

Review Article

Bacillus cereus Characteristics, Virulence Factors Profiles and Toxin Production

Aya R. Mohammed, Esmat I. El-Said, Salah F. Abd ElAal, Rania M. Kamal*

Department of Food Control, Zagazig University, Zagazig City, 44511, Sharkia Governorate, Egypt.

***Correspondence**Corresponding author: Rania M. Kamal
E-mail address: raniakamal79@gmail.com**Abstract**

Bacillus cereus is a Gram positive, facultative anaerobic bacterium characterized by large rod-shaped cells and an ability to form heat-resistant endospores. Because this bacterium is so widely distributed throughout nature and is frequently found in soil, it is naturally present in a wide variety of food products of both animal and plant origin. The presence of *B. cereus* and its virulence factors in dairy products may result in food poisoning and other illnesses. *B. cereus* causes two types of intestinal illness: emetic (vomiting) and diarrheal. Furthermore, the bacterium causes a variety of systemic and local infections in both immunosuppressed and immunocompetent persons. Different toxins and pathogenic factors like nonhemolytic enterotoxin Nhe, hemolytic enterotoxin Hbl, enterotoxin FM and cytotoxin K are accountable for diarrheal syndrome, meanwhile the depsipeptide cereulide toxin causes emetic syndrome. Because of its genetic similarity to *Bacillus anthracis* and *Bacillus thuringiensis*, the ability to detect this pathogen in food is frequently difficult. We reviewed characters, virulent attributes, toxins profiles of this bacterium. Current control methods are limited so future control strategies must be developed.

KEYWORDS*B. cereus*, Cereulide, Emetic syndrome, Diarrheal toxins.**INTRODUCTION**

One of the most challenging public concerns is food safety. Food pathogens and their toxins are responsible for a variety of food-related health threats that cause disease and death worldwide (Assefa, 2019). *Bacillus cereus* (*B. cereus*) is an example of these foodborne pathogens (Soleimani *et al.*, 2018). This species is commonly known for food pathogenicity in the *Bacillus* genus (Soleimani *et al.*, 2018).

Bacillus cereus is a Gram-positive, rod shape, motile, spore-forming bacterium that related to the genus *Bacillus* (Montville and Matthews, 2005). Under adverse environmental conditions, the microorganism produces oval endospores that can survive for extended periods of time, even in extreme conditions (Delbrassinne *et al.*, 2012). Because of the bacterial ability to grow at temperatures ranging from 4°C to 50°C and resist heat and chemicals, this pathogen is popularly found in soil, air, grains, rice (row and cooked), vegetables, meat, and milk (Setlow, 2006).

Taxonomy and characteristics of *B. cereus*

The *B. cereus* group species complex, also known as *B. cereus sensu lato* (s.l.), is a subgroup of strongly related *Bacillus* species. Members of this group are Gram-positive, spore-forming, and widely spread throughout the environment (Carroll *et al.*, 2020). This *B. cereus* group includes at least twelve species that are closely related: *B. cereus*, *B. thuringiensis*, *B. anthracis*, *B. mycoides*, *B. pseudomycoides*, *B. cytotoxicus*, *B. weihenstephan-*

ensis, *B. wiedmanni*, *B. toyonensis*, and the recently identified, *B. albus*, *B. tropicus*, *B. paranthracis*, *B. pacificus*, *B. mobilis*, *B. Luti*, *B. proteolyticus*, *B. nitrateducentis*, *B. paramycoides*, *B. gaemokensis*, *B. bingmayongensis*, *B. manliponensis* and *B. fungorum* (Bianco *et al.*, 2021).

Bacillus cereus strains can grow in a moderately wide temperature range; most lineages seem to be mesophilic (with an optimal growth temperature of 37°C and survival below 10°C), while psychotropic strains (with an optimal growth temperature below 10°C and slow growth at 37°C) have also been described, this allows them to be easily isolated from foods even when refrigerated. Some lineages are linked to milk and dairy production environments, including psychrotolerant strains (Guinebretiere *et al.*, 2008).

Ecology of *B. cereus*

Because *B. cereus* is so ubiquitous, its presence in most raw foods is considered inescapable. The original source of *B. cereus* spore contamination in food is soil. Similar genotypic strains, for example, have been found in milk and dairy farm soil (Christiansson *et al.*, 1999). The soil is *B. cereus*' principal environmental reservoir, where it can finish its saprophytic life cycle (Vilain *et al.*, 2006). When conditions are favourable, *Bacillus cereus* spores germinate in soil, grow on decomposing organic matter in the multicellular filamentous phenotype for translocation through the soil, and eventually resporulate when nutrients are diminished, and conditions are no longer favourable.

Contamination routes

Bacillus cereus can easily contaminate a variety of foods and it is substantially impossible to obtain raw materials free from *B. cereus* spores (Johnson, 1984; Te Giffel et al., 1996). concerning food products, this microorganism has been obtained from the air in a cow shed (Ehling-Schulz et al., 2019), rice, spices, milk and dairy products, vegetables, meat, starchy foods, desserts, and cakes (Ghelardi et al., 2002). This organism is a significant issue for the food industry because it is difficult to be removed from food products due to its ability to tolerate a wide range of circumstances despite environmental stresses that would normally restrict bacterial survival (Andersson et al., 1995). The milk production system is highly complicated, with dead ends, pockets, and traps that have difficulty in cleaning process and can serve as a constant reservoir for spoilage microorganisms (e.g., valves, shafts, and gaskets). Milking equipment can also be a source of contamination because spores can attach and germinate in dairy equipment (for example, pasteurizers, tanks, and packaging machines), resulting in post-treatment contamination of milk (Te Giffel et al., 1996; Svensson et al., 2000; Eneroth et al., 2001). Milk contamination by *Bacillus cereus* group members is critical not only because of their ability to spoil milk, but also because of their potency to cause diseases in humans. (Andersen Borge et al., 2001; Janštová et al., 2006).

Bacillus cereus Culture and Isolation

The agar plate-based counting is the conventional method for *B. cereus* detection, which is guided by ISO 7932:2004, and was last revised nearly 15 years ago. For each dilution, the plating steps must be duplicated using certain media. Then the plates must be incubated at 30°C for 24 to 48 hours. The selective media mannitol yolk poly-myxin B agar (MYP) and polymyxin pyruvate egg yolk mannitol bromothymol blue agar (PEMBA) is frequently used for its selective isolation and count from food products. *B. cereus* grows fast on these media (within just 24 hours) forming colonies surrounded by an opaque halo caused due to the ability of the microorganism's lecithinases to degrade the lecithin present in the egg yolk. On PEMBA medium, Colonies are crenated, 5mm diameter, turquoise to peacock blue with a zone of egg yolk precipitation after 18-24 h incubation. *B. cereus* colonies on MYP medium have a pink-purple color, surrounded by a distinctive halo formed of pink precipitation, allowing their identification. Moreover, when assessing *B. cereus* counts, the presence of vegetative cells, spores, or a mixture of both forms should be considered. A thermal treatment (80°C for 10 minutes) is performed to selectively detect spores (Reyes et al., 2007; Mohamed et al., 2016; Spanu et al., 2016) This step produces a dual effect, inactivating vegetative cells and activating spore germination.

Recently, alternative approaches for enumerating presumptive *B. cereus* in food based on the NF EN ISO 7932 standard have been confirmed. They use a selective chromogenic medium, COMPASS and BACARA respectively. *B. cereus* colonies appear in green (COMPASS) or orange surrounded by an opaque halo (BACARA) after hydrolysis of the chromogenic substrate. They have comparable specificity, selectivity, and accuracy to the standard reference. Furthermore, they save time and do not require a confirmation action.

Toxins and Virulence factors

Consuming food contaminated by *B. cereus* may lead to gastrointestinal diseases and non- gastrointestinal diseases in-

cluding respiratory tract infections, bacteremia, endocarditis and central nervous system infections (Fricker et al., 2007).The diarrheal disease characterized by abdominal pain and non-bloody diarrhea that occur 4–16 h after eating. These symptoms are caused by the bacterial production of diarrheal toxins hemolysin BL (Hbl), nonhemolytic enterotoxin (Nhe) (Ehling-Schulz et al., 2006b) and cytotoxin CytK (Fagerlund et al., 2004) in the human small intestine. All three enterotoxins are cytotoxic and cell membrane pore-forming toxins (Ramarao and Sanchis, 2013; Kumari and Sarkar, 2016). Hbl and Nhe each consist of three different protein components, named L2, L1, and B, and NheA, NheB and NheC, respectively, while CytK is a single-component toxin (Stenfor Arnesen et al. 2008; Fagerlund et al. 2010). Emetic disease, caused by a thermo- and acidic-stable non-ribosomal peptide called cereulide (emetic toxin), and can occur within 0.5-5 hours of ingesting food that has been contaminated then the symptoms appear as an acute attack of nausea, vomiting, and copious abdominal cramping (Ehling-Schulz et al., 2005; 2015).

Bacillus cereus not only produces toxins but also releases several compounds (cytotoxic factors, degradation enzymes, proteases, hemolysins, cell-surface proteins) that may play a role in its virulence (Ramarao and Sanchis, 2013; Guillemet et al., 2016). For example, InhA1 and NprA are synthesized metalloproteases that disturb the body immune defense system and contribute to *B. cereus* pathogenicity (Charlton et al., 1999; Ramarao and Lereclus, 2005; Guillemet et al., 2010; Haydar et al., 2018). Enterotoxin T (BcET) and enterotoxin FM (EntFM) are other enterotoxins that believed to play a role in foodborne illness. The pore-forming toxin Hemolysin II (HlyII) stimulates macrophage apoptosis, enabling *B. cereus* to evade immune defense of the host (Tran et al., 2010; 2011). Further virulence factors including sphingomyelinase and exoproteases have been reported to support the pathogenicity of *B. cereus* (Doll et al., 2013; Jeßberger et al., 2015).

Cereulide

Melling et al. made the initial suggestion in 1976 that emetic syndrome is caused by a different toxin. They suggested this by feeding Rhesus monkeys two distinct isolated strains, one from a diarrheal outbreak and the other from a vomiting outbreak. The first strain resulted in fluid retention, whereas the second did not. This indicated that vomiting symptoms are caused by a distinctive toxin (Melling et al., 1976). Then, (Agata et al., 1995) identified cereulide as the emetic toxin factor.

Cereulide is a toxic cyclic dodecadepsipeptide (1.2 kDa) produced enzymatically by a large protein complex in a process called non ribosomal protein synthesis (NRPS) (Horwood et al., 2004).The cyclic and lipophilic dodecadepsipeptide containing three repetitions of four amino acids, D-Oxy- Leu—DAla—L-Oxy-Val—L-Val, which resembles valinomycin, a well-known antibiotic formed by *Streptomyces* with the sequence D-Oxy-Hyi—D-Val—L-Oxy-Lac—L-Val (Agata et al., 1994). The ces gene locus, which is located on a 270-kb megaplasmid known as pCER270 (or pCERE01) has coding for the enzymes concerned with cereulide synthesis. This plasmid is significantly linked to the toxin plasmid pX01 from *Bacillus anthracis* (Ehling-Schulz, et al., 2006a). CesA (10 kb) and cesB (8 kb) genes encode the cereulide synthetase. CesA is mainly accountable for the d-Ala-d-O-Leu fragment, while the CesB is necessary for the generation of l-Val-l-O-Val fragment. (Marxen et al., 2015).

Cereulide, which is produced by emetic *B. cereus* strains, can be found in foodstuffs, creating a significant risk of food poisoning. Furthermore, the toxin has been shown to be heat, low pH, and trypsin digestion resistant, allowing passage through

the stomach into the digestive tract. The fact that cereulide can withstand heat treatment at 12°C for 2 hours, roasting, frying, microwave cooking, and pH values ranging from 2 to 10 demonstrates why traditional food processing methods fail to eliminate its presence in consumer foods (Agata et al., 2002; Rajkovic et al., 2006).

Cereulide formation in the digestive tract is rarely observed, if it occurs at all. Intoxication is usually caused by the ingestion of toxin preformed in food by ces-positive *B. cereus* strains prior to consumption (Ehling-Schulz et al., 2015).

Enterotoxins

Diarrhea is caused by at least five *B. cereus* enterotoxins. Two of them are protein complexes (hemolysin BL and non-hemolytic enterotoxins). The other three (enterotoxin FM, enterotoxin T, and cytotoxin K) are single protein substances (Beecher et al., 1995; Lund et al., 2000). Diarrhea occurs after enterotoxins are produced in the intestine where invasion and outgrowth of cells and/or most likely spores occur after ingesting contaminated foods (Ceuppens et al., 2012).

Hemolysin BL

Hbl is an enterotoxin developed by diarrheal *B. cereus* strains. It is a three-partite protein consisting of a binding protein B encoded by the hblA gene and two lytic components L1 and L2 encoded by the hblC and hblD genes, respectively. The first enterotoxin recognized in *B. cereus* was Hbl (Beecher and Wong, 1994). All three components are required for Hbl's biological activity, additionally, Hbl must be combined with Nhe to be biologically active (Sastalla et al., 2013). The Hbl potential was comparable to that of cholera toxin, thus supposing that it could be regarded as a principal virulence factor in diarrheal cases caused by *B. cereus* (Beecher et al., 1995).

Non-hemolytic enterotoxin (Nhe)

The three-component enterotoxin Nhe was discovered for the first time in a *B. cereus* strain that was associated with a large food poisoning outbreak in Norway (Lund and Granum, 1996). This three-component pore-forming toxin is made up of the proteins NheA (41.0 kDa), NheB (39.8 kDa), and NheC (36.5 kDa), which work together to form a pore in the cell membrane and induce cell lysis (Lindback et al., 2004). The proteins were distinguishable from Hbl components and exhibited no hemolytic activity. About 92 to 100% of *B. cereus* isolates produce Nhe toxin. In comparison, only 42 strains were able to synthesize the enterotoxin Hbl, revealing the predominance of Nhe in *B. cereus* strains, which is found more frequently than Hbl production (Schoeni and Wong, 2005; Moravek et al., 2006). This demonstrates that Nhe serves as the most predominating toxin in diarrheal food poisoning, proving that *B. cereus* cytotoxicity is strongly related to the concentration of Nhe and weakly related to Hbl concentration (Fagerlund et al., 2008).

Cytotoxin K

CytK was the first toxin proved to be the only cytotoxic, necrotic, and hemolytic toxin of the *B. cereus* strain NVH391/98, which was implicated in a serious food poisoning outbreak in France in 1998. When six people suffered from bloody diarrhea, and three died, during this time, none of the known *B. cereus* enterotoxins could be discovered, and CytK has been shown to be

toxic towards Vero cells (Lund et al., 2000). This 34-kDa protein was also isolated from fractionated supernatants of an endophthalmitis-associated *B. cereus* isolate in the same year and termed "hemolysin IV". (Beecher et al., 2000).

CytK is a single protein, β -barrel pore-forming toxin. There are two variants of CytK that share 89% of identity: CytK-1, and CytK2. CytK1 was identified in 1998 from a strain of *B. cereus* that caused a large foodborne outbreak (FBO) in France and that conducted to the death of three elderly people (Lund et al., 2000). Although CytK2 is five times less toxic than CytK-1, it seems more frequently associated with strains causing FBO (Ramarao and Sanchis, 2013).

Applicable Control Strategies of *B. cereus*

Recent research has shown that high hydrostatic pressures (HHP) could be effective in inactivating bacterial spores when applied in combination with moderate temperature heat treatments (>60°C) (Soni et al., 2016). According to (Schneider et al., 2017), and based on the National Institutes of Health (NIH), the National Food Processors Association (NFPA), and the FDA Food Code 2013, the following strategies are helpful for destroying *B. cereus*. Both vegetative cells and spores can be destructed by steaming food at $\geq 145^\circ\text{F}$ (63°C) under pressure, roasting, frying, and grilling. Cooking at 145°F (63°C) and reheating at 165°F (74°C) for 15 seconds kills the vegetative cells. However, if the toxin was produced in food, it is hazardous to consume. Therefore, the best way to prevent spore germination is to cool foods quickly after heating them.

CONCLUSION

This organism is the primary cause of the majority of food-related diseases because of the high prevalence of *B. cereus* food contamination and active production and secretion of HBL, Nhe, and CytK enterotoxins. *B. cereus*' ability to form highly resistant spores, permitting it to spread in a wide range of environments, is what makes it a successful food pathogen. As a result, *B. cereus* presents in a broad range of foodstuffs, posing a serious food safety risk. Current FDA and other regulatory agencies control practices are inadequate for removing *B. cereus* from food. New techniques for controlling or eliminating *B. cereus* from food matrices are needed.

CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

REFERENCES

- Agata, N., Mori, M., Ohta, M., Suwan, S., Ohtani, I., Isobe, M., 1994. A novel dodecadepsipeptide, cereulide, isolated from *Bacillus cereus* causes vacuole formation in HEp-2 cells. FEMS Microbiol. Lett. 121, 31–34.
- Agata, N., Ohta, M., Mori, M., Isobe, M., 1995. A novel dodecadepsipeptide, cereulide, is an emetic toxin of *Bacillus cereus*. FEMS Microbiol. Lett. 129, 17–19.
- Agata, N., Ohta, M., Yokoyama, K., 2002. Production of *Bacillus cereus* emetic toxin (cereulide) in various foods. Int. J. Food Microbiol. 73, 23–27.
- Andersen Borge, G.I., Skeie, M., Sorhaug, T., Langsrud, T., Granum, P.E., 2001. Growth and toxin profiles of *Bacillus cereus* isolated from different food sources. Int. J. Food Microbiol. 69, 237–246.
- Andersson, A., Ronner, U., Granum, P.E., 1995. What problems does the food industry have with the spore-forming pathogens *Bacillus cereus* and *Clostridium perfringens*?. Int. J. Food Microbiol. 28, 145–155.
- Assefa, A., 2019. Prevalence of *Escherichia coli* O157: H7 in foods of animal origin in Ethiopia: A meta-analysis. Cogent. Food Agric. 5,

- 1642981.
- Beecher, D.J., Olsen, T.W., Somers, E.B., Wong, A.C., 2000. Evidence for contribution of tripartite hemolysin BL, phosphatidylcholine-preferring phospholipase C, and collagenase to virulence of *Bacillus cereus* endophthalmitis. *Infect. Immun.* 68, 5269–5276.
- Beecher, D.J., Schoeni, J.L., Wong, A.C., 1995. Enterotoxic activity of haemolysin BL from *Bacillus cereus*. *Infect. Immun.* 63, 4423–4428.
- Beecher, D.J., Wong, A., 1994. Identification of hemolysin BL-producing *Bacillus cereus* isolates by a discontinuous hemolytic pattern in blood agar. *Appl. Environ. Microbiol.* 60, 1646–1651.
- Bianco, A., Capozzi, L., Monno, M.R., Del Sambro, L., Manzulli, V., Pesole, G., Parisi, A., 2021. Characterization of *Bacillus cereus* group isolates from human bacteremia by whole-genome sequencing. *Front. Microbiol.* 11, 599524.
- Carroll, L.M., Wiedmann, M., Kovac, J., 2020. Proposal of a taxonomic nomenclature for the *Bacillus cereus* group which reconciles genomic definitions of bacterial species with clinical and industrial phenotypes. *MBio.* 11, e00034–20
- Ceuppens, S., Uyttendaele, M., Drieskens, K., Heyndrickx, M., Rajkovic, A., Boon, N., Van de Wiele, T., 2012. Survival and germination of *Bacillus cereus* spores without outgrowth or enterotoxin production during in vitro simulation of gastrointestinal transit. *Appl. Environ. Microbiol.* 78, 7698–7705.
- Charlton, S., Moir, A., Baillie, L., Moir, A., 1999. Characterization of the exo-sporium of *Bacillus cereus*. *J. Appl. Microbiol.* 87, 241–245.
- Christiansson, A., Bertilsson, J., Svensson, B., 1999. *Bacillus cereus* spores in raw milk: factors affecting the contamination of milk during the grazing period. *J. Dairy Sci.* 82, 305–314.
- Delbrassinne, L., Andjelkovic, M., Dierick, K., Denayer, S., Mahillon, J., Van Looc, J., 2012. Prevalence and levels of *Bacillus cereus* emetic toxin in rice dishes randomly collected from restaurants and comparison with the levels measured in a recent foodborne outbreak. *Foodborne Pathog. Dis.* 9, 809–814.
- Doll, V.M., Ehling-Schulz, M., Vogelman, R., 2013. Concerted action of sphingomyelinase and non-hemolytic enterotoxin in pathogenic *Bacillus cereus*. *PLoS One.* 8, e61404. <https://doi.org/10.1371/journal.pone.0061404>.
- Ehling-Schulz, M., Frenzel, E., Gohar, M., 2015. Food-bacteria interplay: pathometabolism of emetic *Bacillus cereus*. *Front. Microbiol.* 6, 704
- Ehling-Schulz, M., Fricker, M., Grallert, H., Rieck, P., Wagner, M., Scherer, S., 2006a. Cereulide synthetase gene cluster from emetic *Bacillus cereus*: Structure and location on a mega virulence plasmid related to *Bacillus anthracis* toxin plasmid pXO1. *BMC Microbiol.* 6, 20.
- Ehling-Schulz, M., Guinebretiere, M. H., Monthán, A., Berge, O., Fricker, M., Svensson, B., 2006b. Toxin gene profiling of enterotoxic and emetic *Bacillus cereus*. *FEMS Microbiol. Lett.* 260, 232–240.
- Ehling-Schulz, M., Lereclus, D., Koehler, T.M., 2019. The *Bacillus cereus* group: *Bacillus* species with pathogenic potential. *Microbiol. Spectr.* 7, doi:10.1128/microbiolspec.GPP3-0032-2018.
- Ehling-Schulz, M., Vukov, N., Schulz, A., Shaheen, R., Andersson, M., Märtilbauer, E., Scherer, S., 2005. Identification and partial characterization of the nonribosomal peptide synthetase gene responsible for cereulide production in emetic *Bacillus cereus*. *Appl. Environ. Microbiol.* 71, 105–113.
- Eneroth, S. A., Svensson, B., Molin, G., Christiansson, A., 2001. Contamination of pasteurised milk by *Bacillus cereus* in the filling machine. *J. Dairy Res.* 68, 189–196.
- Fagerlund, A., Lindback, T., Granum, P.E., 2010. *Bacillus cereus* cytotoxins Hbl, Nhe and CytK are secreted via the Sec translocation pathway. *BMC Microbiol.* 10, 1–8.
- Fagerlund, A., Lindbäck, T., Storset, A.K., Granum, P. E., Hardy, S. P., 2008. *Bacillus cereus* Nhe is a pore-forming toxin with structural and functional properties similar to the ClyA (HlyE, SheA) family of haemolysins, able to induce osmotic lysis in epithelia. *Microbiol.* 154, 693–704.
- Fagerlund, A., Ween, O., Lund, T., Hardy, S.P., Granum, P.E., 2004. Genetic and functional analysis of the cytK family of genes in *Bacillus cereus*. *Microbiol.* 150, 2689–2697.
- Fricker, M., Messelhäußer, U., Busch, U., Scherer, S., Ehling-Schulz, M., 2007. Diagnostic real-time PCR assays for the detection of emetic *Bacillus cereus* strains in foods and recent food-borne outbreaks. *Appl. Environ. Microbiol.* 73, 1892–1898.
- Ghelardi, E., Celandroni, F., Salvetti, S., Barsotti, C., Baggiani, A., Senesi, S., 2002. Identification and characterization of toxigenic *Bacillus cereus* isolates responsible for two food-poisoning outbreaks. *FEMS Microbiol. Lett.* 208, 129–134.
- Guillemet, E., Cadot, C., Tran, S.L., Guinebretiere, M.H., Lereclus, D., Ramaraao, N., 2010. The InhA metalloproteases of *Bacillus cereus* contribute concomitantly to virulence. *J. Bacteriol.* 192, 286–294.
- Guillemet, E., Lérééc, A., Royer, C., Tran, S., Barbosa, I., Sansonetti, P., Lereclus, D., Ramaraao, N., 2016. The bacterial repair protein Mfd confers resistance to the host nitric-oxide response. *Sci. Rep.* 6, 29349.
- Guinebretiere, M. H., Thompson, F. L., Sorokin, A., Normand, P., Dawyndt, P., Ehling-Schulz, M., Svensson, B., Sanchis, V., Nguyen-The, C., Heyndrickx, M., De Vos, P., 2008. Ecological diversification in the *Bacillus cereus* Group. *Environ. Microbiol.* 10, 851–865.
- Haydar, A., Tran, S. L., Guillemet, E., Darrigo, C., Perchat, S., Lereclus, D., Coquet, L., Jouenne, T., Ramaraao, N., 2018. InhA1-mediated cleavage of the metalloprotease NprA allows *Bacillus cereus* to escape from macrophages. *Front. Microbiol.* 9, 1063.
- Horwood, P.F., Burgess, G.W., Jane Oakey, H., 2004. Evidence for non-ribosomal peptide synthetase production of cereulide (the emetic toxin) in *Bacillus cereus*. *FEMS Microbiol. Lett.* 236, 319–324.
- ISO (International Organization for Standardization), 2006. ISO 7932: 2004. Microbiology of food and animal feeding stuffs. Horizontal method for the enumeration of presumptive *Bacillus cereus*. Colony-count technique at 30°C. ISO, Geneva.
- Janstova, B., Drackova, M., Vorlova, L., 2006. Effect of *Bacillus cereus* enzymes on the milk quality following ultra-high temperature processing. *Acta Vet. Brno.* 75, 601–609.
- Jeßberger, N., Krey, V.M., Rademacher, C., Böhm, M.E., Mohr, A.K., Ehling-Schulz, M., Scherer, S., Märtilbauer, E., 2015. From genome to toxicity: a combinatory approach highlights the complexity of enterotoxin production in *Bacillus cereus*. *Front. Microbiol.* 6, 1–15. <https://doi.org/10.3389/fmicb.2015.00560>
- Johnson, K.M., 1984. *Bacillus cereus* in foodborne illness—An update. *J. Food Prot.* 47, 145–153.
- Kumari, S., Sarkar, P.K., 2016. *Bacillus cereus* hazard and control in industrial dairy processing environment. *Food Control.* 69, 20–29.
- Lindback, T., Fagerlund, A., Rødland, M.S., Granum, P.E., 2004. Characterization of the *Bacillus cereus* Nhe enterotoxin. *Microbiol.* 150, 3959–3967.
- Lund, T., De Buyser, M.L., Granum, P.E., 2000. A new cytotoxin from *Bacillus cereus* that may cause necrotic enteritis. *Mol. Microbiol.* 38, 254–261.
- Lund, T., Granum, P.E., 1996. Characterisation of a non-haemolytic enterotoxin complex from *Bacillus cereus* isolated after a foodborne outbreak. *FEMS Microbiol. Lett.* 141, 151–156.
- Marxen, S., Stark, T.D., Rutschle, A., Lucking, G., Frenzel, E., Scherer, S., Ehling-Schulz, M., Hofmann, T., 2015. Depsipeptide intermediates interrogate proposed biosynthesis of cereulide, the emetic toxin of *Bacillus cereus*. *Sci. Rep.* 5, 10637.
- Melling, J., Capel, B.J., Turnbull, P.C.B., Gilbert, R.J., 1976. Identification of a novel enterotoxigenic activity associated with *Bacillus cereus*. *J. Clin. Pathol.* 29, 938–940.
- Mohamed, A.S., Alnakip, M.E.A., Abd-El Aal, S.F., 2016. Occurrence of *Bacillus cereus* in raw milk and some dairy products in Egypt. *Jpn. J. Vet. Res.* 64, 95–102.
- Montville, T.J., Matthews, K.R., 2005. *Food Microbiology: An Introduction*. ASM Press, Washington D.C. 1st ed, pp. 120–123.
- Moravek, M., Dietrich, R., Buerk, C., Broussolle, V., Guinebretiere, M.H., Granum, P. E., Nguyen-The, C., Märtilbauer, E., 2006. Determination of the toxic potential of *Bacillus cereus* isolates by quantitative enterotoxin analyses. *FEMS Microbiol. Lett.* 257, 293–298.
- Rajkovic, A., Uyttendaele, M., Ombregt, S. A., Jaaskelainen, E., Salkinaja-Salonen, M., Debevere, J., 2006. Influence of type of food on the kinetics and overall production of *Bacillus cereus* emetic toxin. *J. Food Prot.* 69, 847–852.
- Ramaraao, N., Lereclus, D., 2005. The InhA1 metalloprotease allows spores of the *Bacillus cereus* group to escape macrophages. *Cell. Microbiol.* 7, 1357–1364.
- Ramaraao, N., Sanchis, V., 2013. The pore-forming haemolysins of *Bacillus cereus*: A review. *Toxins.* 5, 1119–1139.
- Reyes, J.E., Bastias, J.M., Gutiérrez, M.R., Rodríguez, M., 2007. Prevalence of *Bacillus cereus* in dried milk products used by Chilean School Feeding Program. *Food Microbiol.* 24, 1–6.
- Sastalla, I., Fattah, R., Coppage, N., Nandy, P., Crown, D., Pomerantsev, A. P., Leppla, S. H., 2013. The *Bacillus cereus* Hbl and Nhe tripartite enterotoxin components assemble sequentially on the surface of target cells and are not interchangeable. *PLoS One.* 8, e76955.
- Schneider, K. R., Schneider, R. M. G., Silverberg, R., Kurdmongkoltham, P., Bertoldi, B., 2017. Preventing Foodborne Illness: *Bacillus cereus*. *FSHN15-06/FS269*, rev. EDIS. 14,1–5.
- Schoeni, J.L., Wong, A.C.L., 2005. *Bacillus cereus* food poisoning and its toxins. *J. Food Prot.* 68, 636–648.
- Setlow, P., 2006. Spores of *Bacillus subtilis*: Their resistance to and killing

- by radiation, heat and chemicals. J. Appl. Microbiol. 101, 514–525.
- Soleimani, M., Hosseini, H., Pilevar, Z., Mehdizadeh, M., Carlin, F., 2018. Prevalence, molecular identification and characterization of *Bacillus cereus* isolated from beef burgers. J. Food Saf. 38, e12414.
- Soni, A., Oey, I., Silcock, P., Bremer, P., 2016. *Bacillus* spores in the food industry: A review on resistance and response to novel inactivation technologies. Compr. Rev. Food Sci. Food Saf. 15, 1139–1148.
- Spanu, C., Scarano, C., Spanu, V., Pala, C., Casti, D., Lamon, S., Cossu, F., Ibba, M., Nieddu, G., De Santis, E.P.L., 2016. Occurrence and behavior of *Bacillus cereus* in naturally contaminated ricotta salata cheese during refrigerated storage. Food Microbiol. 58, 135–138.
- Stenfors Arnesen, L.P., Fagerlund, A., Granum, P.E., 2008. From soil to gut: *Bacillus cereus* and its food poisoning toxins. FEMS Microbiol. Rev. 32, 579–606.
- Svensson, B., Eneroth, Å., Brendehaug, J., Molin, G., Christiansson, A., 2000. Involvement of a pasteurizer in the contamination of milk by *Bacillus cereus* in a commercial dairy plant. J. Dairy Res. 67, 455–460.
- Te Giffel, M.C., Beumer, R.R., Leijendekkers, S., Rombouts, F.M., 1996. Incidence of *Bacillus cereus* and *Bacillus subtilis* in foods in the Netherlands. Food Microbiol. 13, 53–58.
- Tran, S.L., Guillemet, E., Gohar, M., Lereclus, D., Ramarao, N., 2010. CwpFM (EntFM) is a *Bacillus cereus* potential cell wall peptidase implicated in adhesion, biofilm formation, and virulence. J. Bacteriol. 192, 2638–2642.
- Tran, S.L., Guillemet, E., Ngo-Camus, M., Clybourn, C., Puhar, A., Moris, A., Gohar, M., Lereclus, D., Ramarao, N., 2011. Haemolysin II is a *Bacillus cereus* virulence factor that induces apoptosis of macrophages. Cell. Microbiol. 13, 92–108.
- Vilain, S., Luo, Y., Hildreth, M.B., Brözel, V.S., 2006. Analysis of the life cycle of the soil saprophyte *Bacillus cereus* in liquid soil extract and in soil. Appl. Environ. Microbiol. 72, 4970–4977.