enterocins A and B (Vandera et al., 2017).

Van Zyl et al. (2016) found that *L. plantarum* excluded *L. monocytogenes* from the gut of mice. There are limited data regarding using of probiotics to prevent or treat *L. monocytogenes* infections in farmed rabbits. Zhao et al. (2020) demonstrated that lactic acid bacteria can reduce *L. monocytogenes* infection and modulate the cellular and humoral immune responses of infected rabbits. Recently, treatment of *L. monocytogenes* infected rabbits with multi-strains probiotics (a mixture of *L. acidophilus, Bacillus subtilis*, and *E. faecium* strains) restored the reduced growth and intestinal barriers, decreased the severity of signs and mortalities, attenuated the excessive inflammatory reactions, and ameliorated the microscopic lesions in liver, brain, and spleen compared with enrofloxacin treatment (Abd El-Hamid et al., 2022).

PHYTOBIOTICS

The anti-*Listeria* effect of some herbs and their extracts has been studied. For instance, Thai spices proved efficacy in the treatment of listerosis associated food poisoning (Thongson *et al.*, 2005). In addition, extracts of many plants such as *Prangos ferulacea*, *Allium vineale*, *Chaerophyllum macropodum*, *Sophorae radix*, *Psoraleae semen*, oregano, rosemary, clove, and sage showed high efficacy against *L. monocytogenes* infection (Sagun *et al.*, 2006; Yoon and Choi, 2012; Witkowska *et al.*, 2013). Skariyachan *et al.* (2015) demonstrated that cinnamon extract could be very effective against *L. monocytogenes* via inhibition of the general secretory pathway, thus decreasing the pathogen virulence.

CONCLUSION

Listeriosis is regarded as a disease of clinical and productive significance for rabbits industry. Therefore, the pathogenesis and mechanism of *L. monocytogenes* virulence in rabbit should be more studied and explained. In addition, successful preventive control measures against listeriosis in rabbit farms should be implemented.

CONFLICT OF INTEREST

The author declare that they have no conflict of interest.

REFERENCES

- Abate, O., Bollo, E., Lotti, D., Bo, S., 1998. Cytological, immunocytochemical and biochemical cerebrospinal fluid investigations in selected central nervous system disorders of dogs. Zentralblatt Veterinarmedizin Reihe B 45, 73-85.
- Abd El-Waneas, S.A., 1985. Studies on listeriosis in rabbits. M.V.Sc., Faculty of Veterinary Medicine, Cairo University, Egypt.
- Abd El-Ghaffar, S. Kh., Abd El-Gwad, A.M., 1997. Some studies on *Listeria* monocytogenes in rabbits. Assuit Veterinary Medical Journal 38, 7-19. Available at: https://www.aun.edu.eg/veterinary_medicine/ some-studies-*Listeria*-monocytogenes-rabbits
- Abd El-Hamid, M.I., Ibrahim, D., Hamed, R.I., Nossieur, H.H., Elbanna, M.H., Baz, H., Abd-Allah, E.M., El Oksh, A.S.A., Ibrahim, G.A., Khalifa, E., Ismail, T.A., Awad, N.F.S., 2022. Modulatory impacts of multistrain probiotics on rabbits' growth, nutrient transporters, tight junctions and immune system to fight against *Listeria monocytogenes* infection. Animals (Basel) 12, 2082.
- Abdel Moteleb, T.Y., Salem, B., El-Zanaty, K., 1990. Outbreak of listeriosis in rabbits. The 4th Scientific Congress, Faculty of Veterinary Medicine, Assuit University, Egypt, pp.1045-1049.
- Ahmed, M.A.A., 2013. Pathological studies on some rabbit diseases causing nervous manifestations. M.V.Sc., Faculty of Veterinary Medicine, Zagazig University, Egypt.
- Amalaradjou, M.A.R., Bhunia, A.K., 2012. Modern approaches in probiotics research to control foodborne pathogens. Advances in Food and Nutrition Research 67, 185-239.
- Anton, W., 1934. Kritich experimental beitrag zur biologic des bacterium monocytogens. Zentralblatt für Bakteriologie, Mikrobiologie, und Hygiene A 131, 89-103.
- Arena, M.P., Silvain, A., Normanno, G., Grieco, F., Drider, D., Spano, G., Fiocco, D., 2016. Use of Lactobacillus plantarum strains as a bio-control strategy against food-borne pathogenic microorganisms. Frontiers in Microbiology 7, 464.
- Bajard, S., Rosso, L., Fardel, G., Flandrois, J.P., 1996. The particular behaviour of *Listeria monocytogenes* under sub-optimal conditions. International Journal of Food Microbiology 29, 201-211.
- Barbosa, J., Magalhaes, R., Santos, I., Ferreira, V., Brandao, T.R., Silva, J., Almeida, G., Teixeira, P., 2013. Evaluation of antibiotic resistance patterns of food and clinical *Listeria monocytogenes* isolates in Portugal. Foodborne Pathogens and Disease 10, 861-866.
- Barbuddhe, S.B., Chakraborty, T., 2009. Listeria as an enteroinvasive gastrointestinal pathogen. Current Topics in Microbiology and Immunology 337, 173-195.
- Belen Lopez, M., Briones, V., Fernandez-Garayzabal, J.F., Vazquez-Boland, J.A., Garcia, J.A., Blanco, M.M., Suarez, G., Dominguez, L., 1993. Serological response in rabbits to *Listeria monocytogenes* after oral or intragastric inoculation. FEMS Immunology and Medical Microbiology 7, 131-134.
- Boisivon, A., Guiomar, C., Carbon, C., 1990. *In vitro* bactericidal activity of amoxicillin, gentamicin, rifampicin, ciprofloxacin and trimethoprimsulfamethoxazole alone or in combination against *Listeria monocytogenes*. European Journal of Clinical Microbiology & Infectious Diseases 9, 206-209.
- Bortolussi, R., McGregor, D.D., Kongshavan, P.A.L., Galsworthy, S., Albirit-

ton, W., Davies, J.W., Seeliger, H.P.R., 1984. Host defense mechanisms for perinatal and neonatal *Listeria monocytogenes* infection. Survey and Synthesis of Pathology Research 3, 311-332.

- Carpentier, B., Cerf, O., 2011. Review-persistence of *Listeria monocytogenes* in food industry equipment and premises. International Journal of Food Microbiology 145, 1-8.
- Charpentier, E., Courvalin, P., 1999. Antibiotic resistance in *Listeria* spp. Antimicrobial Agents and Chemotherapy 43, 2103-2108.
- Cullere, M., Dalle Zotte, A., 2018. Rabbit meat production and consumption: State of knowledge and future perspectives. Meat Science 143, 137-146.
- Baker, D.G., 1998. Natural pathogens of laboratory mice, rats, and rabbits and their effects on research. Clinical Microbiology Reviews, 11, 231-266.
- De Cesare, A., Parisi, A., Mioni, R., Comin, D., Lucchi, A., Manfreda, G., 2017. Listeria monocytogenes circulating in rabbit meat products and slaughterhouses in Italy: Prevalence data and comparison among typing results. Foodborne Pathogens and Disease 14, 167-176.
- Dalle Zotte, A., Szendro, Z., 2011. The role of rabbit meat as functional food. Meat Science 88, 319-331.
- Dhama, K., Verma, A.K., Rajagunalan, S., Kumar, A., Tiwari, R., Chakraborty, S., Kumar, R., 2013. *Listeria monocytogenes* infection in poultry and its public health importance with special reference to food borne zoonoses. Pakistan Journal of Biological Science 16, 301-308.
- D'Orazio S.E., 2014. Animal models for oral transmission of *Listeria mono*cytogenes. Frontiers in Cellular and Infection Microbiology 11, 15.
- Drolia, R., Tenguria, S., Durkes, A.C., Turner, J.R., Bhunia, A.K., 2018. *Listeria* adhesion protein induces intestinal epithelial barrier dysfunction for bacterial translocation. Cell Host Microbe 23, 470-484.
- Drolia, R., Amalaradjou, M.A.R., Ryan, V., Tenguria, S., Liu, D., Bai, X., Xu, L., Singh, A.K., Cox, A.D., Bernal-Crespo, V., 2020. Receptor-targeted engineered probiotics mitigate lethal *Listeria* infection. Nature Communications 11, 6344.
- Ehsani, A., Rezaeiyan, A., Hashemi, M., Aminzare, M., Jannat, B., Afshari, A., 2019. Antibacterial activity and sensory properties of *Heracleum persicum* essential oil, nisin, and *Lactobacillus acidophilus* against *Listeria monocytogenes* in cheese. Veterinary World 12, 90-96.
- Fraser, J.A., Sperber, W.H., 1988. Rapid detection of *Listeria* species in food and environmental samples by esculin hydrolysis. Journal of Food Protection 51, 672-765.
- Gray, M.L., Killinger, A.H., 1966. *Listeria monocytogenes* and *Listeria* infections. Bacteriological Review 30, 309-382.
- Greenberg, S.S., Ouyang, J., Zhao, X., Giles, T.D., 1998. Human and rat neutrophils constitutively express neuronal nitric oxide syntheses mRNA. Nitric oxide Research 2, 203-212.
- Harty, J.T., Bevant, M.J., 1995. Specific immunity to *Listeria monocytogenes* in the absence of IFNγ. Immunity 3, 109-117.
- Hass, A., Kreft, J., 1988. *Listeria* biotechnological aspect of a pathogenic microorganism. International Industrial Biotechnology 8, 157-174.
- Hatab, M.E., Abd El-Latif, M.M., 2006. Studies on some bacterial associated with abortion in rabbits. Assuit Veterinary Medical Journal 52, 285-293.
- Hoelzer, K., Pouillot, R., Dennis, S., 2012. Animal models of listeriosis: a comparative review of the current state of the art and lessons learned. Veterinary Research 43, 1-27.
- Holt, J.G., Noel, R., Krieg, Peter, H.A., Sneath, James, T., Staley, T., 1994. Bergey's Manual of Determinative Bacteriology, Ninth Edition, 566-570.
- Ibrahim, G.A., brahim H.N., 2016. Bacteriological, clinic-pathological studies of *Listeria monocytogenes* in rabbits and detection of some virulence genes by polymerase chain reaction (PCR). Egyptian Journal of Chemistry and Environmental Health 2, 250-264.
- Janakiraman, V., 2008. Listeriosis in pregnancy: diagnosis, treatment, and prevention. Reviews in Obstetrics and Gynecology 1, 179-185.
- Johansson, J., Freitag, N.E., 2019. Regulation of *Listeria monocytogenes* virulence. Microbiology Spectrum 7, 1-20.
- Kabakci, N., Yarim, M., 2004. The expression of CD14 antigen in experimental encephalitic listeriosis in rabbits. Revue de Médecine Vétérinaire 155, 151-155.
- Kahn, M.A., Scott, L., Susan, E. Aiel-Io., Dana, G. Allen., David, P.A., Otto, M.R., Philip, T.R., Alice, M.W., 2006. The Merck Veterinary Manual, Ninth Edition. Whitehouse Station, NJ, USA.
- Khan, J.A., Rathore, R.S., Khan, S., Ahmad, I., 2013. *In vitro* detection of pathogenic *Listeria monocytogenes* from food sources by conventional, molecular and cell culture method. Brazilian Journal of Microbiology 44, 751-758.
- Koopmans, M.M., Brouwer, M.C., Geldhoff, M., Seron, M.V., Houben, J., van der Ende, A., van de Beek, D., 2014. Cerebrospinal fluid in-

flammatory markers in patients with *Listeria monocytogenes* meningitis. BBA Clinical 1, 44-51.

- Kruger, N., Low, C., Donachie, W., 1995. Phenotopic characterization of the cells of inflammatory response in ovine encephalitic listeriosis. Journal of Comparative Pathology 113, 263-275.
- Larsen, S.A., Feeley, J.C., Jones, W.L., 1974. Immune response to *Listeria monocytogenes* in rabbits and humans. Applied Microbiology 27, 1005-1013.
- Low, J.C., Donachie, W., 1991. Clinical and antibody responses of lambs to infection by *Listeria monocytogenes*. Research in Veterinary Science 51, 185-192.
- Lianou, A., Sofos, J.N., 2007. A review of the incidence and transmission of *Listeria monocytogenes* in ready-to-eat products in retail and food service environments. Journal of Food Protection 70, 2172-2198
- Malik, S.V., Barbuddhe, S.B., Chaudhari, S.P., 2002. *Listeria* infections in humans and animals in the Indian subcontinent: a review. Tropical Animal Health and Production 34, 359-381.
- Marco, A., domingo, M., Prts, N., Alimira, J., Briones, V., Dominguez, L., 1992. Differences in pathogenicity between *L. monocytogenes* and *L. ivanovii* after I/P and subcut inoculation. The 11th International Symposium on Problems of Listeriosis, Eigtveds. Copenhagen, 11-14 May.
- Meloni, D., Galluzzo, P., Mureddu, A., Piras, F., Griffiths, M., Mazzette, R., 2009. *Listeria monocytogenes* in RTE foods marketed in Italy: prevalence and automated EcoRI ribotyping of the isolates. International Journal of Food Microbiology 129, 166-173.
- Mengesha, D., Zewde, B.M., Toquin, M.T., Kleer, J., Hildebrandt, G., Gebreyes, W.A., 2009. Occurrence and distribution of *Listeria monocytogenes* and other *Listeria* species in ready-to-eat and raw meat products. Die Berliner und Münchener Tierärztliche Wochenschrift 122, 20-24.
- Moursi, M.K., Hala, M.E., Kawther, H.S., Fatma, M.Y., 2006. *Listeria* infection in rabbits at Ismailia province. South Canal Veterinary Medical Journal 1, 203-220.
- Munoz, A.I., 2012. Distribution of *Listeria monocytogenes* serotypes isolated from foods, Colombia, 2000-2009. Biomedica 32, 408-417.
- Murray, E.G.D., Webb, R.A., Swann, M.B.R., 1926. A disease of rabbits characterized by large mononuclear leucocytosis, caused by a hitherto undescribed bacillus Bacterium monocytogenes (n. sp.). Journal of Pathology and Bacteriology 29, 407-439.
- Ndahi, M.D., Kwaga, J.K., Bello, M., Kabir, J., Umoh, V.J., Yakubu, S.E., Nok, A.J., 2014. Prevalence and antimicrobial susceptibility of *Listeria monocytogenes* and methicillin resistant *Staphylococcus aureus* strains from raw meat and meat products in Zaria, Nigeria. Letters in Applied Microbiology 58, 262-269.
- Nakari, U.M., Rantala, L., Pihlajasaari, A., Toikkanen, S., Johansson, T., Hellsten, C., Raulo, S.M., Kuusi, M., Siitonen, A., Rimhanen Finne, R., 2014. Investigation of increased listeriosis revealed two fishery production plants with persistent *Listeria* contamination in Finland in 2010. Epidemiology and Infection 24, 1-9.
- Najdenski, H., Vesselinova, A., 2002. Experimental mixed infection of rabbits with Yarsenia enterocolitica and *Listeria monocytogenes*. Journal of Veterinary Medicine Series B: Infectious Diseases and Veterinary Public Health 49, 97-104.
- OIE, 2014. *Listeria monocytogenes*. Chapter 2.9.7. Manual of Diagnostic Tests and Vaccines for Terrestrial Animals. p. 1-18. Available from: http://www.oie.int/manual-of-diagnostic-tests-and-vaccines-for-terrestrial-animals/.
- Okada, Y., Okutani, A., Suzuki, H., Asakura, H., Monden, S., Nakama, A., Maruyama, T., Igimi, S., 2011. Antimicrobial susceptibilities of *Listeria monocytogenes* isolated in Japan. Journal of Veterinary Medical Science 73, 1681-1684.
- Okerman, L., 1999. Diseases of domestic rabbits. Library of Veterinary Practice, second edition-Blackwell science Ltd. U.K.
- Orndorff, P.E., Hamrick, T.S., Smoak, I.W., Havell, E.A., 2006. Host and bacterial factors in listeriosis pathogenesis. Veterinary Microbiology 114, 1-15.
- Pamukcu, T., Yarim, G.F., Kaba-kci, N., Yarim, M., Duru, O., 2004. Experimental listeriosis in rabbits: Biochemical changes in serum and cerebrospinal fluid. Revue de Médecine Vétérinaire 156, 253-258. Available at: https://www.semanticscholar.org/paper/Experimental-listeriosis-in-rabbits%3A-Biochemical-in-Pamuk%C3%A7u-Yarim/63291ece6fb0827f7372f98b5161fe05f3d2150e
- Patton, N.M., Hagen, K.W., Gorham, J.R., Flatt, R.E., 2000. Domestic Rabbits: Diseases and Parasites. A Pacific Northwest Extension Publication Oregon-Idaho-Washington.
- Poulsen, K.P., Czuprynski, C.J., 2013. Pathogenesis of listeriosis during pregnancy. Animal Health Research Review 14, 30-39.
- Popovic, I., Heron, B., Covacin, C., 2014. Listeria: an Australian perspective

(20012010). Foodborne Pathogens and Disease 11, 425-443

- Quinn, P.J., Markery, B.K., Carter, M.E., Donnelly, W.J., Leonard, F.C., 2002. Veterinary Microbiology and Microbial Diseases. Blockwell Science Ltd. 1st Published. Radoshevich, L., Cossart, P., 2018. *Listeria monocytogenes*: Towards a complete picture of its physiology and pathogenesis. Nature Reviews Microbiology 16, 32-46.
- Radostits, O.M., Gay, C.C., Hinchcliff, K.W., Constable, P.D., 2008. Veterinary Medicine. A textbook of the disease of cattle, horses, sheep, pigs and goats. 10th ed. Philadelphia (PA): Saunders.
- Rahimi, E., Yazdi, F., Farzinezhadizadeh, H., 2012. Prevalence and antimicrobial resistance of *Listeria* species isolated from different types of raw meat in Iran. Journal of Food Protection 75, 2223-2227.
- Regan, T., MacSharry, J., Brint, E., 2014. Tracing innate immune defences along the path of *Listeria monocytogenes* infection. Immunology & Cell Biology 92, 563-569.
- Rivero, G.A., Torres, H.A., Rolston, K.V.I., 2003. Listeria monocytogenes infection in patients with cancer. Diagnostic Microbiology and Infectious Disease 47, 393-398.
- Rocha, K.R., Perini, H.F., de Souza, C.M., Schueler, J., Tosoni, N.F., Furlaneto, M.C., Furlaneto-Maia, L., 2019. Inhibitory effect of bacteriocins from enterococci on developing and preformed biofilms of *Listeria monocytogenes*, *Listeria ivanovii* and *Listeria innocua*. World Journal of Microbiology and Biotechnology 35, 96.
- Sagun, E., Durmaz, H., Tarakci, Z., Sagdic, O., 2006. Antibacterial activities of the extracts of some herbs used in Turkish herby cheese against *Listeria monocytogenes* serovars. International Journal of Food Properties 9, 255-260.
- Schlech, W.F., Lavigne, P.M., Bortolussi, R.A., Allen, A.C., Haldane, E.V., Wort, A.J., Hightower, A.W., Johnson, S.E., King, S.H., Nichols, E.S., Broome, C.V., 1983. Epidemic listeriosis- Evidence for transmission by food. New England Journal of Medicine 308, 203-206.
- Seeliger, H.P.R., 1987. Listeriosis: Diagnosis and treatment of listeriosis. In: Schonberg, A., Ed., Joint WHO-ROI Consultation on Prevention and Control of Listeriosis. Veterinary Medicine Hefte 5, 63-75.
- Seeliger, H.P.R., Jones, D., 1986. Genus Listeria pirie. In Sneath, P.H.A., Mair, N.S., Sharpe, M.E., Holt, J.G. (eds.). Bergeys Manual of Systemic Bacteriology, Vol. 2, the Williams and Wilkins Company, Baltimore, pp. 1235-1245.
- Shin, T., Weinstock, D., Castro, M.D., Acland, H., Walter, M., Kim, H.Y., Purchase, H.G., 2000. Immunohistochemical study of constitutive neuronal and inducible nitric oxide synthase in the central nervous system of goat with natural listeriosis. Journal of Veterinary Science 1, 77-80.
- Skariyachan, S., Pachiappan, A., Joy, J., Bhaduri, R., Aier, I., Vasist, K.S., 2015. Investigating the therapeutic potential of herbal leads against drug resistant *Listeria monocytogenes* by computational virtual screening and *In vitro* assays. Journal of Biomolecular Structure and Dynamics 33, 2682-2694.
- Soni, D.K., Singh, R.K., Singh, D.V., Dubey, S.K., 2013. Characterization of *Listeria monocytogenes* isolated from Ganges water, human clinical and milk samples at Varanasi, India. Infection, Genetics and Evolution 14, 83-91.
- Southwick, F.S., Purich, D.L., 1996. Mechanisms of disease Intracellular pathogenesis of listeriosis. New England Journal of Medicine 334, 770-776.
- Stratakos, A.Ch., Ijaz, U.Z., Ward, P., Linton, M., Kelly, C., Pinkerton, L., Scates, P., McBride, J., Pet, I., Criste, A., Stef, D., Couto, J.M., Sloan, W.T., Dorrell, N., Wren, B.W., Stef, L., Gundogdu, O., Corcionivoschi, N., 2020. *In vitro* and *in vivo* characterisation of *Listeria monocytogenes* outbreak isolates. Food Control 107, 106784.
- Suez, J., Zmora, N., Segal, E., Elinav, E., 2019. The pros, cons, and many unknowns of probiotics. Nature Medicine 25, 716-729.

- Taha, M., Abd El-Motelib, T.Y., El-Ballal, S.S., El-Zanaty, K., 1991. Experimental studies on a new *Listeria monocytogenes* isolate in rabbits. Egyptian Journal of Comparative Pathology and Clinical Pathology 4, 301-312.
- Thongson, C., Davidson, P.M., Mahakarnchanakul, W., Vibulsresth, P., 2005. Antimicrobial effect of Thai spices against *Listeria monocytogenes* and *Salmonella typhimurium* DT104. Journal of Food Protection 68, 2054-2058.
- Trabelsi, I., Slima, S.B., Ktari, N., Triki, M., Abdehedi, R., Abaza, W., Moussa, H., Abdeslam, A., Salah, R.B., 2019. Incorporation of probiotic strain in raw minced beef meat: study of textural modification, lipid and protein oxidation and color parameters during refrigerated storage. Meat Science 154, 29-36.
- Tiwari, R., Chakraborty, S., Dhama, K., Rajagunalan, S., Singh, S.V., 2013. Antibiotic resistance- an emerging health problem: causes, worries, challenges and solutions a review. International Journal of Current Research 5, 1880-1892.
- Vandera, E., Lianou, A., Kakouri, A., Feng, J., Koukkou, A.I., Samelis, J., 2017. Enhanced control of *Listeria monocytogenes* by *Enterococcus fae-cium* KE82, a multiple enterocin-producing strain, in different milk environments. Journal of Food Protection 80, 74-85.
- Van Zyl, W.F., Deane, S.M., Dicks, L.M.T., 2016. Enterococcus mundtii ST-4SA and Lactobacillus plantarum 423 excludes *Listeria monocytogenes* from the GIT, as shown by bioluminescent studies in mice. Beneficial Microbes 7, 227-235.
- Vazquez-Boland JA, Kuhn M, Berche P, Chakraborty T, Domínguez-Bernal G, Goebel W, González-Zorn B, Wehland J, Kreft J., 2001. *Listeria* pathogenesis and molecular virulence determinants. Clinical Microbiology Reviews 14, 584-640.
- Volokhovel, D.V., Duperrier, S., Neverov, A.A., George, J., Buchrieser, C., Hitchins, A.D., 2007. The presence of the internalin gene in natural atypically hemolytic *Listeria innocua* strains suggests descent from *L. monocytogenes*. Applied and Environmental Microbiology 73, 1928-1939.
- Wang, J., Liu, N., Zhang, F., 2019. Tetramethylpyrazine protects oxidative stability and gelation property of rabbit myofibrillar proteins. Food Science of Animal Resources 39, 623-631.
- Warbureton, D., Anne, B., Eliane Pagottof, D., Cindy, C., 2003. The detection of *Listeria* spp. In foods and environmental samples using pal cam broth. Health products and food Branch (HPFB), Ottawa, Ontario.
- Watson, G., Evans, M., 1985. Listeriosis in a rabbit. Veterinary Pathology 22, 191-193.
- Wesley, I.V., 2007. Listeriosis in animals. In: Ryser ET, Marth EH, editors. Listeria, listeriosis and food safety. 3rd ed. Boca Raton (FL): CRC Press; p. 55-84.
- Witkowska, A.M., Hickey, D.K., Alonso-Gomez, M., Wilkinson, M., 2013. Evaluation of antimicrobial activities of commercial herb and spice extracts against selected food-borne bacteria. Journal of Food Research 2, 37-54.
- Wuenscher, M.D., Kohler, S., Bubert, A., Gerike, U., Goebel, W., 1993. The iap gene of *Listeria monocytogenes* is essential for cell viability, and its gene product, p60, has bacteriolytic activity. Journal of Bacteriology 175, 3491-3501.
- Yoon, Y., Choi, K.H., 2012. Antimicrobial activities of therapeutic herbal plants against *Listeria monocytogenes* and the herbal plant cytotoxicity on Caco-2 cell. Letters in Applied Microbiology 55, 4755.
- Zhao, H., Zhang, F., Chai, J., Wang, J., 2020. Effect of lactic acid bacteria on *Listeria monocytogenes* infection and innate immunity in rabbits. Czech Journal of Animal Science 65, 23-30.