

Competence of *Panax ginseng* on Male Fertility in Cypermethrin Exposed Rats

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Abstract

The rising occurrences of male reproductive disorders, as decreased semen (quality and quantity) and testicular cancer, are of great concern for animal production. Therefore, this study was conducted to determine the toxic impact of cypermethrin on the fertility of male rats. Other goal was using the *Panax ginseng* as androgenic drug for reducing the negative impact of cypermethrin poisoned rats. In the results, oral LD₅₀ of cypermethrin in mature male rats was found to be 374.633±12.187 mg/kg. Based on that, thirty-two adult male rats were equally divided into four groups. Group (1) served as -ve control; group (2) treated with 0.1% Panax in feed and kept as +ve control; group (3) orally dosed 1/40 LD₅₀ of cypermethrin; and group (4) intubated the same dose of cypermethrin and panax admixed with diet for 60 days. The results demonstrated that cypermethrin generated obvious disorders in male fertility as evidenced by a significant decrease in the blood testosterone, LH and FSH hormones, testes weights, sperm count and motility, and live sperm percentage. Sperm cell abnormalities were significantly elevated in cypermethrin poisoned rats compared. Although the group treated by cypermethrin with panax showed a lesser effect. Cypermethrin intoxication showed sever alteration in fertility indices and fetal values. Histopathological examinations on the testes, seminal vesicles, and prostate glands served as confirmation for all findings. A positive control group didn't significantly differ as compared to a negative control. Otherwise, it recorded the best results in percentage of motile and live sperm, especially drops of sperm cell abnormalities. Results concluded that involvement with cypermethrin caused overt defects in male reproductive function, which were cured by administering the androgenic drug Panax.

KEYWORDS

Panax ginseng, Cypermethrin, Fertility, Male rats, Semen quality.

INTRODUCTION

Panax ginseng is a traditional herbal remedy that has been used in China, Japan, South Korea, and other East Asian nations. It has now spread to more than 30 additional countries, where it is now one of the most widely used herbal supplements for health care (Attele *et al.*, 1999). The human brain system, cardiovascular system, immunological system, reproductive system, and endocrine system are all positively regulated by *Panax ginseng* (Kee *et al.*, 2018).

The rising occurrences of male reproductive disorders, as decreased semen (quality and quantity) and testicular cancer, are of great concern for animal production (Andersson *et al.*, 2008; Joshi *et al.*, 2011). Numerous investigations revealed that a variety of environmental toxins, including pesticides, are at least partially responsible for the declining health of male reproductive systems in both people and wildlife (Swan *et al.*, 2003; Lifeng *et al.*, 2006).

A variety of broad-spectrum insecticides with a structure like that of natural pyrethrums are known as synthetic pyrethroids (Harlod *et al.*, 2003; Yilmaz *et al.*, 2008). One of the most often used type II synthetic pyrethroid pesticides is cypermethrin (Assayed *et al.*, 2010). It is widely used to protect agricultural crops, and can accumulate, contaminate the environment, and even-

tually disrupt the food chain. Numerous harmful reproduction defects are brought on by cypermethrin exposure in both human and animal models (Sharma *et al.*, 2018).

In the past, tests using the androgen receptor gene showed that cypermethrin might cause oestrogen transactivity and anti-androgenic effects (Hu *et al.*, 2013; Kjeldsen *et al.*, 2013). In experimental animal models, exposure to cypermethrin drastically reduced the weights of the testes, epididymis, seminal vesicles, and prostate (Sharma *et al.*, 2018; El-Sheshtawy *et al.*, 2019; Katragadda *et al.*, 2021).

Additionally, several studies revealed that cypermethrin inhibits testosterone levels, changes sperm maturation processes, and reduces spermatogenesis (El-Sheshtway *et al.*, 2016; Sharma *et al.*, 2018). Numerous studies have asserted that cypermethrin administration in rodent models affects the levels of pituitary gonadotropins in the blood, including luteinizing hormone and follicle-stimulating hormone (Li *et al.*, 2013; Hashem *et al.*, 2015; Sharma *et al.*, 2018; Vasudha *et al.*, 2018). It is clear from this that cypermethrin therapy inhibits testosterone production, at least in part (Travison *et al.*, 2008).

The goal of the current study was to determine the poisoning effect of cypermethrin on the fertility in adult male rats. On the other hand, using of *Panax ginseng* as an antioxidant and androgenic medications for reducing the negative impact of cyperme-

thrin poisoned rats.

MATERIALS AND METHODS

Cypermethrin

Cypermethrin (Cyperguard 10% EC®) was freshly prepared in distilled water before oral administration (100g/liter).

Panax

Panax ginseng (Imtenan health company) was made from roots of genuine *Panax ginseng* which imported from East Asian countries that described by company.

Experimental animals

A total of 76 adult albino rats, average weighing from 120-150g were used (52 male+24 female) in this study.

Ethical Statement

The experiment was conducted with following the national and international ethical guidelines.

Experimental design

Twenty adult male rats were used for determination of the acute oral LD₅₀. In addition, thirty two adult male rats were divided into four groups: each group of 8 rats for the male reproductive toxicity study.

Group 1: kept without treatment, as a negative control. Group 2: given 0.1% Panax daily at a concentration level admixed with feed and kept as positive control group. Group 3: Dosed orally with cypermethrin 5 times/week at concentration level of 9.4 mg/kg equivalent to 1/40 of the estimated LD₅₀. Group 4: dosed orally with cypermethrin 1/40 LD₅₀, 5 times/week with the addition of 0.1% panax daily. All treatments were extended for 60 days to cover all spermatogenic cycles (Jackson and Jones, 1966). After 60 days, 5 rats from each group were sacrificed for revealing reproductive toxicity studies as well as for histopathological examination. The rest of the 3 rats from each group were paired with 24 healthy, untreated female rats (1:2) to evaluate the male fertility index.

Blood and tissues sampling

Blood samples were obtained from the retro-orbital venous plexus at the end of the experiment (Halperin et al., 1953). Serum samples were separated for hormonal examination. Rats were sacrificed by anesthesia inhalation under light diethyl ether. For histological monitoring, seminal vesicles, testes, and prostate glands were removed, weighed, and preserved in 10% neutral buffered formalin.

Weight of sex organs

The prostate gland, testes, and seminal vesicles were removed and weighed; the relative weight was calculated according to 100g of body weight.

Spermatozoa examination

The sperm were obtained by maceration of the epididymis

and vas deferentia (Blandau and Jordan, 1941) and checked for the number of sperm cells (Reddy and Bordekar, 1999), progressive motility percent, live and dead sperm percent, and spermatozoa abnormalities as preceded by Bearden and Fluquary (1984).

Hormonal assay

The LH, FSH, and testosterone were estimated in serum using the Rat/Mouse ELISA TEST KIT is produced by Abnova company method as described by Sakuma (2009) and Goodman (1992).

Fertility index

It was done matching to method described by Wand and Col-in (1998). Morphological examination of the gained foetuses was performed according to Cook and Fairweather (1968).

Histopathological examination

Rats were sacrificed, and autopsy samples from the testes, seminal vesicles, and prostate glands were preserved in a 10% neutrally buffered formalin solution for at least one day. Then, the samples were regularly treated using the normal paraffin embedding procedure, and hematoxylin and eosin were applied to them (Bancroft et al., 1994).

Statistical analysis

The data collected were statistically examined, and significance was determined using the Duncan test and computerized using the Analysis of Variance (ANOVA).

RESULTS

LD₅₀ of cypermethrin

The acute oral LD₅₀ of cypermethrin was calculated as 374.633±12.187 mg/kg in adult male rats.

Clinical signs of cypermethrin toxicity

Clinical signs of cypermethrin toxicity were noticed at the group of cypermethrin treated rats, such as dyspnea, thick eye discharge, intermittent diarrhea, and loss of body weight, during the first few days of intoxication. Later, animals adapted, and gradually, their body condition improved.

Effect of cypermethrin and panax, on the male rats' genital weight

As reported in Table 1, there was a significant decrease in the relative weight of testes, seminal vesicles, and prostate glands at P < 0.05 in the cypermethrin-treated group as compared to all groups. However, the group of cypermethrin-combined with Panax partially improved the values of the relative weight for testes, seminal vesicles, and prostate gland, but body weight still significantly decreased in comparison to the -ve and +ve control groups.

Effect of cypermethrin and Panax on hormonal analysis

Results in Table 2 indicated a noticeable decline (P<0.05) in levels of testosterone, LH and FSH compared to cypermethrin-treated group, while group of cypermethrin with Panax recorded a significant decrease in testosterone, LH and FSH values

as compared to -ve and +ve control groups. Otherwise, Panax – treated rats (+ve control) showed significant increase in testosterone, LH and FSH as evaluated to –ve control group.

Table 1. Effect of cypermethrin and panax on relative weight of sex organs.

Groups	Testes(g)	Seminal vesicles(g)	Prostate glands (g)
1	1.594±0.022 ^a	0.342±0.01 ^a	0.221±0.006 ^a
2	1.678±0.024 ^b	0.353±0.01 ^b	0.228±0.008 ^b
3	1.179±0.04 ^{abc}	0.202±0.009 ^{abc}	0.154±0.006 ^{abc}
4	1.39±0.038 ^{abc}	0.249±0.03 ^{abc}	0.162±0.006 ^{abc}

Data are presented as mean±SE. In each column, data followed by different superscript letters (^{a,b,c}) show significant difference at P<0.05.

Table 2. Effect of cypermethrin and panax on male sex hormones.

Groups	Testosterone (ng/L)	LH (ng/L)	FSH (ng/L)
1	3.264±0.07 ^a	5.61±0.09 ^a	5.978±0.005 ^a
2	4.38±0.104 ^{ab}	8.298±0.1 ^{ab}	6.762±0.04 ^{ab}
3	1.94±0.05 ^{abc}	1.65±0.06 ^{abc}	2.19±0.04 ^{abc}
4	2.364±0.09 ^{abc}	4.33±0.08 ^{abc}	5.24±0.02 ^{abc}

Data are presented as mean±SE. In each column, data followed by different superscript letters (^{a,b,c}) show significant difference at P<0.05.

Effect of cypermethrin and panax on sperm quality parameters

Table 3 revealed a significant decrease (P < 0.05) in sperm cell count and percentage of motile and live sperms, with a substantial rise of defective sperm cells in group-dosed cypermethrin as compared to all groups Fig.1. Co-treatment of Panax with cypermethrin improved these values but was still significantly different in the respective -ve and +ve groups.

Effect of cypermethrin and panax, on maternal index changes

Table 4 illustrates that pregnancy percent was recorded at 50% and 66.6% in the group dosed with cypermethrin and the group dosed with cypermethrin with Panax, respectively, in comparison to the -ve and +ve controls (100%). The percentages of

Table 3. Effect of cypermethrin and panax on sperm quality.

Groups	Sperm Count (x10 ⁶)	Sperm Motility (%)	Live Sperms (%)	Sperm Abnormalities
1	10.0±0.17 ^a	90.0±1.51 ^a	89.0±1.14 ^a	5.0±0.76 ^a
2	10.0±0.184 ^b	92.6±0.87 ^b	94±0.70 ^b	3.0±0.70 ^b
3	4.5±0.184 ^{abc}	40.0±3.53 ^{abc}	50.0±3.53 ^{abc}	35±3.53 ^{abc}
4	5.5±0.156 ^{abc}	60.0±3.53 ^{abc}	65.0±3.53 ^{abc}	25.0±3.53 ^{abc}

Data are presented as mean±SE. In each column, data followed by different superscript letters (^{a,b,c}) show significant difference at P<0.05.

Table 4. Effect of cypermethrin and panax on feoto-maternal index of male albino rats.

	Group 1	Group 2	Group 3	Group 4
Pregnancy	6 (100%)	6 (100%)	3 (50%)	4 (66.6%)
Implantation No. (litter)	8.5±0.42 ^a	9.1±0.47 ^b	6.3±0.33 ^{abc}	7.5±0.288 ^b
Resorption No. (litter)	0	0	2.7	0
Body weight (g)	5.16±0.12 ^a	5.45±0.1 ^b	4.1±0.09 ^{abc}	5.066±0.09 ^{bc}
Length weight (cm)	4.02±0.08 ^a	4.16±0.07 ^b	3.006±0.07 ^{abc}	4.04±0.06 ^c
A live feti (%)	99.2	99.5	90	95
A dead feti (%)	0.8	0.5	10	5
Malformati (%)	0	0	4.5	0
Placenta Weight (g)	0.75±0.01 ^a	0.77±0.013 ^b	0.71±0.01 ^{abc}	0.74±0.008

Data are presented as mean±SE. In each column, data followed by different superscript letters (^{a,b,c}) show significant difference at P<0.05.

abortion and resorption were seen in the cypermethrin-dosed groups. In addition, a significant decrease (P < 0.05) in the number of implants/litters was noticed in cypermethrin-treated rats as compared to all groups.

Effect of cypermethrin and panax on foetal parameters

Table 4 indicates a reduction in foetal body weight, length, and placental weight as well as in alive percent in the cypermethrin-treated group in relation to all groups, with a substantial raise in dead and malformed fetuses served (P< 0.05). Meanwhile, groups exposed to the cypermethrin combination with panax returned placental, foetal body weight, and length to be non-different as compared to the control group. Otherwise, the fetuses weight extensively decreased (P < 0.05) as compared to the -ve value.

Histopathological examination of male sex organs

As shown in Figure 1 the histopathological findings revealed marked degeneration of spermatogenic series, spermatogonia and sertoli cells in testes of cypermethrin treated rats as compared to all groups and showed regeneration in cypermethrin with Panax group.

As shown in Figure 2 seminal vesicles showed marked degeneration of seminal vesicle mucosal folds and lining epithelia in cypermethrin treated group, in cypermethrin with Panax group revealed restoring of only primary mucosal folds.

As shown in Figure 3 prostate glands showed marked atrophy of lining epithelial with only basal cells lining associated interstitial edema in cypermethrin treated group, while restoration of histological and secretory activities in group of cypermethrin with Panax.

DISCUSSION

In the current study, male fertility and reproductive performance were studied in adult orally intoxicated rats with 9.4 mg/kg. cypermethrin equivalent to 1/40 of estimated LD₅₀ (5 times/

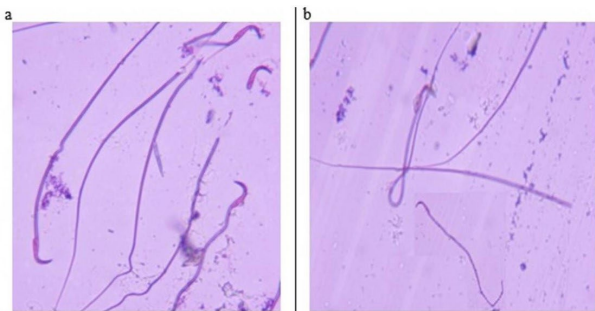


Fig. 1. Abnormalities of sperms; (a) show detached head, (b) show coiled tail.

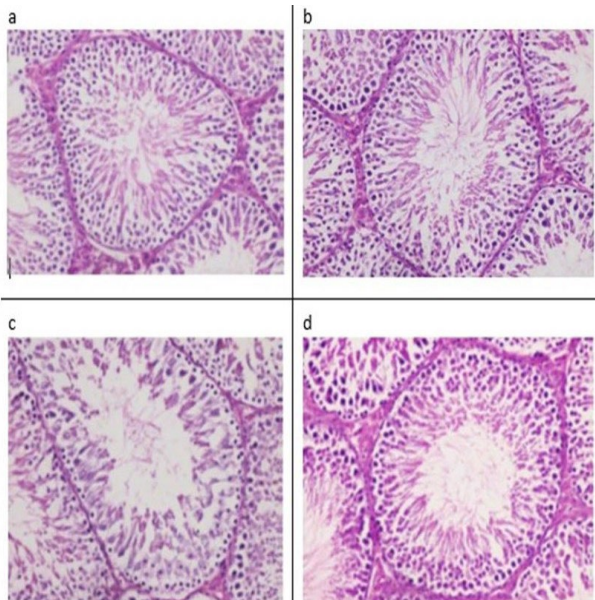


Fig. 2. Histopathology of testes showing (a) -ve control group Seminiferous tubules are normal criteria of spermatogenic series and normal Leydig cells appearance, (b) +ve control group with mild hyperplasia of Leydig cells. (c) cypermethrin group showing marked degeneration of spermatogenic series and persistence of spermatogonia and sertoli cells, (d) cypermethrin with Panax group showing regeneration of spermatogenic series, while the lumen harbor normal and degenerated spermatids, with mild proliferation of Leydig cells .H&EX400

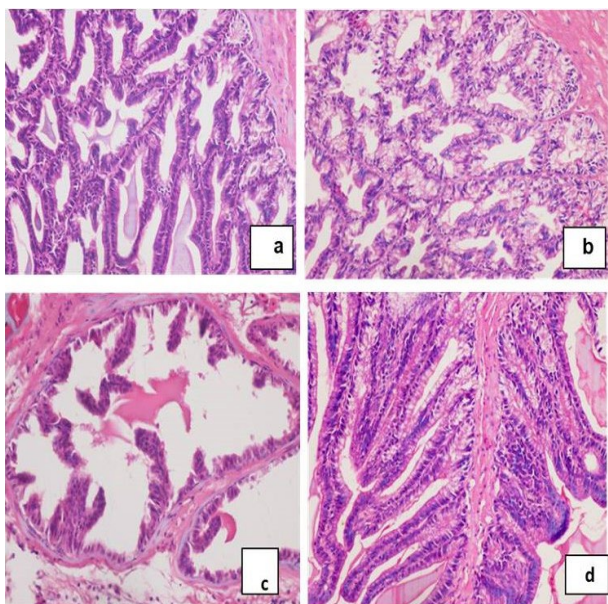


Fig. 3. Histopathology of seminal vesicle showing; (a) -ve control primary and secondary mucosal folds (with fusion at some areas) lined by simple columnar epithelium with basal elongated nuclei, some revealing secretory activities with clear apical cytoplasm. cytoplasm. (b) +ve control group showing normal histological criteria with evidence of increased secretory activities (clear cytoplasm) (c) cypermethrin group showing marked degeneration of seminal vesicle mucosal folds and lining epithelia (d)cypermethrin with Panax group showing restoring of only primary mucosal folds (double headed arrows) that have secretory activities in form of apical clear cytoplasm.

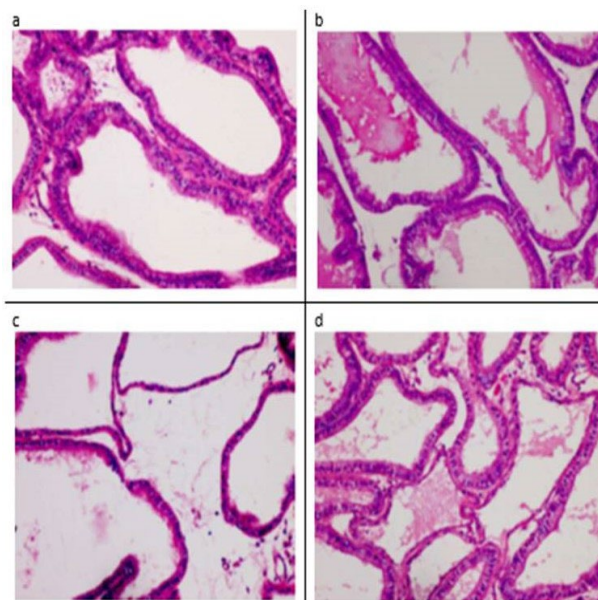


Fig 4. Histopathology of prostate (a) -ve control Prostate secretory alveoli showing; normal prostate secretory alveoli lined by simple cuboidal cells with features of secretory activities in form of eosinophilic granules at the apical border (b) +ve control group; in addition to presence of eosinophilic secretion in the lumen. (c) cypermethrin treated group showing marked atrophy of the lining epithelial with only basal cells lining associated by interstitial edema. (d) cypermethrin with Panax group showing restoration of histological and secretory activities of most tubules, while some still atrophied.

week) as well as to evaluate the amelioration of these adverse effects by using a panax-containing diet (0.1 g/kg diet/day) compared to a control throughout a 60-day period.

The king of all herbs, *Panax ginseng*, is frequently referred to as a potential supplement for enhancing overall health. Antioxidant and anti-inflammatory compounds are abundant in *Panax ginseng* (Leung and Wong, 2013) and androgenic activity (Sengupta and Dutta, 2022).

A typical synthetic pyrethroid with a broad range is cypermethrin. Due to its widespread application in several industries, it might be harmful to people, pets, and the environment. Data from the literature showed that cypermethrin damaged the male reproductive organs and reduced fertility in experimental animals (Hu et al., 2013; Mahdi et al., 2016; Chouabia et al., 2021; Kartagadda et al., 2021).

The acute oral LD₅₀ was calculated to be 374.633 ± 12.187 mg/kg b.wt in adult intoxicated male rats. This finding was compared with those won by Nagarjuna et al. (2009) estimated the LD₅₀ of cypermethrin as 205 mg/kg for rats orally. Oral LD₅₀ values of cypermethrin was to be 250mg/kg and 4132mg/kg in corn oil and in water respectively (Raj et al., 2013). The acute oral LD₅₀ of cypermethrin in albino rats was reported to be 416.98mg/kg, while it was 618.2mg/kg for formulated type II pyrethroids cypermethrin (Brijender and Saxena, 2017). These various findings may be attributed to different species, route of administration, vehicle, sex and climate beside regional variations (Uboh et al., 2011).

Clinical signs of cypermethrin were noticed, such as dyspnea, thick eye discharge, intermittent diarrhoea, and loss of body weight, during the first few days of intoxication. Later, animals adapted, and gradually, their body condition improved. Regarding mortality, the present observation revealed that mortalities occurred in 8% of cypermethrin poisoned rats only within the 1st week of treatment.

Clinical signs in cypermethrin of poisoned rats were manifested by dyspnea, thick eye discharge, intermittent diarrhea, tremors, loss of body weight during the first few days of intoxication. Later the rats gradually adapted their body condition improved. Similar findings were recorded by El-sheshtway et al. (2016); Ramon-Yusuf et al. (2017); Simon et al. (2018) and Priyanka et al. (2023).

Rats either male or female exposed to cypermethrin at different doses for different periods, showed varying degree of mild to moderate toxic symptoms and behavioral changes with the

appearance of intermittent diarrhea, decreased feed intake, thick eye discharge, dyspnea, ataxia, salivation after displaying signs of incoordination and tremors, in addition, there were signs of CNS stimulation and followed by depression (Islam and Hoque 2015). Contrary, no observable effects attributable to cypermethrin were detected by Sangha and Kamalpreet (2011), however, Kašuba et al. (2022) showed no signs of systemic toxicity or death in treated rats all time of experiment.

These adverse signs of cypermethrin could be attributed to its neurotoxic action, which lead to inhibition of acetylcholinesterase (ACh) resulting in the retention of ACh in synaptic gaps of neurons, and to their interaction with Na channels inducing neurons depolarization (Idris et al., 2012).

The obtained data showed that cypermethrin induced negative effects in male reproduction in all cypermethrin poisoned rats, whereas rats receiving cypermethrin with Panax showed a lesser trend. concerning the weight of sex organs, there was significant decrease in the relative weights of testes, seminal vesicles, and prostate glands in cypermethrin poisoned rats compared to controls. These results were in accordance with those reported by Sharma et al. (2013); El-Sheshtway et al. (2016); Alaa-Eldin et al. (2017); Murad et al. (2020) and Katragadda et al. (2021). In contrast, cypermethrin had no change in the weight of testes, seminal vesicles, or prostate glands in rats (Fang et al. 2013). Also, there was increase in the weight of testes in cypermethrin-treated rats (Simon et al. 2018).

There were direct destructive impact of cypermethrin on testes, seminal vesicles, and prostate gland tissues may be the cause of the observed reduction in weight of testes and accessory sex organs following cypermethrin treatment (Elbetieha et al., 2001), as confirmed by the histopathological findings. In testicles obtained from cypermethrin poisoned rats, showed marked degeneration of spermatogenic series and persistence of spermatogenesis and Sertoli cells (Fig. 1), the observed degeneration in the current study was consistent with previous studies of rat testis and accessory glands (Joshi et al., 2011; Ahmad et al., 2012; Alaa-Eldin et al., 2017; Murad et al., 2020). The weight of the reproductive organs decreases due to the circulating amount of androgen, which affects the structural and functional integrity of the reproductive organs. Cypermethrin caused a significant drop in testosterone, LH, and FSH levels, which may result in a decrease in organ weights. Similarly, cypermethrin may exert its excess effects on the weights of sex organs through its affinity for male sex hormones (Mathur and Song, 1995).

Results for the hormonal analysis agreed with those reported by Joshi et al. (2011); Sharma et al. (2013), El-Sheshtawy et al. (2019) and Ubah et al. (2021). The recorded depression in testosterone, LH, and FSH levels may be due to cypermethrin's direct cytotoxicity on testicular tissues and cells (Cremonese et al., 2017), which was confirmed by microscopic examination. The drop in hormone levels and said that it may be due to either cypermethrin's direct impact on the pathway in the testis that produces androgen or its indirect impact on the hypothalamus and anterior pituitary gland, which may influence the testes and sexual function (Rajawat et al. 2014).

An additional testicular target for pyrethroids was suggested as a potential reason for the decrease in testosterone, FSH, and LH levels. The hypothalamus-pituitary axis may be affected by the pyrethroid as well. A reduction in LH may potentially be a contributing factor to the low level of testosterone since LH stimulates the Leydig cells to generate testosterone. Cypermethrin's potential hormonal action has a variety of impacts on the endocrine system (Joshi et al. 2011).

Otherwise, panax-treated rats recorded an improvement in hormonal values as compared to cypermethrin poisoned rats recorded an increment in their values in relation to the controls, which could be related to the stimulant effect of *Panax ginseng* on male sex hormones, thus increasing sexual male function (Lee et al., 2017). In addition, He Lin et al. (2021) reported that *Panax ginseng* may be utilized as a natural androgen supplement to raise levels of testosterone and luteinizing hormone and encour-

age the pituitary gland's release of gonadotropins.

Sperm quality parameters revealed a significant decrease in the count sperm cells, percentage of viable, motile sperm in all cypermethrin-treated rats, either with or without panax in comparison to the control. However, however a significant increase in sperm cell abnormalities. Alike results were described in rats by Sharma et al. (2013); El-Sheshtway et al. (2016); Katragadda et al. (2021) and Mahdi et al. (2016) in rabbits.

The decrease in sperm count was caused by the stoppage of spermatogenesis and the persistence of spermatogonia. Insecticide buildup in the testicular tissue may have had a negative impact on the Sertoli cell population, impairing spermatogenesis and lowering the number of sperm cells Cremonese et al. (2017). Following pyrethroid treatment, epididymal sperm count, motility, viability, and morphology were considerably affected; the decrease in sperm count may be caused by a negative effect of pyrethroids on spermatogenesis (Ben Slima et al., 2013).

In addition, Madhubabu and Yenugu (2017) reported that spermatogenesis, sperm production, and sperm function are all impacted by pyrethroid toxicity, which acts at the molecular level. The information in this study also shows that adult rats exposed to cypermethrin over an extended period saw a considerable reduction in testicular weight, a marker of diminished spermatogenesis. The decrease in fertility is probably due to the decline in spermatogenesis.

The number of females who became pregnant after mating with the treated males, the number of implantations, and the number of resorptions in females who were impregnated by cypermethrin poisoned rats were all significantly lower in this study's results when compared to the -ve and +ve controls. These findings coincided with those of Ahmad et al. (2012) in rats exposed to low doses of different pesticides. These effects may be attributed to poor development of fertilized ova due to an alteration in sperm quality, whereas the increased sperm cell abnormalities may have left them incapable of fertility (Rao et al., 1996).

Furthermore, the current experiment has shown that exposure of adult male rats to cypermethrin either alone or with Panax induced hazardous effects on foetal values such as decreased placental, foetal body weights and lengths, and decreased alive fetuses percent, while results displayed a significant increase in dead and malformed foetal percent in the form of hematoma and stunt tail in comparison to -ve and +ve controls; similar results were reported by Ahmad et al. (2012) in rats. These results could be ascribed to sperm cell abnormalities, which are usually taken as characteristic criteria and applied to monitoring the mutagenic potential of many chemicals. Sharma et al. (2013)

In addition, the current study showed that Panax co-treatment reverted the poisonous effect of cypermethrin on male fertility and reproductive parameters, in which group 4 dosed cypermethrin and fed on a Panax-containing diet recorded a significant improvement in all altered parameters as compared to group administrated cypermethrin alone and recorded no change than -ve and +ve controls in placental weight, foetal body weight, and length, even though all fertility parameters still significantly differed in comparison to -ve and +ve controls. Also, microscopic examination of the testicle and accessory glands obtained from this group showed a nearly normal architecture.

Otherwise, the effective role of Panax on the several toxic materials was in accordance with results, as noticed by Aslan et al. (2021) on cisplatin-induced testicular damage in rats; He lin et al. (2021) in the treatment of androgen-deficient rats (via metabolomic and gut microbiota), the authors attributed the improvement of reproductive functions of male rats by Panax to its antioxidant and androgenic activities.

The +ve control group (Panax-treated rats) showed similar results to those of negative controls and recorded the best results in some values of percent motile and live sperm decline in sperm cell abnormalities and a significant increase in testes, LH, and FSH levels as compared to -ve controls. These results were supported by previous findings that found that Panax significantly increased testosterone levels and spermatozoa parameters in

animal models (Xu et al., 2014; Khouri et al., 2015; Park et al., 2016; Kee et al., 2018).

Clinical in vivo and in vitro studies' outcomes have demonstrated recently that *Panax ginseng* and its constituent ingredients have a hormonal-like action (Tian et al., 2020). The *Panax ginseng* polysaccharide, which also increases the expression level of androgen receptor mRNA, boosts testosterone, and assures the appropriate function of androgen, may be the cause of the elevated hormonal values (Park et al., 2017).

Panax's effects can be linked to its impact on the hypothalamus-pituitary-adrenal cortical axis, which boosts vigor and fortitude, restores and improves the body's immunological response, encourages lifespan, metabolism, and the creation of healthy cells (Kopalli et al., 2017). Ginseng is a cyclic adenosine monophosphate-responsive element modulator, which helps to improve testicular issues, sperm quality, and sperm motility, according to a number of preclinical investigations (Lee et al., 2017; Kee et al., 2018; He lin et al., 2021). Ginsenosides, the main active ingredients in ginseng, may impact estrogen and androgen activity as well as behave as an aphrodisiac, according to recent reviews (Park et al., 2017).

CONCLUSION

The results of the present study show that cypermethrin directly affects male fertility and reproduction and that these harmful effects may be prevented or mitigated by employing *Panax ginseng* as a detoxifying, antioxidant, and androgenic agent.

CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

REFERENCES

- Ahmad, L., Khan, A., Khan, M.Z., Hussain, I., Mahmood, F., Sleemi, M.K., Abdullah, I., 2012. Toxicopathological effects of cypermethrin upon male reproductive system in rabbits. *Pesticide Biochem. Physiol.* 103, 194-201.
- Alaa-Eldin, E.A., El-Shafei, D.A., Abouhashem, N.S., 2017. Individual and combined effect of chlorpyrifos and cypermethrin on reproductive system of adult male albino rats. *Environ Sci. Pollut. Res.* 24, 1532-1543.
- Andersson, A.M., Jørgensen, N., Main, K.M., Toppari, J., Meyts, E.R.D., Lefers, H., Skakkebaek, N., 2008. Adverse trends in male reproductive health: we may have reached a crucial 'tipping point'. *Inter. J. Androl.* 31, 74-80.
- Aslan, E., Kumalar, K., Güzel, H., Demirel, H.H., Çelik, S., Pektaş, M.B., 2021. Effects of *Panax ginseng* on cisplatin-induced testicular damage of rats. *Anatolian J. Botany* 5, 37-43.
- Assayed, M.E., Salem, H.A., Khalaf, A.A., 2010. Protective effects of garlic extract and vitamin C against cypermethrin reproductive toxicity in male rats. *Res. J. Vet. Sci.* 3, 13-27.
- Attele, A.S., Wu, J.A., Yuan, C.S., 1999. Ginseng pharmacology: multiple constituents and multiple actions. *Biochem. Pharmacol.* 58, 1685-1693.
- Bancroft, J.D., Cook, H.C., Stirling, R.W., 1994. Manual of histological techniques and their diagnostic application. In *Manual of histological techniques and their diagnostic application* pp. 457-457.
- Bearden, H.J., Fuquay, J.W., 1984. Applied animal reproduction. Reston Publishing Company, Inc., Reston, Virginia, pp.158-165.
- Ben Slima, A., Ali, M.B., Barkallah, M., Traore, A.I., Boudawara, T., Allouche, N., Gdoura, R., 2013. Antioxidant properties of Pelargonium graveolens L'Her essential oil on the reproductive damage induced by deltamethrin in mice as compared to alpha-tocopherol. *Lipids Health Dis.* 12, 1-9.
- Blandau, R.J., Jordan, E.S., 1941. A technique for the artificial insemination of the white rat. *J. Lab. Clin. Med.* 26, 1361-1363.
- Brijender, B., Saxena, P.N., 2017. Estimation of Median Lethal Dose of Cypermethrin and Betacyfluthrin. *Inter. J. Toxicol. Pharmacol. Res.* 9, 194-198.
- Chouabia, A., Djemli, S., Abdennour, C., Mallem, L., Kahalerras, L., Arkoub, F. Z., Tahraoui, A., 2021. Protective effect of *Salvia officinalis* against cypermethrin induced reprotoxicity in male Wistar rats. *Pharmacognosy Journal* 13, 6.
- Cook, M.J., Fairweather, F.A., 1968. Methods used in teratogenic testing. *Laboratory Animals* 2, 2, 219-228.
- Cremonese, C., Piccoli, C., Pasqualotto, F., Clapauch, R., Koifman, R. J., Koifman, S., Freire, C., 2017. Occupational exposure to pesticides, reproductive hormone levels and sperm quality in young Brazilian men. *Reproductive Toxicology* 67, 174-185.
- Elbetieha, A., Da'As, S.I., Khamas, W., Darmani, H., 2001. Evaluation of the toxic potentials of cypermethrin pesticide on some reproductive and fertility parameters in the male rats. *Archives of Environmental Contamination and Toxicology* 41, 522-528.
- El-Sheshtawy, S.M., El-Gobary, G.I.A., Omar, N.A., Shawky, N.A., 2019. Ameliorating the Toxic Effects of Cypermethrin by Sesame oil in Male Rabbits. *Slov Vet Res.* 56, 51-59.
- El-Sheshtawy, S.M., El-Gebaly, L.S., Ebaid, M.R.B., El-Sheikh, N.I., 2016. Toxic effects of cypermethrin on male fertility and some hepatic biochemical parameters in male albino rats Egypt *J Chem Environ Health* 2, 66-77.
- Li, Y.F., Pan, C., Hu, J.X., Li, J., Xu, L.C., 2013. Effects of Cypermethrin on Male Reproductive System in Adult Rats. *Biomed Environ Sci.* 26, 3, 201-208.
- Goodman, M.F., 1992. Canine ovulation timing. *Probl. Vet. Med.* 4, 433-444.
- Harlod, E., James, A., Doglas, C., Cherly, L., Franklin, M., Glenn, H., 2003. *The Merck Veterinary Manual*, Merck and Co., Inc., Rahaway, N.J., U.S.A., 9th ed.
- Hashem, H.E., Abd El-Haleem, M.R., Abass, M.A., 2015. Epithelial and stromal alterations in prostate after cypermethrin administration in adult albino rats (histological and biochemical study). *Tissue and Cell* 47, 366-372.
- He Lin, Jiarui Zhao, Zhongying Liu, Zhiqiang Liu, Lin, Z., 2021. Efficacy of *Panax ginseng* supplementation on androgen deficiency rats via metabolomics and gut microbiota. *Journal of Functional Foods* 87, 104810.
- Hu, Jin-xia., Yan-fang Li., Jing Li., Chen Pan; Zhen He., Hong-yan Dong., Xu, L., 2013. Toxic effects of cypermethrin on the male reproductive system: with emphasis on the androgen receptor. *Journal Applied Toxicology* 33, 576-585
- Idris, S.B., Ambali, S.F., Ayo, J.O., 2012. Cytotoxicity of chlopyrifos and cypermethrin: The ameliorative effects of antioxidants. *African Journal of Biotechnology* 11, 16461-16467.
- Islam, M., Hoque, S., Mizanul, M., 2015. Clinico-Haematological and Histopathological Features of the Swiss Albino Mice *Mus musculus* L. in Response to Chronic Cypermethrin Exposure. *Sch. Acad. J. Biosci.* 3, 421-428.
- Jackson, H., Jones, A., 1966. Antifertility compounds in male and female. Thomas, Springfield, Illinois.
- Joshi, S.C., Bansal, B., Jasuja, N.D., 2011. Evaluation of reproductive and developmental toxicity of cypermethrin in male albino rats. *Toxicological & Environmental Chemistry* 93, 593-602.
- Kašuba, V., Blanka T., Ana, L., Anja, K., Nevenka, K., Vedran, M., Mirta, M., Alica, P., Davor Ž., Suzana, Ž., 2022. Evaluation of Toxic Effects Induced by Sub-Acute Exposure to Low Doses of Cypermethrin in Adult Male Rats *Toxics* 10, 717.
- Katragadda, V., Adem, M., Mohammad, R.A., Sri Bhayam, S., Battini, K., 2021. Testosterone recuperates deteriorated male fertility in cypermethrin intoxicated rats. *Toxicological Research* 37, 125-134.
- Kee, J.Y., Han, Y.H., Mun, J.G., Um, J.Y., Hong, S.H., 2018. Pharmacological effect of prohibited combination pair *Panax ginseng* and *Veratrum nigrum* on colorectal metastasis in vitro and in vivo. *Journal of Ethnopharmacology* 220, 177-187.
- Khouri, Nabil A., Daradka, Haytham M., Allouh, Mohammed Z., Alkofahi, Ahmad S., 2015. Comparative effects of *Orchis anatolica* vs. the red Korean *Panax ginseng* treatments on testicular structure and function of adult male mice. *Journal of Complementary and Integrative Medicine* 12, 33-41.
- Kjeldsen, L.S., Ghisari, M., Bonefeld-Jørgensen, E.C., 2013. Currently used pesticides and their mixtures affect the function of sex hormone receptors and aromatase enzyme activity. *Toxicology and Applied Pharmacology* 272, 453-464.
- Kopalli, S.R., Cha, K.M., Lee, S.H., Ryu, J.H., Hwang, S.Y., Jeong, M. S., Kim, S.K., 2017. Pectinase-treated *Panax ginseng* protects against chronic intermittent heat stress-induced testicular damage by modulating hormonal and spermatogenesis-related molecular expression rats. *Journal of Ginseng Research* 41, 578-588.
- Lee, Y.M., Yoon, H., Park, H.M., Song, B.C., Yeum, K.J. 2017. Implications of red *Panax ginseng* in oxidative stress associated chronic diseases. *Journal of Ginseng Research* 41, 113-119.
- Leung, K.W., Wong, A.S., 2013. Ginseng and male reproductive function. *Spermatogenesis* 3, e26391.
- Li, Y.F., Chen, P.A.N., Hu, J.X., Jing, L.I., Xu, L.C., 2013. Effects of cyperme-

- thrin on male reproductive system in adult rats. *Biomedical and Environmental Sciences* 26, 201-208.
- Lifeng, T., Shoulin, W., Junmin, J., Xuezhao, S., Yannan, L., Qianli, W., Longsheng, C., 2006. Effects of fenvalerate exposure on semen quality among occupational workers. *Contraception* 73, 92-96.
- Madhubabu, G., Yenugu, S., 2017. Allethrin toxicity causes reproductive dysfunction in male rats. *Environmental Toxicology* 32, 1701-1710.
- Mahdi, Y.S., Falih, I.B., Zaid, N.Y., 2016. Toxicological effects of cypermethrin on sperm morphology in male rabbit. *Int. J. Adv. Res. Biol. Sci.* 3, 46-51.
- Mathur, V.K., Song, F.M., 1995. The Dynamics of Regional Population and Employment Growth. *Review of Urban and Regional Development Studies* 7, 70-88.
- Murad, H.M., Abdulameer, S.A., Asker Aljuboory, D.S., Neamah, D.A., Maktoof, A.H., 2020. Defensive Effects of Breberine against Cypermethrin Induced Male Reproductive System Toxicity in Rabbits. *Systematic Reviews in Pharmacy* 11, 9.
- Nagarjuna. A., Doss. P. 2009. Acute oral toxicity and histopathological studies of cypermethrin in rats. *Indian Journal of Animal Res.* 43, 235-240.
- Park, H.J., Choe, S., Park, N.C., 2016. Effects of Korean red ginseng on semen parameters in male infertility patients: a randomized, placebo-controlled, double-blind clinical study. *Chinese Journal of Integrative Medicine* 22, 490-495.
- Park, J., Bui, P.T., Song, H., Kim, S.K., Rhee, D.K., Kim, E.Y., Lee, Y. J. 2017. Ginseng on nuclear hormone receptors. *The American Journal of Chinese Medicine* 45, 6, 1147-1156.
- Priyanka, S., Anshu, R., Khushboo, C., Preety M., Sudesh, R., 2023. Systematic Toxicity of Cypermethrin and Alterations in Behavior of Albino Rats. *ACS Omega* 8, 14766-14773.
- Rajawat, N. K., Soni, I., Mathur, P., Gupta, D., 2014. Cyfluthrin-induced toxicity on testes of Swiss albino mice. *Int. J. Curr. Microbiol. Sci.*, 3, 334-343.
- Raj J., Mohineesh, C., Tirath, D., Monika P., Anupuma, R., 2013. Determination of Median Lethal Dose of Combination of Endosulfan and Cypermethrin in Wistar Rat. *Toxicology International* 20, 1.
- Ramon-Yusuf, S.B., Aliu, Y.O., Salawu, O.A., Chahoud, I., Ambali, S.F., 2017. Maternal and foetal toxicity induced by exposure to mixture of dimethoate and cypermethrin in albino rats. *Journal of Toxicology and Environmental Health Sciences* 9, 6, 59-65.
- Rao, M.V., Thakoz, F., Shineg, G., Sharh, K.P., Roy, G.K., 1996. Contraceptive efficacy of testosterone enanthate in males. *J. of Animal Morphology and Physiology* 43, 15-21.
- Reddy, K., Bordekar, A.D., 1999. Spectrophotometric analysis of resazurin reduction test and semen quality in men. *Indian Journal of Experimental of Biology* 37, 782-786.
- Sakuma, Y., 2009. Gonadal steroid action and brain sex differentiation in the rat. *J. Neuroendocrinol.* 21, 410-414.
- Sangha, G.k., Kamalpreet, K. 2011. Cypermethrin induced changes in biochemical constituents of plasma of female albino rats, *Indian J. Anim. Res.* 45, 186-191.
- Sengupta, P., Dutta, S., 2022. *Panax ginseng* as reproductive medicine in male infertility: with a brief focus on herb-drug interaction. *Chemical Biology Letters* 9, 1, 279-279.
- Sharma, P., Huq, A.U., Singh, R., 2013. Cypermethrin induced reproductive toxicity in male. *Journal of Environmental Biology* 34, 857-862.
- Sharma, P., Khan, I.A., Singh, R. 2018. Curcumin and quercetin ameliorated cypermethrin and deltamethrin-induced reproductive system impairment in male wistar rats by upregulating the activity of pituitary-gonadal hormones and steroidogenic enzymes. *International Journal of Fertility & Sterility* 12, 72.
- Simon, U., David, O., Peter, R., Joseph, R., Ijeoma, C., Celestine, N., 2018. Pathological effects of cypermethrin on the testes and accessory sexual glands of Yankasa rams. *Arch. Pathol. Clin. Res.* 2, 6-12.
- Swan, S.H., Kruse, R.L., Liu, F., Barr, D.B., Drobnis, E.Z., Redmon, J.B., 2003. Semen quality in relation to biomarkers of pesticide exposure. *Environmental Health Perspectives* 111, 1478-1484.
- Tian M., Li Lin-Nan, ZhenG Rui-Rong, Yang Li, Wang Zheng-Tao, 2020. Advances on hormone-like activity of *Panax ginseng* and ginsenosides. *Chinese Journal of Natural Medicines* 18, 526-535.
- Travison, T.G., Shackelton, R., Araujo, A.B., Hall, S.A., Williams, R.E., Clark, R.V., O'Donnell, A.B, McKinlay, J.B., 2008. The natural history of symptomatic androgen deficiency in men: onset, progression, and spontaneous remission. *J. Am. Geriatric Soc.* 56, 831-839.
- Ubah, S.A., Agbonu, O.A., Columbus, P.K., Abah, K.O., Chibuogwu, I.C., Abalaka, S.E., Ajayi, I.E., 2021. Effects of date fruit (*Phoenix dactylifera*) on sperm cell morphology and reproductive hormonal profiles in cypermethrin-induced male infertility in Wistar rats. *Scientific African* 11, 713.
- Uboh, F.E. Asuquo, E.N., Eteng, M.U., Akpanyung, E.O., 2011. Endosulfan induces renal toxicity independent of the route exposure in rats. *Am. J. Biochem. Mol. Biol.* 1, 359.
- Vasudha, K., Meghapriya, A., Kishori, B., 2018. Recovery of male reproductive health in cypermethrin-exposed rats by testosterone. *Int. J. Res. Anal. Rev.* 5, 311-317.
- Wand, M.H., Colin, G.R., 1998. *Fundamentals of Toxicologic Pathology*. Academic Press, New York, Sydney, Tokyo, pp. 478.
- Xu, Y., Ding, J., Ma, X.P., Ma, Y.H., Liu, Z.Q., Lin, N., 2014. Treatment with *Panax ginseng* antagonizes the estrogen decline in ovariectomized mice. *International journal of molecular sciences* 15, 7827-7840.
- Yilmaz, S. N., Çömelekoğlu, Ü., Coşkun, B., Ballı, E., Özge, A., 2008. Effects of cypermethrin on isolated frog sciatic nerve: An ultrastructural study. *Turkish Journal of Medical Sciences* 38, 121-125.