

# Immunomodulatory Role of Dietary Thyme against *Saprolegnia parasitica* Infection in Cultured Nile tilapia (*Oreochromis niloticus*)

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## Abstract

The fish industry has grown considerably worldwide, and fungal infections represent a significant aspect that increases economic losses and challenges through high mortality rates. The Egyptian aquaculture industry is particularly vulnerable to *Saprolegnia parasitica*, a deadly fish pathogen. Using phytobiotics as immunomodulators, antioxidants, and health promoters in aquaculture have been proven recently as an alternative strategy for banned malachite green. This research aimed to examine the thyme effect (*Thymus vulgaris*) on the immune status of cultured Nile tilapia against Saprolegniasis. A total of 50 fish (*Oreochromis niloticus*) with skin lesions were gathered from a private fish farm in Alexandria, Egypt. Skin swabs, gill swabs, and muscle tissue were obtained from each fish. After the mycological examination, results revealed that 35 isolates out of 150 examined samples (23.33%) were positive for fungal growth, of which 15 (10%) isolates were identified as *Saprolegnia* species. In addition, other fungi were detected; 5 (3.33%), 8 (5.33%), 4 (2.67%), and 3 (2%) isolates were identified as *Penicillium* species, *Aspergillus flavus*, *Alternaria* species, and *Fusarium* species, respectively. To evaluate the immunomodulatory effect of thyme, 300 healthy Nile tilapia fish with a mean weight of 30.0±5.0 g were brought to be experimentally designed; they were distributed into four groups (with three replicates) and were fed on an experimental diet including 0.0, 0.5, 1.0, and 1.5 thyme oil (gm/100gm diet) continuously for two months. Then fish were infected with *S. parasitica* zoospores, which were thoroughly mixed with their diet. According to results, after two months of feeding, catalase (CAT), serum lysozyme activity, and total protein dramatically increased according to the levels of thyme added; the acquisition was for the group fed on a 1.5 g/100g diet. Additionally, the expression of interleukin-10 (IL-10) and interleukin-1 $\beta$  (IL-1 $\beta$ ) in liver tissues increased similarly. Hence, it is concluded that employing thyme would improve the well-being and yield of the farmed Nile tilapia.

## KEYWORDS

Nile tilapia, *S. parasitica*, Thyme, Immunity, IL-10, IL-1 $\beta$ 

## INTRODUCTION

People in many countries rely heavily on freshwater fish as a source of protein (Hussain *et al.*, 2011). However, freshwater and marine fish populations have drastically declined, and fish farming has risen globally during the past ten years (Iqbal *et al.*, 2012); as a result, fish culture has become a critical industry commercially worldwide. One of the primary causes of financial losses in the ornamental and food fish farming businesses is fungal infections (Torto-Alalibo *et al.*, 2005; Ali, 2009). Saprolegniasis is one of the significant infections leading to considerable economic losses and high mortalities in freshwater fish. Moreover, it affects fish eggs, influencing high mortality within the hatching stage (Hussain *et al.*, 2013). Saprolegniasis is the leading cause of heavy winter mortalities in northern freshwater fish farms in Egypt (Emara *et al.*, 2020), including tilapia, considered the most widely cultivated fish in Egypt and the second worldwide after carp (Zahran *et al.*, 2017). *Saprolegnia* species are fungi-like organisms that fall within the oomycetes classification (Osman *et al.*, 2008). Oomycetes represent a typical saprophytic opportunist, infecting injured, infected, or stressed fish; however, they have

been reported in numerous reports as primary infection agents in fish (Roberts, 2012). *Saprolegnia parasitica* is linked to losses in various fish species, such as Nile tilapia (*Oreochromis niloticus*), where the mold is responsible for more than 95% of accumulative mortalities under experimental conditions (Ali *et al.*, 2019). *S. parasitica* deaths are more common in fish farms than in natural settings because the farmed animals are frequently subjected to ongoing stress and a variety of contaminants, which ultimately raise the spread and risk of the infection.

In most cases, Saprolegniasis is a white, cotton-like lesion on the head or dorsal fin that spreads to cover the entire body over time before turning red, brown, or green. In untreated cases, *Saprolegnia* infection leads to death by osmoregulatory failure (Rezinciuc *et al.*, 2018). The development of Saprolegniasis is significantly influenced by temperature; the majority of *Saprolegnia*-related fatalities are associated with the late autumn, winter, and early spring seasons, where outbreaks frequently occur at lower water temperatures (Kumar *et al.*, 2020). Moreover, other stress factors, such as water quality, handling, or crowding, are recurrently accompanied by outbreaks of Saprolegniasis (Ali *et al.*, 2013).

Multiple virulence factors are employed by *S. parasitica*, with hyphae deploying virulence factors against a fish cell. In addition, *Saprolegnia* spores secrete adhesive materials that can bind to lectins (Almeida et al., 2009). The transfer of effector proteins into infected cells causes the oomycete infection. *S. parasitica* is a pathogen with long-haired hook bundles that are rapidly generated within the secondary cyst phase. These bundles may act as a possible pathogenic factor by enhancing the ability of the host epidermis cells to adhere to the extracellular matrix and proteins (thrombospondin and fibronectin), resulting in tissue injury and dehydration due to the release of bodily fluids and poisonous substances (Wawra et al., 2012; Reziuc et al., 2018).

Infections are often associated with immunosuppression (Shehata and Abdel-Hakim, 2016), and there is no proven therapy for this infection. The routine application of disinfectants is commonly followed, including anti-*Saprolegnia* agents such as malachite green and formalin, which have recently been banned in some countries (Madrid et al., 2015). Therefore, previous studies recommended utilizing non-synthetic, safe alternative treatments, such as feed additives, that are required to improve the immune response, improve growth, and enhance resistance to certain diseases (Ali et al., 2014; Heikkinen et al., 2016).

One of the crucial issues in aquaculture is the effective control of *S. parasitica*. Therefore, it is suggested to employ a variety of plant extracts while treating aquaculture. Due to its variant bioactivities, including antioxidant, antibacterial, antiprotozoal, antifungal, and antiviral properties, thyme (*Thymus vulgaris*), an aromatic plant in the *Lamiaceae* family, has been extensively utilized in traditional medicine (Alsafah and Faragi, 2017). In addition, 4-allylphenol, carvacrol (15%), eugenol, cymene, and thymol (40%) are particularly abundant in thyme, reflecting its strong antioxidant effect (Sönmez et al., 2015).

Thus, the present study aimed to isolate and identify *Saprolegnia* species from infected Nile tilapia (*Oreochromis niloticus*) and assess the effect of thyme (*Thymus vulgaris*) on oxidative biomarkers, transcription of immune-related genes and antifungal activity of cultured Nile tilapia (*Oreochromis niloticus*) by improving its immune status against Saprolegniasis.

## MATERIALS AND METHODS

### Study design and fish sampling

A total number of 50 Nile tilapia fish (*Oreochromis niloticus*) having a mean weight of 30.0±5.0 g were obtained from a private fish farm in Alexandria, Egypt, suffered from gray, white skin lesions, detached scales from the body surface, and necrosis of fins and membranous part of gills. Samples were transmitted to the Bacteriology Unit in AHRI (Damanhour Provincial Lab) to be examined. A gross examination of dead and diseased live fish was carried out for the presence of lesions and ulceration. To remove surface bacterial pollutants, fish covered in fungal mats resembling cotton wool were washed with double-distilled water.

### Mycological examination

Skin lesions, infected gills, and infected muscles of the collected fish were swabbed and immersed in 0.1% sterile peptone water (Hashemi et al., 2012). The samples were inoculated and maintained at 25°C for up to 5 days on Sabouraud dextrose agar (SDA) containing chloramphenicol and chlortetracycline (100 mg of each), accompanied by routine daily monitoring for any anticipated fungal growth. A small amount of *Saprolegnia* species hyphae was sub-cultured on new plates of SDA media for purifi-

cation. The fungal isolates were placed on slides with coverslips in a lactophenol cotton blue dye solution and microscopically inspected for vegetative bodies and spores. Fungal isolates were characterized on the basis of colonial features, pigment production, and the micro-morphology of the spores produced (Ellis et al., 2007; Pitt and Hocking, 2009).

### Thyme extract preparation

A commercial product of thyme oil (Sigma-Aldrich, USA) was used to be incorporated into the experimental feed in the form of pellets (Dorojan et al., 2015). Four treatments (T2= 0.5, T3 = 1.0, and T4= 1.5 g/100 g diet) were used according to the thyme oil level. The control treatment (T1) did not get a thyme supplement. The pellets have been put in the air to dry well (Table 1).

Table 1. Ingredients and proximate chemical composition of experimental diet (g/kg on dry weight basis).

Ingredients	g/kg
Soybean meal (45% CP)	500
Fish meal (70.0% CP)	90
Wheat bran	200
Corn oil	15
Ground corn	100
Corn oil	20
Fish oil	45
Vitamin Premix <sup>1</sup>	15
Mineral Premix <sup>2</sup>	15

Vitamin Premix<sup>1</sup> (/kg in premix): vitamin A 67 IU, vitamin D 16.2 IU, vitamin E 7.4 g, vitamin K3 340 mg, vitamin B1 670 mg, vitamin B2 1000 mg, vitamin B6 800 mg, vitamin B12 1.4 mg, vitamin C 10 g, D-pantothenic acid 2.65 g, folic acid 330 mg, nicotinamide 5.35 g, choline chloride 35 g, biotin 34 mg, inositol 8g.  
Mineral Premix<sup>2</sup>: Fe 14 g, Cu 350 mg, Zn 4 g, Mn 1.4 mg, Mg 10 g, Co 30 mg, I 40 mg, Se 35 mg.

### Experimental fish rearing and management

A total of 300 Nile tilapia fish (*Oreochromis niloticus*) of 30.0±5.0 g average weight were brought from different farms in Alexandria, Egypt, and transmitted alive in polyethylene plastic bags supplemented with 2/3 air to the Fish Disease Unit in AHRI (Alexandria Provincial Lab). Then, they were acclimatized for 14 days and fed on a control diet with no feed additive supplementation (30% crude protein). After that, fish were put into prepared glass aquaria (100×80×60 cm) containing 100 L of water with continuous aeration with adjusted dissolved oxygen (5 mg/L) and an adjusted temperature of 25°C during the experimental period. Fish were divided into four groups (three replicates) and fed experimental diets containing thyme (0.5, 1.0, and 1.5 g/100 g diet) twice daily for two months. Clean, transparent water was applied all over the experiment, and the water was sampled every two weeks to monitor the quality. The water parameters were within normal limits for temperature (20–25°C), DO (5.4–5.7 mg/L), NH<sub>3</sub> (0.074–0.082 mg/L), and pH (7.4–7.6). The experiment settings were maintained within ranges that supported fish viability (Al-agawany et al., 2020).

### Challenge test

Isolated *Saprolegnia* strains were subcultured on fresh agar plates of SDA media with chloramphenicol and chlortetracycline (100 mg each). The tested *Saprolegnia* strains' zoospores were collected, counted, and adjusted to 1×10<sup>3</sup> in a Neubauer chamber (ERMA, Tokyo, Japan) (Stueland et al., 2005). The challenge

test was performed on each experimental group with *S. parasitica* zoospores, which were thoroughly mixed with a diet and provided twice daily for one month.

For 24 h, fish were deprived of food. Then three fish were obtained from every tank. Blood samples were drawn from caudal veins and were centrifuged at 3000 rpm for 15 min, and a portion of the liver was extracted and preserved in 2 ml RNA later at -80°C in liquid nitrogen. Sampling was carried out before and after infection by *S. parasitica*.

Antioxidant stress markers and non-specific immunity

Lysozyme assay

The lysozyme activity was measured as described by Kumari et al. (2006). Lysoplates were prepared by dissolving 1% agarose in 0.06 M PBS (pH 6.3) with 500 mg of LG1Micrococcus lysodeikticus added to 1 L of agarose. The agarose mixture was distributed onto six plates, each with a diameter of 4 to 5 nm. 25 µL of serum samples and standard lysozyme were added to each well. After 18 h, the diameter of the cleared zones was measured, and the lysozyme concentration was estimated.

Serum total protein determination

The method reported by Siwicki and Anderson (2000) was employed to determine the serum total protein concentration colorimetrically, using a commercial kit supplied by Spectrum, Cairo, Egypt.

Determination of liver tissue catalase (CAT) activity

The technique developed by Yarahmadi et al. (2016) was applied to measure liver catalase activity.

Gene expression analysis

Fish were randomly selected on day 60 of the feeding trial to analyze immune-related genes, including interleukin-1β (IL-1β) and interleukin-10 (IL-10). The primers used were supplied by Metabion (Germany). Table 2 demonstrates the cycling conditions, primer sequences, amplicon sizes, and target genes for SYBR-Green RT-PCR.

RNA extraction

RNA was extracted from tissue samples using the QIAamp RNeasy Mini kit (Qiagen, Germany, GmbH) by adding 30 mg of the tissue sample to 600 µl of RLT buffer containing ten µl of mercaptoethanol per ml. Tubes were inserted into adapter sets and clamps of the Qiagen tissue lyser to homogenize samples. For 2 min, high-speed (30 Hz) shaking steps were used to cause disruption. Finally, the cleaned lysate was mixed with one volume of 70% ethanol. The processes were carried out under the QIAamp RNeasy Mini Kit Purification of Total RNA from Animal Tissues Protocol (Qiagen, Germany, GmbH).

SYBR-Green RT-PCR

Primers were used in a 25 µl reaction, including 3 µl of the RNA template, 0.25 µl of RevertAid Reverse Transcriptase (200 U/µL), 8.25 µl of water, 0.5 µl of each primer at a concentration of 20 pmol, and 12.5 µl of the 2x QuantiTect SYBR-Green PCR Master Mix (Qiagen, Germany, GmbH). The reaction was performed uti-

lizing a Stratagene MX3005P real-time PCR apparatus (RT-PCR).

SYBR-Green RT-PCR analysis

Amplification curves and CT values were calculated by the Stratagene MX3005P. The CT of the positive control group was compared with that of each sample utilizing the "CtΔCt" method (Yuan et al., 2006), employing the following ratio to determine the variance of gene expression on the RNA of the various samples:

$$\Delta\Delta Ct = \Delta Ct \text{ reference} - \Delta Ct \text{ target}$$

$$\Delta Ct \text{ target} = Ct \text{ control} - Ct \text{ treatment and } \Delta Ct \text{ reference} = Ct \text{ control} - Ct \text{ treatment}$$

$$\Delta\Delta Ct = \Delta Ct \text{ reference} - \Delta Ct \text{ target}$$

$$\Delta Ct \text{ target} = Ct \text{ control} - Ct \text{ treatment}$$

Table 2. Primers sequences of the target genes and cycling conditions for SYBR-Green RT-PCR.

Target gene	Primers sequences	Reverse transcription		Primary denaturation	Amplification (40 cycles)			Dissociation curve (1 cycle)			Reference
		Reverse transcription	transcription		Secondary denaturation	Annealing	Extension	Secondary Denaturation	Annealing	Final denaturation	
<i>EF-1α</i>	CCTTCAACGGCTCAGGTCATC TGTGGCAGTGTGGCAATC	50°C 30 min	62°C 30 sec	94°C 15 min	62°C 30 sec	72°C 30 sec	94°C 1 min	62°C 1 min	94°C 1 min	Gröner et al. (2015)	
<i>IL-10</i>	CTGCTAGATCAGTCCGTCGAA GCAGAACCGTGTCCAGGTAA	50°C 30 min	60°C 30 sec	94°C 15 min	60°C 30 sec	72°C 30 sec	94°C 1 min	60°C 1 min	94°C 1 min	Staden et al. (2016)	
<i>IL-1 β</i>	GCTGGAGAGTCCGTGGAAGAAGATATAG CCTGGAGCATCATGGCGTG	50°C 30 min	62°C 30 sec	94°C 15 min	62°C 30 sec	72°C 30 sec	94°C 1 min	62°C 1 min	94°C 1 min	Castro et al. (2011)	

*EF-1α*: Eukaryotic translation elongation factor 1 alpha; *IL-10*: Interleukin-10; *IL-1 β*: interleukin-1β

Statistical analysis

SPSS V20 was utilized to analyze the data statistically. Data were shown as mean ± SE of three replicates. The Bartlett and Kolmogorov-Smirnov tests were applied to test the data for homogeneity of variances and normality of distribution prior to statistical analysis. The Duncan test was used as a post hoc test to analyze mean differences at a 5% probability level. Statistical significance was set at P < 0.05.

RESULTS

Incidence of *Saprolegnia* species

Mycological analysis of 50 inspected *Oreochromis niloticus* skins, gills, and muscles revealed 15 isolates of *Saprolegnia parasitica* (Table 3). The wet preparation of the gills, muscles, and skin revealed masses of immature and mature sporangia having a significant number of sporangia pores, and the hyphae emerged profusely branched and were non-septated, as shown in Figure 1. The positive colonies on SDA began with cysts of long hairs that were initially white and cottony in color before turning gray.

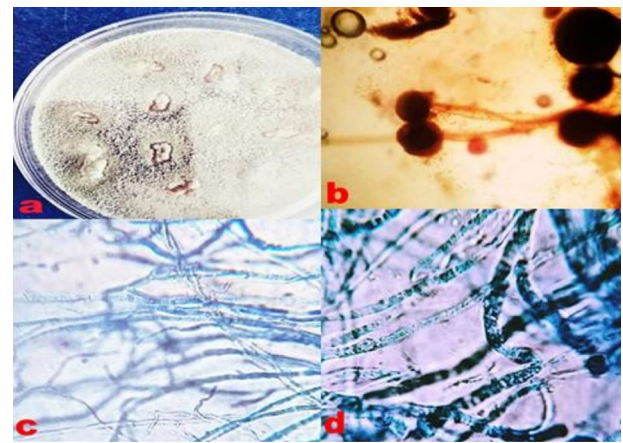


Fig. 1: a: *Saprolegnia* species cultures on SDA started as long hairs with whitish cottony color. b: The wet smear of skin showing masses of mature sporangia. c, d: The hyphae looked profuse, separated and were non-septated, stained with Lacto-phenol cotton blue, 400 X.

Immunity biomarkers

Serum lysozyme (LYZ) activities were influenced by thyme

diet supplementation, and lysozyme activity was remarkably raised (P<0.05) in the thyme diet concluded group according to dose percentage in comparison with the control group (Figure 2).

Oxidative stress indicators

The serum catalase enzyme (CAT) and total protein activities were remarkably affected by dietary thyme supplementation (P<

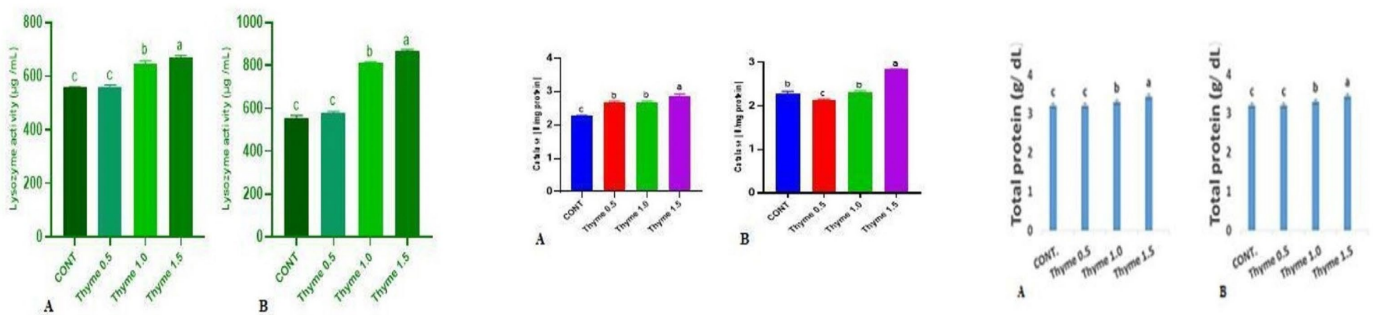


Fig. 2. Immune parameters in Nile Tilapia (*Oreochromis niloticus*) fed on diet with different levels of thyme (*Thymus vulgaris*) for 2 months (A: non-infected cases; B: infected cases). Data are expressed as ± SE. Different letters above bars indicate the significant difference among the treatments (P < 0.05).

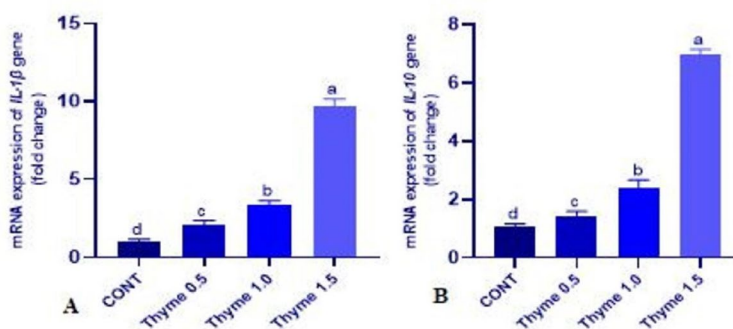


Fig. 3. mRNA levels of interleukin-1β (*IL-1β*) (A) and Interleukin 10 (*IL-10*) (B) genes in the liver of Tilapia fish fed diets supplemented with thyme oil for 2 months.

Table 3. Incidence of molds in the examined Nile Tilapia fish

Sources of examined samples (50 for each)	<i>Saprolegnia parasitica</i> No. (%)	<i>Penicillium</i> species No. (%)	<i>Aspergillus flavus</i> No. (%)	<i>Alternaria</i> No. (%)	<i>Fusarium</i> No. (%)
Skin swabs	7 (14)	3 (6)	4 (8)	2 (4)	0 (0)
Gills swabs	5 (10)	1 (2)	3 (6)	2 (4)	1(2)
Muscles tissue	3 (6)	1 (2)	1 (2)	0 (0)	2 (4)
Total (150)	15 (10)	5 (3.33)	8 (5.33)	4 (2.67)	3 (2)



0.05) according to dose percentage in comparison with the control group (Figures 2).

#### Liver gene expression

Thyme-contained diets remarkably encouraged *IL-10* and *IL-1 $\beta$*  gene expression in all thyme groups (Figure 3), and the top expression levels were observed in the 1.5 g of thyme per 100 g diet, but the lowest levels were in the control group.

## DISCUSSION

Tilapia species constitute the significant bulk of the Nile River, with more than 33.0% of the total catches (Gafard, 2015). The aquaculture industry has been considered one of the fastest-growing agribusinesses. The spread of *S. parasitica* infections in Nile tilapia has dramatically increased since malachite green, the most potent anti-*Saprolegnia* chemical, was prohibited in many regions worldwide. Mycological examination of 50 Nile tilapia fish (*Oreochromis niloticus*) with skin lesions revealed that isolated molds belonged to the following genera: *Saprolegnia*, *Penicillium*, *Aspergillus flavus*, *Alternaria*, and *Fusarium*. Similar results were recorded by Ammar (2001), El-Ahl (2010), and Refai et al. (2010). The fungal contamination of fish could be attributed to improper sanitation, contaminated feeds, and water supply, and workers' hands influencing the fish's health status (Kumar et al., 2020). Our findings about the infection rate of *S. parasitica* in Nile tilapia agreed with that obtained by Phillips et al. (2008) and Zahran et al. (2017), who described *S. parasitica* as one of the essential mycotic infections leading to economic loss in cultured ecosystems.

Robert et al. (2003) revealed that rapid decreases in water temperature impaired the fish's immune system and temporarily suppressed mucus production by goblet cells in the dermal layers of the skin, which acted as a physical barrier preventing fungal spores from contacting the fish skin. Additionally, mucus has antimicrobial elements that can eliminate invasive zoospores, including proteolytic enzymes, complement, lysozyme, C-reactive protein, and immunoglobulin. Without mucus, the skin is exposed, and fungi start growing in numbers and extending their hyphae into the muscular tissue.

One of the most promising strategies for disease management in aquaculture is enhancing fish defense mechanisms through the prophylactic administration of natural plant products. Plant extract is more valuable and eco-friendly, and its application has increased significantly after malachite green treatment was forbidden worldwide. Thyme (*Thymus vulgaris*) is a herb that has been used in traditional medicine for a variety of purposes, including antitussive, antioxidant, antispasmodic, bronchodilator, anti-asthmatic, expectorant, carminative, anthelmintic, antimicrobial, and antiseptic (Ocaña and Reglero, 2012; Alsafah and Al-Faragi, 2017; Soliman et al., 2021). Additionally, essential thyme oil has a higher efficacy as an antifungal against *Saprolegnia* species.

Regarding the immunomodulatory effect of *Thymus vulgaris*, the current study confirmed that it had many benefits for Nile tilapia health by increasing the level of Lysozyme, a crucial bactericidal enzyme, with abundance in epithelial secretions. Lysozymes are fish's most essential immunity factors to resist infections (Mirghaed et al., 2020). In this study, lysozyme secretion in fish was increased by thyme treatment. According to Farsani et al. (2019), the dietary herbal treatment improved lysozyme activity, which aided fish in resisting infections. Several studies have approved this theory, which showed that thyme treatment stimulated fish immune responses through increasing lysozyme secretion (Perez-Sánchez et al., 2015; Diler et al., 2016; Hoseini and Yousefi, 2019; Zargar et al., 2019; Yousefi et al., 2022).

In this study, thyme treatment stimulated the immune response by increasing total serum protein, constituting a signifi-

cant measure of the fish's nutritional condition and overall health (Hoseini and Tarkhani, 2013). To the same extent, Hoseini and Yousefi (2019) and Yousefi et al. (2022) proved that thyme had hepatoprotective effects.

Natural antioxidant catalase is one of the most important antioxidant enzymes that protect fish from oxidative damage due to free radicals, nitrogen species, and reactive oxygen (Halliwell and Gutteridge, 2007). The present results showed that thyme administration elevates catalase levels in the same manner that Zheng et al. (2009) approved after using thyme to control *S. parasitica* in freshwater fish. Similarly, in rainbow trout, thyme oil administration significantly increased the catalase enzyme (Giannenas et al., 2012).

Interleukin (IL) represents a cytokine that promotes inflammation and is crucial for innate immunity. In humans, there are 100 different forms of IL. Recent genetic investigations have revealed that the first cytokine in fish was IL-1 (Wang et al., 2009). In mammals, the *IL-1 $\alpha$*  and *IL-1 $\beta$*  genes are located on the same chromosome and are close to each other. However, only the *IL-1 $\beta$*  gene has been discovered in fish thus far.

In this study, significant increases in IL-10 and IL-1 $\beta$  were achieved by increasing the concentration of thyme. IL-1 $\beta$  influenced a fish's immune system by enhancing phagocytosis, stimulating the lysozyme activities of macrophages (Hoseini and Yousefi, 2019), and modulating IL-17 family members' expression, which was an essential defense against infections (Kono et al., 2011). The current results were in line with Zargar et al. (2019) and Yousefi et al. (2022), who found that *Thymus vulgaris* essential oil considerably upregulated the degrees to which immune-related genes such as the cluster of differentiation 4 (CD4), the lysozyme gene, complement 3 (C3), and *IL-1 $\beta$*  were expressed. According to previous research, bacterial infection, LPS stimulation, and the administration of immune stimulants could enhance *IL-10* expression (Zhang et al., 2009), and this agreed with our finding of a significant increase in IL-10 compared with the control in the case of elevated thyme concentration. IL-10 represented a pleiotropic regulatory and a crucial anti-inflammatory cytokine that regulated the immune response, preventing the severe consequences of inflammation (Moore et al., 2001).

In this study, the highest levels of lysozyme (LYZ), total protein activities, and serum catalase enzyme (CAT), as well as a significant increase in *IL-10* and *IL-1 $\beta$*  genes, were at doses of 1.5 g/100 g and 1.0 g/100 g *Thymus vulgaris* diet; similar findings were observed by (Zaki et al., 2012).

## CONCLUSION

In general, our study suggests that *Thymus vulgaris* has the potential to be used as a healthy control for Nile tilapia through increasing immunity. This reflects their future potential application as preventive measures against winterkill through frequent application in commercial aquaculture, including broodstock and fingerling overwintering.

## CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

## REFERENCES

- Alagawany, F., Farag, M.R., Abdelnou, S.A., 2020. A review on the beneficial effect of thymol on health and production of fish. *Reviews in Aquaculture* 13, 632-641. <http://doi.10.1111/raq.12490>
- Ali, E.H., 2009. Antifungal activity of sodium chloride on *Saprolegnia diclina* and *Aphanomyces* sp. *Acta Mycol.* 44, 125-38.
- Ali, S.E., Amr, A.A., Skaar, G.I., Evensen, Q., Charo-Karisa, H., 2019. Efficacy and safety of boric acid as a preventive treatment against *Saprolegnia* infection in Nile tilapia (*Oreochromis niloticus*). *Sci. Rep.* 9, 18013. <http://doi.10.1038/s41598-019-54534-y>
- Ali, S.E., Thoen, E., Evensen, Ø., Skaar, I., 2014. Boric acid inhibits germination and colonization of *Saprolegnia* spores *in vitro* and *in vivo*.

- PLoS One 9, e91878. <http://doi:10.1371/journal.pone.0091878>
- Ali, S.E., Thoen, E., Vralstad, T., Kristensen, R., Evensen, O., Skaar, I., 2013. Development and reproduction of *Saprolegnia* species in biofilms. *Vet. Microbiol.* 163, 133-141.
- Almeida, A., Cunha, A., Gomes, N.C.M., Alves, E., Costa, L., Faustino, M.A.F., 2009. Phage therapy and photodynamic therapy: low environmental impact approaches to inactive microorganisms in fish farming plants. *Drugs* 7, 268-313. <http://doi:10.3390/md7030268>
- AlSafah, A.H., AL-Faragi, J.K., 2017. Influence of thyme (*Thymus vulgaris*) as feed additives on growth performance and antifungal activity on *Saprolegnia* spp. in *Cyprinus carpio* L. *Journal of Entomology and Zoology Studies* 5, 1598-1602.
- Ammar, M.A.M., 2001. Sanitary assessment of some common fresh water fish in Assiut. M.V.Sc. Thesis, Fac. Vet. Med., Assuit Univ, Egypt.
- Castro, R., Zou, J., Secombes, C.J., Martin, S.A.M., 2011. Cortisol modulates the induction of inflammatory gene expression in a rainbow trout macrophage cell line. *Fish and Shellfish Immunology* 30, 215-223.
- Diler, O., Gormez, O., Diler, I., Metin, S., 2016. Effect of oregano (*Origanum onites* L.) essential oil on growth, lysozyme and antioxidant activity and resistance against *Lactococcus garvieae* in rainbow trout, *Oncorhynchus mykiss* (Walbaum). *Aquaculture Nutrition* 23, 844-851. <https://doi.org/10.1111/anu.12451>
- Dorojan, O.G., Cristea, V., cretu, M., Coadă, M.T., Dediu, L., Grecu, I.R., 2015. Effect of thyme (*Thymus vulgaris*) and vitamin E on growth performance and body composition of *Acipenser stellatus* juveniles. *Aquaculture, Aquarium, Conversation and Legislation – International Journal of the Bioflux Society* 2015, 8, 195-202.
- El-Ahl, M.H.S., 2010. Studies on fungi in fish and fish products and their control. Ph.D. Thesis, Dept. of Microb, Fac. of Vet. Med., Cairo Univ., Egypt.
- Ellis, D., Davis, S., Alexiou, H., Handke, R., Bartley, R., 2007. Descriptions of Medical Fungi. Nexus Print Solutions, Adelaide, South Australia, Australia.
- Emara, E.K.M., Gaafar, A.Y., Shetaia, Y.M., 2020. In vitro screening for the antifungal activity of some Egyptian plant extracts against the fish pathogen *Saprolegnia parasitica*. *Aquaculture Research* 51, 4461-4470. <https://doi.org/10.1111/are.14791>
- Farsani, M.N., Hoseinifar, S.H., Rashidian, G., Ghafari, H., Ashouri, F.G., Van Doan, H., 2019. Dietary effects of *Coriandrum sativum* extract on growth performance, physiological and innate immune responses and resistance of rainbow trout (*Oncorhynchus mykiss*) against *Yersinia ruckeri*. *Fish Shellfish Immunol.* 91, 233-240. <https://doi.org/10.1016/j.fsi.2019.05.031>
- Gafard, M., 2015. General Authority for Fish Resources Development. Annual fishery statistics book, Cairo, Egypt.
- Giannenas, I., Triantafyllou, E., Stavrakakis, S., Margaroni, M., Mavridis, S., Steiner, T., Karagouni, E., 2012. Assessment of dietary supplementation with carvacrol or thymol containing feed additives on performance, intestinal microbiota and antioxidant status of rainbow trout (*Oncorhynchus mykiss*). *Aquaculture* 350, 26-32. <https://doi.org/10.1016/j.aquaculture.2012.04.027>
- Gröner, F., Ziková, A., Kloas, W., 2015. Effects of the pharmaceuticals diclofenac and metoprolol on gene expression levels of enzymes of biotransformation, excretion pathways and estrogenicity in primary hepatocytes of Nile tilapia (*Oreochromis niloticus*). *Comparative Biochemistry and Physiology, Part C* 167, 51-57.
- Halliwell, J.M.C., Gutteridge, M., 2007. Free Radicals in Biology and Medicine, fourth ed., Oxford University Press.
- Hashemi, K.S.M., Sadeghpour, H.M., Gholampour, A.I., 2012. Isolation of *Saprolegnia* and the Influence of Root Ethanolic Extract of *Ruta graveolens* on *Saprolegnia* spp Growth. *International Journal of Bioscience, Biochemistry and Bioinformatics* 2, 64-67.
- Heikkinen, J., Tiirola, M., Mustonen, S.M., Eskelinen, P., Navia-Paldanius, D., von Wright, A., 2016. Suppression of *Saprolegnia* infections in rainbow trout (*Oncorhynchus mykiss*) eggs using protective bacteria and ultraviolet irradiation of the hatchery water. *Aquac Res.* 47, 925-939.
- Hoseini, S. M., Tarkhani, R., 2013. Effect of short-term treatment with potassium permanganate on stress markers and blood biochemistry in goldfish *Carassius auratus*. *Aquaculture Research* 44, 869-875. <https://doi.org/10.1111/j.1365-2109.2012.03091.x>
- Hoseini, S.M., Yousefi, M., 2019. Beneficial effects of thyme (*Thymus vulgaris*) extract on oxytetracycline-induced stress response, immunosuppression, oxidative stress and enzymatic changes in rainbow trout (*Oncorhynchus mykiss*). *Aquaculture Nutrition* 25, 298-309.
- Hussain, M., Hussain, W.H., Mohamad, M.A., 2013. Pathogenicity of *Achlya proliferoids* and *Saprolegnia diolina* (*Oreochromis niloticus*) associated with Saprolegniasis outbreaks in cultured Nile Tilapia (*Oreochromis niloticus*). *World Journal of Fish and Marine Science* 5, 188-193.
- Hussain, S.M., Javed, M., Javid, A., Javid, T., Hussain, N., 2011. Growth responses of *Catla catla*, *Labeo rohita* and *Cirrhina mrigala* during chronic exposure of iron. *Pak J Agric Sci.* 48, 225-230.
- Iqbal, Z., Sheikh, U., Mughal, R., 2012. Fungal infections in some economically important freshwater fishes. *Pak. Vet. J.* 32, 422-426.
- Kono, T., Korenaga, H., Sakai, M., 2011. Genomics of fish IL-17 ligand and receptors: a review. *Fish and Shellfish Immunology* 31, 635-643. <http://doi:10.1016/j.fsi.2010.11.028>
- Kumari, J., Sahoo, P.K., Swain, T., Sahoo, S.K., Sahu, A.K., Mohanty, B.R., 2006. Seasonal variation in the innate immune parameters of the Asian catfish *Clarias batrachus*. *Aquaculture* 252, 121-127. <https://doi.org/10.1016/j.aquaculture.2005.07.025>
- Kumar, S., Mandal, R.S., Bulone, V., Srivastava, V., 2020. Identification of Growth Inhibitors of the Fish Pathogen *Saprolegnia parasitica* Using in silico Subtractive Proteomics, Computational Modeling, and Biochemical Validation. *Front Microbiol.* 11, 571093. <http://doi:10.3389/fmicb.2020.571093>
- Madrid, A., Godoy, P., González, S., Zaror, L., Moller, A., Werner, E., Cuelar, M., Villena, J., Montenegro, I., 2015. Chemical characterization and anti-oomycete activity of Laureliopsis philippianna essential oils against *Saprolegnia parasitica* and *S. australis*. *Molecules* 20, 8033-8047. <http://doi:10.3390/molecules20058033>
- Mirghaed, A.T., Hoseini, S.M., Hoseinifar, S.H., Doan, H.V., 2020. Effects of dietary thyme (*Zataria multiflora*) extract on antioxidant and immunological responses and immune-related gene expression of rainbow trout (*Oncorhynchus mykiss*) juveniles. *Fish and Shellfish Immunology* 106. <http://doi:10.1016/j.fsi.2020.08.002>
- Moore, K.W., de Waal, M.R., Coffman, R.L., O'Garra, A., 2001. Interleukin-10 and the interleukin-10 receptor. *Annu Rev Immunol.* 19, 683-765. <http://doi:10.1146/annurev.immunol>
- Ocaña, A., Reglero, G., 2012. Effects of thyme extract oils (from *Thymus vulgaris*, *Thymus zygis*, and *Thymus hemaleis*) on cytokine production and gene expression of oxLDL-stimulated THP-1-macrophages. *J. Obes.* 2012, 104706. <http://doi:10.1155/2012/104706>
- Osman, H.A., Solman, W.E., Noor El Deen, A.E., Mohamed, L.A., 2008. Induction of Saprolegniasis in *Oreochromis niloticus* with special reference to its biological control. *Global Vet.* 2, 32-37.
- Perez-Sánchez, J., Benedito-Palos, L., Estensoro, I., Petropoulos, Y., Calduch-Giner, J.A., Browdy, C.L., Sitja-Bobadilla, A., 2015. Effects of dietary NEXT ENHANCE® 150 on growth performance and expression of immune and intestinal integrity related genes in gilthead sea bream (*Sparus aurata* L.). *Fish and Shellfish Immunology.* 44, 117-128. <https://doi.org/10.1016/j.fsi.2015.01.039>
- Phillips, A.J., Anderson, V.L., Robertson, E.J., Secombes, C.J., Van, W.P., 2008. New insights into animal pathogenic oomycetes. *Trends Microbiol.* 16, 13-19. <http://doi:10.1016/j.tim.2007.10.013>
- Pitt, J.I., Hocking, A.D., 2009. *Fungi and Food Spoilage*. 3<sup>rd</sup> Ed. Published by Springer Dordrecht Heidelberg, London, New York.
- Refai, M.K., Laila, A., Amany, M., Kenawy, E., Shima, S.M.A., 2010. The Assessment of Mycotic Settlement Of Freshwater Fishes In Egypt. *Journal of American Science* 6, 595-602.
- Rezinciuc, S., Sandoval-Sierra, J.V., Ruiz-León, Y., van West, P., Diéguez-Urbeondo, J., 2018. Specialized attachment structure of the fish pathogenic oomycete *Saprolegnia parasitica*. *PLoS One* 13, e0190361. <https://doi.org/10.1371/journal.pone.0190361>
- Robert, M.D., David, J.W., Jeffery, S.T., 2003. Saprolegniasis (Winter Fungus) and Branchiomycosis of Commercially Cultured Channel Catfish SRAC Publication No. 4700
- Roberts, R.J., 2012. *Fish pathology*. 4<sup>th</sup> ed. Wiley-Blackwell, USA.
- Shehata, A.H.S., Abdel-Hakim, S.A., 2016. Treatment Trails of Saprolegniasis in *Oreochromis niloticus*. *Alex. J. Vet. Sci.* 49,99-104. <http://doi:10.5455/ajvs.226020>
- Siwicki, A., Anderson, D., 2000. Nonspecific Defense Mechanisms Assay in Fish: II. Potential Killing Activity of Neutrophils and Macrophages, Lysozyme Activity in Serum and Organs and Total Immunoglobulin Level in Serum. *FAO project GCP/ INT/JPA, IFI, Olsztyn, Poland*, pp. 105-112.
- Soliman, M.M., Adil, A., Mohammed, M.M., 2021. Hepatoprotective effect of *Thymus vulgaris* extract on sodium nitrite-induced changes in oxidative stress, antioxidant and inflammatory marker expression. *Sci Rep.* 11, 5747 <http://doi:10.1038/s41598-021-85264-9>
- Sönmez, A.Y., Bilen, S., Alak, G., Hisar, O., Yanik, T., Biswas, G., 2015. Growth performance and antioxidant enzyme activities in rainbow trout (*Oncorhynchus mykiss*) juveniles fed diets supplemented with sage, mint and thyme oils. *Fish Physiol Biochem.* 41,165-175. <http://doi:10.1007/s10695-014-0014-9>
- Standen, B.T., Peggs, D.L., Rawling, M.D., Foey, A., Davies, S.J., Santos, G.A., Merrifield, D.L., 2016. Dietary administration of a commercial mixed-species probiotic improves growth performance and

- modulates the intestinal immunity of tilapia, *Oreochromis niloticus*. Fish and Shellfish Immunology 49, 427e435.
- Stueland, S., Heier, B.T., Skaar, I., 2005. A simple in vitro screening method to determine the effects of drugs against growth of *Saprolegnia parasitica*. Mycological Progress 4, 273-279.
- Torto-Alalibo, T., Tian, M., Gajendran, K., Waugh, M., van West, P., Kamoun, S., 2005. Expressed sequence tags from the oomycete fish pathogen *Saprolegnia parasitica* reveals putative virulence factors. BMC Microbial. 5, 46. <http://doi.org/10.1186/1471-2180-5-46>
- Wang, T., Bird, S., Koussounadis, A., Holland, J.W., Carrington, A., Zou, J., Secombes, C.J., 2009. Identification of a novel IL-1 cytokine family member in teleost fish. J Immunol. 183, 962-974. <http://doi.org/10.4049/jimmunol.0802953>
- Wawra, S., Bain, J., Durward, E., de Bruijn, I., Minor, K.L., Matena, A., Löbach, L., Whisson, S.C., Bayer, P., Porter, A.J., Birch, P.R.J., Secombes, C.J., van West, P., 2012. Host-targeting protein 1 (SpHtp1) from the oomycete *Saprolegnia parasitica* translocates specifically into fish cells in a tyrosine-O-sulphate-dependent manner. Proc Natl Acad Sci USA. 109, 2096-2101.
- Yarahmadi, P., Ghafari, F.H., Khazaei, A., Khodadadi, M.G., Jalali, M.A., 2016. Protective effects of the prebiotic on the immunological indicators of rainbow trout (*Oncorhynchus mykiss*) infected with *Aeromonas hydrophila*. Fish Shellfish Immunol. 54, 589-597.
- Yousefi, M., Ghafarifarsani, H., Hoseini, S.M., Hoseinifar, S.H., Abtahi, B., Vatnikov, Y.K., Kulikov, E.V., Van-Doan, H., 2022. Effects of dietary thyme essential oil and prebiotic administration on rainbow trout (*Oncorhynchus mykiss*) welfare and performance. Fish and Shellfish Immunology. 120, 737-744. <https://doi.org/10.1016/j.fsi.2021.12.023>
- Yuan, J.S., Reed, A., Chen, F., Stewart, C.N., 2006. Statistical analysis of real time PCR data B.M.C. Bioinformatics. 22, 85. <http://doi.org/10.1186/1471-2105-7-85>
- Zahran, E., Hafez, E.E., Hossain, F.M.A., Elhadidy, M., Shaheen, A.A., 2017. Saprolegniosis in Nile Tilapia: Identification, Molecular Characterization, and Phylogenetic Analysis of Two Novel Pathogenic *Saprolegnia* Strain. Journal of Aquatic Animal Health 29, 43-49. <https://doi.org/10.1080/08997659.2016.125969>
- Zaki, M., Alabib, E.M., Nour, A.M., Tonsy, H.D., Mahmoud, S.H., 2012. Effect of some medicinal plants diet on mono sex Nile tilapia (*Oreochromis niloticus*), growth performance, feed utilization and physiological parameters. Egyptian journal of aquatic biology and fisheries. 4, 220-227. <http://doi.org/10.21608/EJABF.2011.2101>
- Zargar, Z., Rahimi-Afzal, E., Soltani, A., Taheri, M.H.A., Ebrahimzadeh-Mousavi, M., Soltani, P.Y., 2019. Growth performance, immune response and disease resistance of rainbow trout (*Oncorhynchus mykiss*) fed *Thymus vulgaris* essential oils. Aquacult Res. 50, 3097-3106. <https://doi.org/10.1111/are.14243>
- Zhang, Z., Swain, T., Bogwald, J., Dalmo, R.A., Kumari, J., (2009). Bath immunostimulation of rainbow trout (*Oncorhynchus mykiss*) fry induces enhancement of inflammatory cytokine transcripts, while repeated bath induce no changes. Fish Shellfish Immunol. 26, 677-678. <http://doi.org/10.1016/j.fsi.2009.02.014>
- Zheng, Z., Tan, J.Y., Liu, H., Zhou, X., Xiang, X., Wang, K., 2009. Evaluation of oregano essential oil (*Origanum heracleoticum* L.) on growth, antioxidant effect and resistance against *Aeromonas hydrophila* in channel catfish (*Ictalurus punctatus*). Aquaculture 292, 214-218. <https://doi.org/10.1016/j.aquaculture.2009.04.025>