Risk of antimicrobial and multidrug resistance on Avian Pathogenic Escherichia coli in public health

Ummi Rahayu¹, Freshinta J. Wibisono², Wiwiek Tyasningsih^{3,4}, Mustofa H. Effendi^{4,5,6*}, Budiastuti Budiastuti⁷, Budiarto Budiarto⁵, Dian A. Permatasari^{4,5}, Aswin R. Khairullah⁸, Saifur Rehman⁹, John Y.H. Tang⁶, Riza Z. Ahmad⁸, Bima P. Pratama¹⁰, Ikechukwu B. Moses¹¹, Dea A.A. Kurniasih¹²

²Faculty of Veterinary Medicine, Universitas Wijaya Kusuma Surabaya, Jl. Dukuh Kupang XXV No.54, Dukuh Kupang, Dukuh Pakis, Surabaya 60225, East Java, Indonesia.

³Division of Veterinary Microbiology, Faculty of Veterinary Medicine, Universitas Airlangga, Kampus C Mulyorejo, Jl. Dr. Ir. H. Soekarno, Surabaya, East Java, 60115, Indonesia.

⁴Research Group on Antimicrobial Resistance, Faculty of Veterinary Medicine, Universitas Airlangga, Kampus C Mulyorejo, Jl. Dr. Ir. H. Soekarno, Surabaya, East Java, 60115, Indonesia.

⁵Division of Veterinary Public Health, Faculty of Veterinary Medicine, Universitas Airlangga, Kampus C Mulyorejo, Jl. Dr. Ir. H. Soekarno, Surabaya, East Java, 60115, Indonesia.

⁶School of Food Industry, Faculty of Bioresources, and Food Industry, Universiti Sultan Zainal Abidin (Besut Campus), Besut 22200, Malaysia.

⁷Study Program of Pharmacy Science, Faculty of Health Science, Universitas Muhammadiyah Surabaya, Jl. Raya Sutorejo No.59, Dukuh Sutorejo, Mulyorejo, Surabaya 60113, East Java, Indonesia.

[®]Research Center for Veterinary Science, National Research and Innovation Agency (BRIN), Jl. Raya Bogor Km. 46 Cibinong, Bogor, West Java, 16911, Indonesia.

²Department of Pathobiology, Faculty of Veterinary and Animal Sciences, Gomal University, RV9W+GVJ, Indus HWY, Dera Ismail Khan 27000, Pakistan.

¹⁰Research Center for Agroindustry, National Research and Innovation Agency (BRIN), Jl. Raya Puspiptek 60, South Tangerang, West Java, 15310, Indonesia.

"Department of Applied Microbiology, Faculty of Science, Ebonyi State University. Abakaliki 480211, Nigeria.

12 Research Center for Public Health and Nutrition, National Research and Innovation Agency (BRIN), Jl. Raya Bogor Km. 46 Cibinong, Bogor, West Java, 16911, Indonesia.

ARTICLE INFO

Recieved: 26 April 2025

Accepted: 27 May 2025

*Correspondence:

Corresponding author: Mustofa H. Effendi E-mail address: mhelmieffendi@gmail.com

Keywords:

AMR, APEC, MDR, Poultry, Public health.

Introduction

Escherichia coli is a pathogenic bacterium commonly found in poultry raised for consumption (such as quail, chicken, duck, and turkey), which causes the significant disease colibacillosis (Hidanah *et al.*, 2023; Li *et al.*, 2016). This disease is caused by Avian Pathogenic *Escherichia coli* (APEC), a highly virulent strain capable of causing severe illnesses in poultry (Ibrahim, 2019; Wibisono *et al.*, 2022). APEC's transmission through the food chain presents potential risks to human health, particularly via the consumption of contaminated poultry products (Amir *et al.*, 2017; Saraiva *et al.*, 2022).

Poultry such as chickens, turkeys, ducks, and quails are raised in large populations and are hig*hly* susceptible to bacterial infections (Abd El-Ghany, 2019). These infections can significantly affect poultry populations, leading to the widespread use of antibiotics for treatment and prevention, which contributes to the development of antibiotic resistance and the potential for harmful residues in the environment and human food supply (Nwobodo *et al.*, 2022). Pathogenic *E. coli* in consumption poultry possess numerous virulence factors that contribute to their infection and pathogenicity, allowing them to adapt and thrive (Kalule *et al.*, 2018).

The major concern with *E. coli* infections is antibiotic resistance, often manifesting as antimicrobial resistance (AMR) and multidrug resistance (MDR). The use of antibiotics in animal agriculture has led to the emergence of resistant strains of pathogenic bacteria (Xu *et al.*, 2022). Aarestrup (2012) notes that global antibiotic use in animals is nearly double that in humans. Annually, about 63.1 ± 1.5 tons of antibiotics are used in livestock production (Van Boeckel *et al.*, 2015), with more than 80% of

animals raised for food purposes receiving antibiotics. By 2030, global antibiotic use in food animals is projected to increase by 67%, reaching approximately 105,500 tons (Van Boeckel *et al.*, 2015).

Residues from antibiotics in animal products such as quail meat and eggs are higher than those in chicken eggs (8.4%) and duck eggs (2.66%), with the aminoglycoside group showing 9.15% in quail eggs (Anggita *et al.*, 2021). People who consume poultry products, such as meat, eggs, and dairy, are at risk of developing resistance to specific antimicrobials due to antibiotic residues that enter the food chain (Abreu *et al.*, 2023; Roess *et al.*, 2013). This resistance can reduce the effectiveness of antibiotics used to treat human diseases.

The widespread use of antibiotics can also result in the emergence of antibiotic-resistant bacterial strains, with the potential for resistant genes to be transferred to other pathogenic or non-pathogenic bacteria (Peterson and Kaur, 2018; Widodo *et al.*, 2022). Consistent with research by Khairullah *et al.* (2024), resistant bacteria can become pathogenic and cause negative health impacts. These resistance genes can be transferred to other bacteria, exacerbating the problem. This review aimed to assess the risks of AMR and MDR APEC in consumption poultry, focusing on dangerous virulence genes, residual contamination, and environmental health impacts.

Pathogenic Escherichia coli bacteria

Escherichia coli is a rod-shaped, Gram-negative bacterium that is facultatively anaerobic and belongs to the Enterobacteriaceae family. It naturally resides in the digestive tracts of both animals and humans (Santos

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made. ISSN: 2090-6277/2090-6269/ © 2011-2025 Journal of Advanced Veterinary Research. All rights reserved.

ABSTRACT

Escherichia coli is a harmful bacterium commonly found in poultry species like chickens, ducks, quails, and turkeys, which can lead to colibacillosis. This illness is caused by Avian Pathogenic *Escherichia coli* (APEC), a highly virulent strain that poses a risk of transmission through the food chain, endangering human health. Managing infections in poultry frequently involves the widespread use of antibiotics, which promotes the development of antimicrobial resistance (AMR) and multidrug resistance (MDR). The global use of antibiotics in livestock is a significant issue, with an estimated 63,000 tons used annually, expected to rise to 105,500 tons by 2030. Antibiotic residues found in poultry products, such as meat and eggs, can enter the human food supply, contributing to antibiotic resistance and diminishing the effectiveness of treatments for human infections. The transmission of APEC occurs through horizontal pathways, including direct contact between poultry via feces and the environment, and vertical transmission via eggs from infected breeders. Additionally, APEC can cause allergic reactions, cancer, reproductive issues, and toxicity in humans. The consequences of antimicrobial resistance include higher rates of illness, death, and healthcare costs, with the potential for further escalation.

¹Master Program of Veterinary Disease and Public Health Science, Faculty of Veterinary Medicine, Universitas Airlangga, Kampus C Mulyorejo, Jl. Dr. Ir. H. Soekarno, Surabaya, East Java, 60115, Indonesia.

et al., 2020). While *E. coli* itself is generally harmless, certain serotypes can cause diseases, such as colibacillosis in poultry, which is caused by Avian Pathogenic *Escherichia coli* (APEC) with serogroups including O1, O2, and O78 (Kathayat *et al.*, 2021; Watts and Wigley, 2024). Some *E. coli* strains, like the EHEC O157:H7, produce shiga toxins and can lead to zoonotic diseases (Rahman *et al.*, 2020). In poultry, *E. coli* can cause a variety of infections, including yolk sac infection, omphalitis, cellulitis, swollen head syndrome, coligranuloma, and colibacillosis, which are associated with high mortality rates (Islam *et al.*, 2021).

Pathogenic *E. coli* is classified into two main categories: Extraintestinal Pathogenic *Escherichia coli* (ExPEC) and Diarrheagenic *Escherichia coli* (DEC) (Vanstokstraeten *et al.*, 2022; Pokharel *et al.*, 2023). ExPEC includes Uropathogenic *E. coli* (UPEC), Neonatal Meningitis-Associated *E. coli* (NMEC), APEC, and Septicemic *E. coli* (SEPEC). DEC includes Enterotoxigenic *E. coli* (ETEC), Enteroinvasive *E. coli* (EIEC), Enteroaggregative *E. coli* (EAEC), Enteropathogenic *E. coli* (EPEC), Shiga Toxin-producing *E. coli* (STEC), Diffusely Adherent *E. coli* (DAEC), and Adherent-Invasive *E. coli* (AIEC). According to Lim *et al.* (2020), strains such as EPEC, ETEC, EIEC, and EHEC are known to cause foodborne illnesses.

E. coli is primarily transmitted through horizontal routes, including direct contact with infected poultry or through the respiratory tract. The bacteria can survive in feces for weeks, enabling prolonged transmission. Vertical transmission occurs through contaminated eggshells (Joseph *et*

al., 2023). Additionally, the bacteria can be spread via the digestive tract, where they can infect and multiply, leading to further transmission.

The rising threat of AMR and MDR in poultry products for human consumption

The rising incidence of Antimicrobial Resistance (AMR) and Multidrug Resistance (MDR) in poultry is a global concern, as shown in Table 1. In particular, in Indonesia, there is a widespread misuse of antibiotics by the public and farmers, who often administer antibiotics without proper veterinary supervision and fail to calculate the correct dosage (Siahaan *et al.*, 2022). According to Salam *et al.* (2023), antibiotic resistance is a major global issue and has the potential to become a leading cause of death in the next 50 years. Ahmed *et al.* (2024) report that drug-resistant microorganisms are responsible for over 1.2 million deaths annually, with the death toll projected to rise to 10 million by 2050. Furthermore, the World Bank estimates that AMR will increase healthcare costs by USD 1 trillion by 2050. *E. coli* is currently one of the main contributors to AMR, accounting for 43% of all cases (Karpov *et al.*, 2024; Murray *et al.*, 2022).

Antibiotic-resistant bacteria can lead to serious health complications, treatment failures, and even death (Mancuso *et al.*, 2021). Various incidents of AMR in poultry have been documented, including a study by Farghaly *et al.* (2017) where *E. coli* isolates exhibited resistance to amoxi-

Table 1. AMR and MDR data on *Escherichia coli* in the world.

Types of entibioties	Country	Sampla	Sauraa
Types of antibiotics	Country	Sample	Source
Erythromycin (98.7%) Enrofloxacin (96.1%) Tetracycline (95.2%) Doxycycline (93.9%) Lincomycin (90.0%) Streptomycin (90.0%) Ampicillin (87.8%) Sulfamethoxazole/trimethoprim (84.3%) Amoxicillin (81.7%) Cephalotin (78.7%) Florfenicol (69.6%) Amikacin (67.4%) Gentamicin (62.2%) Lomefloxacin (61.3%) Cefotaxime (52.6%)	East China	Sick chicken	(Afayibo <i>et al.</i> , 2022)
Tetracycline (50%) Ciprofloxacin (46.8%)	Polandia	Wild Birds	(Nowaczek et al., 2021)
Tetracycline (64%) Ampicillin (55.1%) Trimetoprim (49.4%)	Qatar	Pigeons	(Alhababi et al., 2020)
Trimetoprim (95.5%) Florfenicol (93.7%) Amoxicillin (93.3%) Doxycycline (92.2%) Spectinomycin (92.2%)	Yordania	Broiler chicken	(Ibrahim <i>et al.</i> , 2019)
Nalidixic acid (84.8%) Ciprofloxacin (69.6%) Tetracycline (58.4%) Chloramphenicol (56.2%)	Korea	Chicken	(Kim et al., 2021)
Tetracycline (97.3%) Ampicillin (97.3%) Sulfamethoxazole/trimethoprim (83.7%) Sulfisoxazole (81.1%) Cefoxitin (78.4%) Streptomycin (75.7%) Gentamicin (45.9%) Nalidixic acid (45.9%)	Nigeria	Chicken	(Aworh et al., 2020)
Ampicillin (97.1%) Nalidixic acid (77.1%) Trimetoprim (70.5%) Streptomycin (65.7%) Sulfamethoxazole/trimethoprim (63.8%) Chloramphenicol (56.2%)	Türkiye	Chicken	(Baran <i>et al.</i> , 2020)

cillin + clavulinic acid (71.4%), ciprofloxacin (57.1%), tetracycline (71.4%), trimethoprim-sulfamethoxazole (100%), chloramphenicol (85.7%), nalidixic acid (57.1%), and streptomycin (71.4%). Ranabhat *et al.* (2024) also reported AMR to gentamicin (90.2%) and tetracycline (67.2%). Hasan *et al.* (2024) found antimicrobial resistance to ampicillin (86%), ciprofloxacin (86%), and trimethoprim-sulfamethoxazole (73%).

Patel *et al.* (2024) found AMR to fluoroquinolone antibiotics, including enrofloxacin (72.46%), levofloxacin (69.56%), and ciprofloxacin (66.66%), followed by amoxyclav (63.77%) and tetracycline (59.42%). Khong *et al.* (2023) reported that *E. coli* isolates showed the greatest resistance to cephalothin (54.4%), tetracycline (27.9%), and streptomycin (29.4%). Salam *et al.* (2024) also reported varying levels of resistance in *E. coli* isolates, with the highest resistance found to amoxicillin-sulbactam (82%), followed by streptomycin (76%), cefotaxime and trimethoprim/ sulfamethoxazole (73%), tetracycline and ciprofloxacin (68%), chloramphenicol (66%), cefaclor (65%), and cefixime (62%).

MDR refers to bacteria that have become resistant to multiple types of antibiotics with different mechanisms of action. The term MDR applies to bacteria that are resistant to three or more different classes of antibiotics, rendering them more dangerous than before (Effendi *et al.*, 2021).

E. coli, as a commensal bacterium that has become multidrug-resistant, presents a significant health risk (Pradika *et al.*, 2019). This is due to *E. coli*'s ability to transfer resistance genes to other pathogenic bacteria within the digestive tract (Wibisono *et al.*, 2022). Putri *et al.* (2023) identified several risk factors for multidrug resistance in *E. coli*, including the type of farm, type of feed, veterinary support, chlorine levels in water, antibiotic programs, and references for antibiotic administration. The consequences of AMR and MDR include increased treatment costs, higher fatality, morbidity, and mortality rates associated with infectious diseases.

Transmission pattern of the Escherichia coli pathogen

Various transmission routes for *E. coli* pathogens from animals to humans, such as those carried by migratory birds, play a significant role in the spread of various human and animal pathogens, including bacteria, viruses, fungi, and parasites (Mihaela and Marina, 2014). Pathogenic bacteria can be transmitted to humans and animals through poultry or birds via contaminated water, particularly if the water is used for household or agricultural purposes (Islam *et al.*, 2021). The aquatic environment serves as a dynamic medium capable of transmitting pathogens over long distances (Chen *et al.*, 2020; Bombaywala *et al.*, 2021), due to the extensive movement of organisms, which creates a complex disease transmission network (Khairullah *et al.*, 2022).

Hernández-Vásquez *et al.* (2022) highlight water pollution as a major environmental issue affecting people's quality of life worldwide, with around two billion people still consuming water contaminated with harmful microorganisms. Sabar *et al.* (2022) indicate that water consumed by the public often contains human and animal feces, with *E. coli* acting as a key indicator of fecal contamination, potentially causing gastrointestinal infections. This contamination results from the consumption of water tainted by human and animal waste, often from unclean water sources. According to research by Swelum *et al.* (2021), *E. coli* can spread through direct contact with birds or other poultry, as well as through the respiratory tract, with bacteria capable of surviving in feces for several weeks. Vertical transmission can also occur, as when eggshells contaminated with feces carry pathogenic *E. coli*.

Bortolaia *et al.* (2010) explain that horizontal transmission occurs when infected poultry spread bacteria to healthy individuals through body fluids, aerosols, feathers, and contaminated droppings. In poultry farms for consumption, *E. coli* can be transmitted vertically to offspring. Vertically transmitted APEC (Avian Pathogenic *E. coli*) can increase mortality rates in chicks within the first week of life due to horizontal transmission in hatcheries and farms. Infected eggs from the reproductive tracts of poultry can spread APEC to embryos during incubation. Additionally, uninfected poultry can become exposed to the bacteria through contact with infected birds via feathers and droppings. These interactions serve as primary channels for the spread of APEC infection.

Arsène *et al.* (2022) note that interactions between humans and animals, both direct and indirect, can facilitate the transfer of antibiotic residues. Direct interactions occur when antibiotics used in animals or plants result in residues that enter the food supply after slaughter or harvest. Indirectly, antibiotic residues accumulate in water and soil through manure or human waste, which can then contaminate food, particularly vegetables, through watering or contact with animal waste.

Khairullah *et al.* (2024) identify the fecal-oral route as a primary mechanism for APEC transmission in poultry, where bacteria excreted through the feces of infected birds contaminate the environment, feed, and water sources. Chickens can ingest APEC through contaminated feed or water, or via direct contact with infected feces. Transmission can also occur through the respiratory tract or retrogradely through the vaginal/cloacal route. Research on colibacillosis cases shows that the fecal-oral and respiratory routes are the most common means of transmission. Additionally, there is evidence of vertical transmission of APEC from infected mother birds to their chicks via eggs, which can lead to increased mortality during the first week of life, as well as horizontal transmission in hatcheries.

Fathima *et al.* (2022) report high levels of antimicrobial resistance in chicks at hatching, which may stem from the gut microbiota of the birds (via vertical transmission) or from the hatchery environment. Horizontal transmission of bacteria occurs within and between groups of chickens, spreading widely from farms to the surrounding environment (Dame-Korevaar *et al.*, 2019; Osman *et al.*, 2018).

Avian Pathogenic Escherichia coli (APEC) in commercial poultry

APEC can act as either a primary or secondary pathogen in various poultry species, including broiler chickens, laying hens, turkeys, and ducks (Collingwood *et al.*, 2014; Hu *et al.*, 2022). This infection is syndromic and leads to colibacillosis, with *E. coli* as the causative agent. One method to determine the pathogenic potential of APEC strains is serotyping, which categorizes *E. coli* based on somatic (O), capsular (K), and flagellar (H) antigens. Certain O serotypes are particularly associated with *E. coli* pathogenicity (Hu *et al.*, 2022). Common serotypes linked to colibacillosis include O1:K1, O2:K1, O5, O8, O35, O150, and O78:K80 (Cordoni *et al.*, 2016).

E. coli is classified into eight phylogenetic groups, namely *E. coli* sensu stricto (A, B1, B2, C, D, E, F) and *E. coli* cryptic clade I. The majority of *E. coli* strains that infect humans (ExPEC) belong to groups B2 and D, while APEC is generally found in groups C, F, B1, and B2 (Jeong *et al.*, 2021). APEC enters the chicken's body via the oral or nasal tract, where it initially explores the mucosa of the digestive and reproductive tracts without immediately causing disease. However, APEC can invade the mucosal layer and spread to extra-intestinal organs such as the heart, liver, lungs, spleen, kidneys, and reproductive organs. This infection causes colibacillosis, which is characterized by high morbidity and mortality rates, production losses, and potential transmission to humans via food (Kathayat *et al.*, 2021; Sarowska *et al.*, 2019).

A significant and contagious disease caused by APEC infection in quail is septicemic colibacillosis (Matin *et al.*, 2017). While *E. coli* is normally part of the intestinal flora, certain strains are pathogenic and can lead to diseases that have substantial economic impacts (Hassan *et al.*, 2021). In 2011, *E. coli* infections caused losses estimated at \$500 billion globally (Kawecki *et al.*, 2012). Pathogenic *E. coli* can lead to peritonitis, pericarditis, airsacculitis, osteomyelitis, cellulitis, synovitis, salpingitis, yolk sac infection, and coliosticemia associated with APEC (Guabiraba and Schouler, 2015; Kunert-Filho *et al.*, 2015; Crémet *et al.*, 2015).

Colibacillosis is a fatal disease that poses a significant threat to the

poultry industry (Kabir, 2010). According to Hegazy *et al.* (2017), APEC causes severe diseases in consumed poultry such as quail, chickens, ducks, and turkeys. In quail, APEC infection can lead to hepatitis and pericarditis in birds aged 21 days and coligranulomatosis in quail aged 8-12 months (Ito *et al.*, 1990). Infected chickens show symptoms of watery diarrhea, anorexia, weakness, weight loss, and death (Kika *et al.*, 2023; Matin *et al.*, 2017). Ducks infected with colibacillosis often exhibit congested lungs and minor bleeding in the heart and air sacs (Stordeur *et al.*, 2004).

Escherichia coli survival enhancer gene

Colibacillosis is a disease caused by pathogenic *E. coli*. The pathogenic potential of *E. coli* is enhanced by the presence of virulence factors, which are regulated by specific gene codes (De Carli *et al.*, 2015). APEC-induced colibacillosis is associated with various genes, each contributing to the pathogenicity of *E. coli* and enabling it to infect poultry (Joseph *et al.*, 2023). According to Rezatofighi *et al.* (2021), avian *E. coli* can serve as a potential reservoir for ExPEC strains in humans, making it a potential zoonotic threat.

The virulence genes of APEC include *fimC*, *papC*, *tsh*, *mat*, *irp2*, *fyuA*, *hylF*, *iucD*, *iroN*, *iss*, *Cva/cvi*, *neuC*, *ompA*, *vat*, *ibeA*, and *ibeB* (Al-Kandari and Woodward, 2019; levy *et al.*, 2020; Newman *et al.*, 2021; Sgariglia *et al.*, 2019). As stated by Croxen and Finlay (2010), *E. coli* can become pathogenic with just a few key genes through a mix-and-match mechanism, which enables the bacteria to cause serious diseases.

APEC strains produce various virulence factors that aid in host tissue colonization, such as adhesins (encoded by *pap*C and *tsh*), iron uptake systems (*iut*A, *irp*2, *sit*, and *iro*N), serum resistance factors (*iss*, *omp*T, and *kps*II), toxins (*vat*), and other factors (cvi/cva and etsB) (Kathayat *et al.*, 2021; Paixao *et al.*, 2016). The virulence of APEC strains, including *omp*T, *iut*A, *iss*, *pap*-C, *cva/cvi*, and *tsh*, is commonly found in pathogenic *E. coli* strains from poultry (Subedi *et al.*, 2018). Furthermore, Kendek *et al.* (2024) reported that 60% of *E. coli* isolates possess the *hly*F gene, which is associated with colisepticaemia in poultry.

Escherichia coli virulence gene mechanism

The *fim*C gene is involved in the mechanism of adherence and colonization of epithelial cells (Islam *et al.*, 2021). Schwan (2011) noted that the *fim*C gene is linked to pyelonephritis. The *pap*C gene encodes pilin, which assists *E. coli* in adhering to the surface of epithelial cells in the urinary tract and other organs, facilitating colonization and infection (Choudhary and Madhwal, 2020; Mohamed *et al.*, 2014).

The *iss* gene is a virulence factor that enhances bacterial survival. The *iss* gene (Increased Serum Survival) has been found in the intestines of quail (Rodriguez-Siek *et al.*, 2005). This virulence gene is pathogenic and has zoonotic potential. The *iss* gene plays a critical role in the virulence of *E. coli* (Dissanayake *et al.*, 2014; Paixao *et al.*, 2016). In a study by Badouei *et al.* (2016), *iss* was found in 90.3% of septicemic quail isolates and 64.3% of fecal isolates. Similarly, Kafshdouzan *et al.* (2013) found the *iss* gene in 68.2% of APEC isolates and in 24.5% of fecal isolates. According to Khusnan and Prihtiyantoro (2020), *iss* was present in 61.9% of *E. coli* isolates from quail.

The *tsh* gene in avian APEC is associated with virulence and infection, often located on virulence plasmids related to ColV. The *tsh* gene influences the ability of *E. coli* to cause disease and plays a key role in the early stages of respiratory infections, helping the bacteria adapt and infect host cells more effectively (Dozois *et al.*, 2000). The *papC* gene (P fimbriae assembly protein C) helps bacteria attach to host cell surfaces in the respiratory, digestive, and urinary tracts (Mansoor and Salman, 2023; Mellata *et al.*, 2003).

The Vat gene, encoding vacuolating autotransporter toxin, secretes Vat, heat-stable enterotoxin 1 (AstA), and Colicin V (ColV), encoded by vat, astA, and cvaA/B (Mortezaei et al., 2013; Huja et al., 2014). The vat

gene allows the toxin to enter the bloodstream and damage blood vessels throughout the body (Soon-Gu *et al.*, 2008). Sharif *et al.* (2018) reported that the *vat* gene was present in 40% of APEC cases in poultry.

The *hly*F gene expressed by *hly*F-producing *E. coli* induces autophagy in eukaryotic cells. This phenotype is associated with increased production of outer membrane vesicles (OMVs) by *hly*F-expressing bacteria. The *hly*F protein has a predicted catalytic domain of the short-chain dehydrogenase/reductase superfamily, suggesting its role in promoting OMV production. Increased OMV production is linked to toxin release. The *hly*F gene is expressed during extraintestinal infections and contributes to the virulence of extraintestinal pathogenic *E. coli* in poultry, causing colibacillosis (Murase *et al.*, 2016; Thomrongsuwannakij *et al.*, 2020).

The *iro*N gene is commonly found in APEC strains, as it encodes siderophores like aerobactin, salmokelin, and yersiniabactin, which help absorb iron, promoting bacterial growth and development. The *iro*N gene's ability to access iron in blood serum is crucial, as *E. coli* can trigger sepsis and infect iron-deficient organs (Su *et al.*, 2016; Kathayat *et al.*, 2021).

Impact of residual antibiotics on humans and the environment

The poultry industry relies heavily on the use of various antibiotics, which has led to the emergence and spread of antimicrobial resistance (AMR) and multidrug resistance (MDR) in birds and poultry intended for consumption. This situation is worsened by the ability of bacteria to transfer genetic material that contains resistance traits. Such transfers can occur both vertically through genetic mutations, as well as horizontally via mechanisms like conjugation, transduction, and transformation, enabling the spread of resistance between bacteria (Yanestria *et al.*, 2022).

The consumption of poultry meat contaminated with resistant bacteria can lead to infections in humans that are harder to treat, increasing the risk of severe complications. Nhung *et al.* (2017) pointed out that the rising incidence of colibacillosis in poultry has led to an increased use of antibiotics for both treatment and prevention, which, unfortunately, has contributed to the growing issue of multidrug resistance (MDR).

Bacanlı and Başaran (2019) noted that antibiotic residues in poultry products can trigger and accelerate the development of bacterial resistance. These residues not only increase the risk of transmitting antibiotic-resistant bacteria to humans but can also cause allergic reactions, such as penicillin allergies, and lead to more serious health problems. For example, residues of substances like sulfamethazine, oxytetracycline, and furazolidone have been linked to cancer, while gentamicin can lead to nephropathy and anaphylactic shock. Additionally, chloramphenicol residues can cause bone marrow toxicity, mutagenic effects, and reproductive disorders in humans.

Research by Beyene (2016) highlighted that antibiotic residues can have severe hidden risks, such as being covalently bound to various intracellular components like glycogen, glutathione (GSH), DNA, RNA, proteins, and phospholipids, contributing to long-term health risks. Chloramphenicol residues have been specifically reported to potentially cause cancer (Shahid *et al.*, 2021). Other compounds, such as nitrofurans, nitroimidazoles, and quinoxalines, are known to interact with or bind to various intracellular molecules, posing additional health threats. Tetracycline residues have been associated with several health issues, including developmental impairments in fetuses, digestive disturbances, pro-inflammatory effects, cytotoxicity, and immune system disorders (Haque *et al.*, 2023).

These conditions make the treatment of disease-causing bacteria increasingly difficult and pose a significant threat to both animal and human health (Effendi *et al.*, 2022). Resistant bacteria can be transmitted from poultry to humans through the food chain, jeopardizing public health (Bennani *et al.*, 2020). Furthermore, waste from livestock containing antibiotics and resistant pathogens can contaminate soil and water sources, negatively impacting ecosystems (Iwu *et al.*, 2020). The rise of

resistant pathogens diminishes the effectiveness of existing antibiotics, making the treatment of infections more challenging and dangerous.

Comprehensive approaches to prevent and control APEC in poultry production

Maintaining clean poultry enclosures is a crucial first step in preventing APEC infections in poultry. According to research by Joseph *et al.* (2023), biosecurity plays a key role in the prevention and control of APEC in poultry farms. Proper sanitation helps minimize the buildup of harmful microorganisms, such as *E. coli*, within the cages and surrounding areas. The use of effective disinfectants to clean equipment, cages, and other poultry facilities is essential for preventing cross-contamination. Additionally, proper management of poultry waste and droppings is vital to prevent the spread of pathogens to the surrounding environment, including feed and water sources. Poulsen *et al.* (2017) emphasized the importance of routine disinfection in farm areas, hatcheries, and poultry maintenance rooms (brooders). The application of an all-in-all-out system can further limit the spread of infection. Moreover, eggs that fall onto the floor should be discarded or prevented from becoming contaminated by promptly collecting or cleaning them.

Proper ventilation in poultry houses helps reduce humidity and improves air circulation, which can lower the infection rates of pathogenic bacteria, including APEC. High humidity can facilitate the spread of airborne bacteria, worsening contamination in the house. As such, maintaining a good ventilation system and proper temperature control is critical for maintaining poultry health (Khairullah *et al.*, 2024).

Contaminated feed and water can serve as major transmission routes for APEC in poultry. Ensuring that these sources remain free from bacterial contamination is essential (Fancher *et al.*, 2020). Feed should be stored in a clean, dry environment to prevent contamination, while the quality of drinking water should be regularly monitored. Providing nutritious feed also strengthens poultry immunity against APEC infections (Kathayat *et al.*, 2021).

Vaccination is an effective method to reduce infection rates and enhance poultry immunity. Several vaccines specifically developed for APEC can be administered to reduce bacterial virulence and boost the immune response. Timely and appropriate vaccination is essential for preventing outbreaks of APEC-related diseases (Ghunaim *et al.*, 2014).

The use of antibiotics in poultry must be managed carefully and in line with medical guidelines to prevent the development of antibiotic resistance. Misuse or overuse of antibiotics can exacerbate AMR issues, potentially making APEC infections more difficult to treat in the future (Ranabhat *et al.*, 2024). Therefore, strict antibiotic use control policies are essential to minimize the risk of antibiotic-resistant bacteria.

Regular monitoring of poultry health is crucial for the early detection of APEC infections. A comprehensive health screening program, including laboratory testing of samples from suspected infected birds, is vital for early identification and timely treatment (Christensen *et al.*, 2021). Early detection can help contain the spread of infection and prevent greater economic losses. Healing (2009) noted that regular monitoring and surveillance are key to controlling primary disease agents.

To control APEC transmission in poultry, both horizontal (between individuals in the flock) and vertical (from parent to offspring) transmission routes must be managed (Joseph *et al.*, 2023). Implementing healthy flock management practices, such as isolating infected and healthy birds, can reduce transmission risk. Additionally, controlling infection in parent birds and using vaccines for parent stock can help prevent vertical transmission (Conan *et al.*, 2012).

Conclusion

AMR and MDR in APEC found in poultry intended for consumption present a significant threat. The presence of virulence genes in these

pathogens enhances their ability to cause severe infections. Antibiotic residues in poultry products, along with the growing resistance of bacteria to multiple antibiotics, represent a major issue that not only affects animal health but also has the potential to transfer to humans through the food chain. The spread of antibiotic-resistant APEC poses a serious risk, as these resistant bacteria can be released into the environment through livestock waste, contaminating soil and water. This increases the likelihood of antibiotic-resistant infections in humans and contributes to the global AMR and MDR crisis. Strict regulation of antibiotic use in poultry farming is crucial, along with enhanced prevention and control measures to mitigate the negative effects on both public health and the environment.

Acknowledgments

The authors thanks to Faculty of Veterinary Medicine, Universitas Airlangga. This study was partly supported by The Skema Penelitian Tesis Magister Airlangga, Universitas Airlangga, Surabaya, Indonesia, in 2025 (grant number: 1851/UN3.LPPM/PT.01.03/2025).

Conflict of interest

The authors have declared no conflict of interest.

References

Aarestrup, F., 2012. Get pigs off antibiotics. Nature 486, 465-466. doi: 10.1038/486465a.

- Abd El-Ghany, W.A., 2019. A comprehensive review on the common emerging diseases in quails. J. World's Poult. Res. 9, 160–174. doi: 10.36380/jwpr.2019.20. Abreu, R., Semedo-Lemsaddek, T., Cunha, E., Tavares, L., Oliveira, M., 2023. Antimicrobial Drug Re-
- Abreu, R., Semedo-Lemsaddek, T., Cunha, E., Tavares, L., Oliveira, M., 2023. Antimicrobial Drug Resistance in Poultry Production: Current Status and Innovative Strategies for Bacterial Control. Microorganisms 11, 953. doi: 10.3390/microorganisms11040953.
- Afayibo, D.J.A., Zhu, H., Zhang, B., Yao, L., Abdelgawad, H.A., Tian, M., Wang, S., 2022. Isolation, molecular characterization, and antibiotic resistance of avian pathogenic *Escherichia coli* in Eastern China. Vet. Sci. 9, 319. doi: 10.3390/vetsci9070319.
- Ahmed, S.K., Hussein, S., Qurbani, K., Ibrahim, R.H., Fareeq, A., Mahmood, K.A., Mohamed, M.G., 2024. Antimicrobial resistance: Impacts, challenges, and future prospects. J. Med. Surg. Public Health 2, 100081. doi: 10.1016/j.glmedi.2024.100081.
- Al-Kandari, F., Woodward, M.J., 2019. Genotypic and phenotypic diversity differences of presumptive commensal and avian pathogenic *E. coli*. Br. Poult. Sci. 60, 79–86. doi: 10.1080/00071668.2018.1544415.
- Alhababi, D.A., Eltai, N.O., Nasrallah, G.K., Farg, E.A., Al Thani, A.A., Yassine, H.M., 2020. Antimicrobial resistance of commensal *Escherichia coli* isolated from food animals in Qatar. Microb. Drug Resist. 26, 420–427. doi: 10.1089/mdr.2019.0402.
- Amir, M., Riaz, M., Chang, Y.F., Akhtar, S., Yoo, S.H., Sheikh, A.S., Kashif, M., 2017. Impact of Unhygienic Conditions during Slaughtering and Processing on Spread of Antibiotic Resistant *Escherichia coli* from Poultry. Microbiol. Res. 8, 7330. doi: 10.4081/mr.2017.7330.
 Anggita, M., Asmara, W., Untari, T., Wibowo, M.H., Artanto, S., Herawati, O., Wahyuni, A.E.T.H.,
- Anggita, M., Asmara, W., Untari, I., Wibowo, M.H., Artanto, S., Herawati, O., Wahyuni, A.L.I.H., 2021. Antibiotic resistance of bacteria from healthy quail cloacal swab. J. Vet. 22, 508–514. Arsène, M.M.J., Davares, A.K.L, Viktorovna, P.I., Andreevna, S.L., Sarra, S., Khelifi, I., Serqueiev.
- Arsene, M.M.J., Davares, A.K.L., Viktorovna, P.I., Andreevna, S.L., Sarra, S., Khelifi, I., Sergueievna, D.M., 2022. The public health issue of antibiotic residues in food and feed: Causes, consequences, and potential solutions. Vet. World 15, 662–671. doi: 10.14202/vetworld.2022.662–671.
- Aworh, M.K., Kwaga, J., Okolocha, E., Harden, L., Hull, D., Hendriksen, R.S., Thakur, S., 2020. Extended-spectrum β-lactamase-producing *Escherichia coli* among humans, chickens and poultry environments in Abuja, Nigeria. One Health Outlook. 2, 9. doi: 10.1186/s42522-020-00014-7.
- Bacanlı, M., Başaran, N., 2019. Importance of antibiotic residues in animal food. Food Chem. Toxicol. 125, 462–466. doi: 10.1016/j.fct.2019.01.033.
- Badouei, M.A., Blackall, P.J., Koochakzadeh, A., Nazarpak, H.H., Sepehri, M.A., 2016. Prevalence and clonal distribution of avian *Escherichia coli* isolates harboring increased serum survival (iss) gene. J. Appl. Poult. Res. 25, 67–73. doi: 10.3382/japr/pfv064.Baran, A., Adıgüzel, M., Yüksel, M., 2020. Prevalence of antibiotic-resistant and extended-spectrum
- Baran, A., Adiguzel, M., Yuksel, M., 2020. Prevalence of antibiotic-resistant and extended-spectrum beta-lactamase-producing *Escherichia coli* in chicken meat from eastern Turkey. Pak. Vet. J. 40, 2074–7764. doi: 10.29261/pakvetj/2020.047.
- Bennani, H., Mateus, A., Mays, N., Eastmure, E., Stärk, K.D., Häsler, B., 2020. Overview of evidence of antimicrobial use and antimicrobial resistance in the food chain. Antibiotics 9, 49. doi: 10.3390/antibiotics9020049.
- Beyene, T., 2016. Veterinary drug residues in food-animal products: its risk factors and potential effects on public health. J. Vet. Sci. Technol. 7, 1–7. doi: 10.4172/2157-7579.1000285.
 Bombaywala, S., Mandpe, A., Paliya, S., Kumar, S., 2021. Antibiotic resistance in the environment:
- Bornbaywara, S., Martobe, A., Paliya, S., Kurnar, S., 2021. Antibiotic resistance in the environment: a critical insight on its occurrence, fate, and eco-toxicity. Environ. Sci. Poll. Res. 28, 24889-24916. doi: 10.1007/s11356-021-13143-x.
 Bortolaia, V., Bisqaard, M., Bojesen, A.M., 2010. Distribution and possible transmission of ampicil-
- Bortolaia, V., Bisgaard, M., Bojesen, A.M., 2010. Distribution and possible transmission of ampicillin-and nalidixic acid-resistant *Escherichia coli* within the broiler industry. Vet. Microbial. 142, 379–386. doi: 10.1016/j.vetmic.2009.10.024.
- Chen, J., Sun, R., Pan, C., Sun, Y., Mai, B., Li, Q.X., 2020. Antibiotics and food safety in aquaculture. J. Agric. Food Chem. 68, 11908–11919. doi: 10.1021/acs.jafc.0c03996.
 Choudhary, S.V., Madhwal, A., 2020. Isolation and identification of *Escherichia coli* as secondary
- Choudhary, S.V., Madhwal, A., 2020. Isolation and identification of *Escherichia coli* as secondary infection in layer birds. Vet. Pract. 21, 174–178.
 Christensen, H., Bachmeier, J., Bisgaard, M., 2021. New strategies to prevent and con-
- Christensen, H., Bachmeier, J., Bisgaard, M., 2021. New strategies to prevent and control avian pathogenic *Escherichia coli* (APEC). Avian Pathol. 50, 370–381. doi: 10.1080/03079457.2020.1845300.
- Collingwood, C., Kemmett, K., Williams, N., Wigley, P., 2014. Is the Concept of Avian Pathogenic *Escherichia coli* as a Single Pathotype Fundamentally Flawed? Front. Vet. Sci. 1, 5. doi: 10.3389/fvets.2014.00005.
- Conan, A., Goutard, F.L., Sorn, S., Vong, S., 2012. Biosecurity measures for backyard poultry in developing countries: a systematic review. BMC Vet. Res. 8, 240. doi: 10.1186/1746-6148-8-240.Cordoni, G., Woodward, M.J., Wu, H., Alanazi, M., Wallis, T., La Ragione, R.M., 2016. Comparative
- Cordoni, G., Woodward, M.J., Wu, H., Alanazi, M., Wallis, T., La Ragione, R.M., 2016. Comparative genomics of European avian pathogenic *E. coli* (APEC). BMC Genomics 17, 960. doi: 10.1186/ s12864-016-3289-7.

- Crémet, L., Broquet, A., Brulin, B., Jacqueline, C., Dauvergne, S., Brion, R., Asehnoune, K., Corvec, S., Heymann, D., Caroff, N., 2015. Pathogenic potential of *Escherichia coli* clinical strains from orthopedic implant infections towards human osteoblastic cells. Pathog. Dis. 73, ftv065. doi: 10.1093/femspd/ftv065
- Croxen, M.A., Finlay, B.B. 2010. Molecular mechanisms of Escherichia coli pathogenicity. Nat. Rev. Microbiol. 8, 26-38, doi: 10.1038/nrmicro2265. Dame-Korevaar, A., Fischer, E.A., van der Goot, J., Stegeman, A., Mevius, D., 2019. Transmission
- routes of ESBL/pAmpC producing bacteria in the broiler production pyramid, a literature review. Prev. Vet. Med. 162, 136–150. doi: 10.1016/j.prevetmed.2018.12.002.
- De Carli, S., Ikuta, N., Lehmann, F.K., da Silveira, V.P., de Melo Pedrebon, G., Fonseca, A.S., Lunge, V.R., 2015. Virulence gene content in *Escherichia coli* isolates from poultry flocks with clinical
- signs of colibacillosis in Brazil Poult. Sci. 94, 2635–2640. doi: 10.3382/ps/pev256. Dissanayake, D.R., Octavia, S., Lan, R., 2014. Population structure and virulence contentof avian pathogenic *Escherichia coli* isolated from outbreaks in Sri Lanka. Vet. Microbiol. 168, 403–
- 412. doi: 10.1016/j.vetmic.2013.11.028. Dozois, C.M., Dho-Moulin, M., Brée, A., Fairbrother, J.M., Desautels, C., Curtiss, R., 2000. Relationship between the *Tsh* autotransporter and pathogenicity of avian *Escherichia coli* and local-ization and analysis of the *Tsh* genetic region. Infect. Immun. 68, 4145–4154. doi: 10.1128/ IAI.68.7.4145-4154.2000.
- Effendi, M.H., Faridah, H.D., Wibisono, F.M., Wibisono, F.J., Nisa, N., Fatimah, F., Ugbo, E.N., 2022. Detection of virulence factor encoding genes on *Escherichia coli* isolated from broiler chick-
- en in Blitar District, Indonesia. Biodiversitas 23, 3437–3442. doi: 10.13057/biodiv/d230716.
 Effendi, M.H., Tyasningsih, W., Yurianti, Y.A., Rahmahani, J., Harijani, N., Plumeriastuti, H., 2021. Presence of multidrug resistance (MDR) and extended-spectrum beta-lactamase (ESBL) of Escherichia coli isolated from cloacal swab of broilers in several wet markets in Surabaya, Indonesia Point Provider 20, 2010 and 2010 and 2010 and 2010.
- Indonesia. Biodiversitas 22, 304–310. doi: 10.13057/biodiv/d220137.
 Fancher, C.A., Zhang, L., Kiess, A.S., Adhikari, P.A., Dinh, T.T.N., Sukumaran, A.T., 2020. Avian Pathogenic *Escherichia coli* and Clostridium perfringens: Challenges in No Antibiotics Ever Broiler Production and Potential Solutions. Microorganisms 8, 1533. doi: 10.3390/microorganisms8101533.
- Farghaly, E.M., Samy, A., Roshdy, H., 2017. Wide prevalence of critically important antibiotic resis-tance in Egyptian quail farms with mixed infections. Vet. Sci. Res. Rev. 3, 17–24. doi: 10.17582/ journal.vsrr/2017.3.1.17.24.
- Fathima, S., Shanmugasundaram, R., Adams, D., Selvaraj, R.K., 2022. Gastrointestinal Microbiota and Their Manipulation for Improved Growth and Performance in Chickens. Foods 11, 1401. doi: 10.3390/foods11101401.
- Ghunaim, H., Abu-Madi, M.A., Kariyawasam, S., 2014. Advances in vaccination against avian pathogenic *Escherichia coli* respiratory disease: potentials and limitations. Vet. Microbiol. 172, 13– 22. doi: 10.1016/j.vetmic.2014.04.019.
- Guabiraba, R., Schouler, C., 2015. Avian colibacillosis: still many black holes. FEMS Microbiol. Lett. 362, fnv118. doi: 10.1093/femsle/fnv118.
- Haque, A.R., Sarker, M., Das, R., Azad, M.A.K., Hasan, M.M., 2023. A review on antibiotic residue in foodstuffs from animal source: global health risk and alternatives. Int. J. Environ. Anal. Chem. 103, 3704-3721. doi: 10.1080/03067319.2021.1912334.
- Hasan, B., Ali, M.Z., Rawlin, G., 2024. Avian Pathogenic Escherichia coli Isolated in Poultry Farms in Bangladesh that Use Antibiotics Extensively. Microb. Drug Resist. 30, 468–475. doi: 10.1089/ mdr.2024.0005.
- Hassan, M.M., El Zowalaty, M.E., Lundkvist, Å., Järhult, J.D., Nayem, M.R.K., Tanzin, A.Z., Ashour, Hassan, M.M., El Zowalay, M.E., Eurockis, A., Janiot, J.D., Nayen, M.E.K., Katzin, A.Z., Kshour, H.M., 2021. Residual antimicrobial agents in food originating from animals. Trends Food Sci. Technol. 111, 141–150. doi: 10.1016/j.itfs.2021.01.075.
 Healing, T., 2009. Surveillance and Control of Communicable Disease in Conflicts and Disasters. Conflict Catastrophe Med. 2009, 197–222. doi: 10.1007/978-1-84800-352-1_13.
 Hegazy, A.M., Ismail, A., Abdallah, H.A., Ibrahim, W.F.S., 2017. Incidence Of Pathogenic Eschesichia
- Ooli In Some Poultry Species In Egypt And Its Effect On Chickens Performance. Kafrelsheikh Vet. Med. J. 15, 11–28.
- Hernández-Vásquez, A., Visconti-Lopez, F.J., Vargas-Fernández, R., 2022. Escherichia coli contami-nation of water for human consumption and its associated factors in Peru: a cross-sectional
- Batori or water for finitial consumption and its associated factors in Pertu: a cross-sectional study. Am. J. Trop. Med. Hyg. 108, 187–194. doi: 10.4269/ajtmh.22-0240.
 Hidanah, S., Sabdoningrum, E.K., Chusniati, S., Nurliyani, N., Khairullah, A.R., Nayan, N., 2023. Effectiveness of Phyllanthus niruri and Andrographis paniculata Extracts on Egg Quality in Laying Hens with Avian Pathogenic *Escherichia coli*. J. Med. Vet. 6, 48–54. doi: 10.20473/jmv. articia.2020.46. Ed. vol6.iss3.2023.48-54.
- Voloissi 2023;40-34.
 Hu, J., Afayibo, D.J.A., Zhang, B., Zhu, H., Yao, L., Guo, W., Wang, X., Wang, Z., Wang, D., Peng, H., Tian, M., Qi, J., Wang, S., 2022. Characteristics, pathogenic mechanism, zoonotic potential, drug resistance, and prevention of avian pathogenic *Escherichia coli* (APEC). Front. Microbiol. 13, 1049391. doi: 10.3389/fmicb.2022.1049391.
- Huja, S., Oren, Y., Biran, D., Meyer, S., Dobrindt, U., Bernhard, J., Ron, E.Z., 2014. Fur is the master regulator of the extraintestinal pathogenic *Escherichia coli* response to serum. MBio 5, e01460-14. doi: 10.1128/mBio.01460-14.
- Ibrahim, R.A., Cryer, T.L., Lafi, S.Q., Basha, E.A., Good, L., Tarazi, Y.H., 2019. Identification of *Escherichia coli* from broiler chickens in Jordan, their antimicrobial resistance, gene characterization and the associated risk factors. BMC Vet. Res. 15, 159. doi: 10.1186/s12917-019-1901-1.
- Ibrahim, W.F., 2019. Isolation, identification and antimicrobial susceptibility testing of recent E. coli serotypes from Japanese Quails reared in Sharkia Governorate, Egypt. Damanhour J. Vet. Sci. 1. 14-17. doi: 10.21608/DJVS.38982.
- Islam, M. S., Sobur, M. A., Talukder, M., Rahman, M. B., Khan, M.F.R., Rahman, M.T., 2020. levy, S., Molecular detection of avian pathogenic *Escherichia coli* (APEC) for the first time in layer farms in Bangladesh and their antibiotic resistance patterns. Microorganisms 8, 1021. doi:
- 10.3300/microorganisms8071021.
 Islam, M.S., Nayeem, M.M.H., Sobur, M.A., Ievy, S., Islam, M.A., Rahman, S., Rahman, M. T., 2021.
 Virulence determinants and multidrug resistance of *Escherichia coli* isolated from migratory birds. Antibiotics 10, 190. doi: 10.3390/antibiotics10020190.
- Ito, H., Okoda, S., Kobayashi, S., Sugiyama, H., Masanori, N., 1990. Colibacillosis of Japanese quail (Coturnix coturnix japonica) occurring in Higashimikawa District. J. Japan. Vet. Med. Assoc. 43, 661–665. doi: 10.12935/jvma1951.43.661.
- W. C.D., Korsten, L., Okoh, A.I., 2020. The incidence of antibiotic resistance within and beyond the agricultural ecosystem: A concern for public health. Microbiologyopen 9, 1035. doi: 10.1002/ mbo3.1035
- Jeong, J., Lee, J.Y., Kang, M.S., Lee, H.J., Kang, S.I., Lee, O.M., Kwon, Y.K., Kim, J.H., 2021. Comparative characteristics and zoonotic potential of Avian Pathogenic *Escherichia coli* (APEC) isolates from chicken and duck in south Korea. Microorganisms 9, 946. doi: 10.3390/microorganisms9050946.
- Joseph, J., Zhang, L., Adhikari, P., Evans, J.D., Ramachandran, R., 2023. Avian Pathogenic Esche-richia coli (APEC) in Broiler Breeders: An Overview. Pathogens 12, 1280. doi: 10.3390/pathogens12111280.
- Kabir, S.M.L., 2010, Avian colibacillosis and salmonellosis: a closer look at epidemiology, pathogenesis, diagnosis, control and public health concerns. Int. J. Environ. Res. Public Health 7, 89–114. doi: 10.3390/ijerph7010089. Kafshdouzan, K., Zahraei, S. T., Nayeri, F. B., Madadgar, O., Yamasaki, S., Hinenoya, A., Yasuda, N.,
- 2013. Distribution of virulence associated genes in isolated *Escherichia coli* from avian colibacillosis. Iran. J. Vet. Med. 7, 1–6.
- Kalule, J.B., Keddy, K.H., Nicol, M.P., 2018. Characterisation of STEC and other diarrheic *E. coli* isolated on CHROMagar[™]STEC at a tertiary referral hospital, Cape Town. BMC Microbiol. 18, 55. doi: 10.1186/s12866-018-1195-7. Karpov, D.S., Kazakova, E.M., Kovalev, M.A., Shumkov, M.S., Kusainova, T., Tarasova, I.A., Gonchar-
- enko, A.V., 2024. Determinants of antibiotic resistance and virulence factors in the genome of Escherichia coli APEC 36 strain isolated from a broiler chicken with generalized colibacillosis.

Antibiotics 13, 945. doi: 10.3390/antibiotics13100945.

- Kathayat, D., Lokesh, D., Ranjit, S., Rajashekara, G., 2021. Avian Pathogenic Escherichia coli (APEC): Kathayat, D., Dokesh, D., Karjiti, S., Kajashekara, G., 2021. Avian Pathogenic Escharticitic Out (APEC). An overview of virulence and pathogenesis factors, zoonotic potential, and control strategies. Pathogens 10, 467. doi: 10.3390/pathogens10040467.
 Kawecki, T.J., Lenski, R.E., Ebert, D., Hollis, B., Olivieri, I., Whitlock, M.C., 2012. Experimental evolution. Trend. Eco. Evo. 27, 547–560. doi: 10.1016/j.tree.2012.06.001.
 Kendek, I.A., Putri, M.F.R., Wibisono, F.J., Effendi, M.H., Tyasningsih, W., Ugbo, E.N., Agumah, N.B., 2004. Adjustice dotted in a full dotted in a full
- 2024. Molecular detection of *hly*F gene on multidrug resistance of avian pathogenic *Escherichia coli* isolated from ducks on wet markets of Surabaya, Indonesia. Biodiversitas 25, , 1246–1253. doi: 10.13057/biodiv/d250341. Khairullah, A.R., Afnani, D.A., Riwu, K.H.P., Widodo, A., Yanestria, S.M., Moses, I.B., Effendi, M.H.,
- Ramandinianto, S.C., Wilowo, S., Fauziah, I., Kusala, M.L.J., Fauzia, K.A., Furqoni, A.H., Raissa, R., 2024. Avian pathogenic *Escherichia coli*: Epidemiology, virulence and pathogenesis, diagnosis, pathophysiology, transmission, vaccination, and control. Vet. World 17, 2747–2762. doi: 10.14202/vetworld.2024.2747-2762. Khairullah, A.R., Rehman, S., Sudjarwo, S.A., Effendi, M.H., Ramandinianto, S.C., Gelolodo, M.A.,
- Widodo, A., Riwu, K.H.P., Kurniawati, D.A., 2022. Detection of mecA gene and methicillin-re-sistant Staphylococcus aureus (MRSA) isolated from milk and risk factors from farms in
- Probolinggo, Indonesia. F1000Res. 11, 722. doi: 10.12688/f1000research.122225.3.
 Khong, M.J., Snyder, A.M., Magnaterra, A.K., Young, M.M., Barbieri, N.L., Weimer, S.L., 2023. Antimicrobial resistance profile of *Escherichia coli* isolated from poultry litter. Poult. Sci. 102, 102305. doi: 10.1016/j.psj.2022.102305. Khusnan, K., Prihtiyantoro, W., 2020. Characterization of Virulent Factors of Sorbitol-Negative
- Kika, T.S., Cocoli, S., Pelić, D.L., Puvača, N., Lika, E., Pelić, M., 2023. Colibacillosis in Modern Poultry
- Kim, H., Kim, Y.A., Seo, Y.H., Lee, H., Lee, K., 2021. Prevalence and molecular epidemiology of extended-spectrum-β-lactamase (ESBL)-producing *Escherichia coli* from multiple sectors of poultry industry in Korea. Antibiotics 10, 1050. doi: 10.3390/antibiotics10091050. Kunert-Filho, H.C., Carvalho, D., Grassotti, T.T., Soares, B.D., Rossato, J.M., Cunha, A,C., Brito, K.C.T.,
- Cavalli, L.S., Brito, B.G., 2015. Avian pathogenic *Escherichia coli* methods for improved diag-nosis. World's Poult. Sci. J. 71, 249–258. doi: 10.1017/S0043933915000264.
- Li, R., Li, N., Zhang, J., Wang, Y., Liu, J., Cai, Y., Chai, T., Wei, L., 2016. Expression of Immune-Related Genes of Ducks Infected with Avian Pathogenic Escherichia coli (APEC). Front. Microbiol. 7,
- G37. doi: 10.3389/fmicb.2016.00637.
 Lim, M.A., Kim, J.Y., Acharya, D., Bajgain, B.B., Park, J.H., Yoo, S.J., Lee, K., 2020. A diarrhoeagenic enteropathogenic *Escherichia coli* (EPEC) infection outbreak that occurred among elementary school children in Gyeongsangbuk-do province of South Korea was associated with consumption of water-contaminated food items. Int. J. Environ. Res. Public Health 17, 3149. doi: 10.3390/ijerph17093149. Mancuso, G., Midiri, A., Gerace, E., Biondo, C., 2021. Bacterial Antibiotic Resistance: The Most Crit-
- ical Pathogens. Pathogens 10, 1310. doi: 10.3390/pathogens10101310. Mansoor, D.K., Salman, R.A., 2023. Detection Of P Fimbriae Genes In *E. coli* Isolated From Urinary
- Tract Infection. Semiconductor Optoelectronics 42, 270-284.
- Matin Mat, Islam M.A., Khatun, M.M., 2017. Prevalence of colibacillosis in chickens in greater My-mensingh district of Bangladesh. Vet. World 10, 29–33. doi: 10.14202/vetworld.2017.29-33.
- Mellata, M., Dho-Moulin, M., Dozois, C.M., Curtiss, R. 3rd, Lehoux, B., Fairbrother, J.M., 2003. Role of avian pathogenic *Escherichia coli* virulence factors in bacterial interaction with chicken heterophils and macrophages. Infect. Immun. 71, 494–503. doi: 10.1128/IAI.71.1.494-503.2003. Mihaela, N., Marina, S., 2014. Wild birds as potential vectors for pathogen dissemination on mi-
- Gration routes in the Danube Delta Wetlands. Int. J. Curr. Microbiol. Appl. Sci. 3, 890–897.
 Mohamed, M.A., Shehata, M.A., Rafeek, E., 2014. Virulence Genes Content and Antimicrobial Resistance in *Escherichia coli* from Broiler Chickens. Vet. Med. Int. 2014, 195189. doi: 10.1155/2014/195189.
- Mortezaei, N., Singh, B., Bullitt, E., Uhlin, B.E., Andersson, M., 2013. P-fimbriae in the presence of anti-*PapA* antibodies: new insight of antibodies action against pathogens. Sci. Rep. 3, 3393. doi: 10.1038/srep03393.
- Murase, K., Martin, P., Porcheron, G., Houle, S., Helloin, E., Pénary, M., Nougayrède, J.P., Dozois, C.M., Hayashi, T., Oswald, E., 2016. *HlyF* produced by extraintestinal pathogenic *Escherichia coli* is a virulence factor that regulates outer membrane vesicle biogenesis. J. Infect. Dis. 213, 856–865. doi: 10.1093/infdis/jiv506. Murray, C.J., Ikuta, K.S., Sharara, F., Swetschinski, L., Aguilar, G.R., Gray, A., Tasak, N., 2022. Glob-
- al burden of bacterial antimicrobial resistance in 2019: a systematic analysis. Lancet 399, 629–655. doi: 10.1016/S0140-6736(21)02724-0.
- Newman, D.M., Barbieri, N.L., de Oliveira, A.L., Willis, D., Nolan, L.K., Logue, C.M., 2021. Characteriz-ing avian pathogenic *Escherichia coli* (APEC) from colibacillosis cases, 2018. PeerJ 9, e11025. doi: 10.7717/peerj.11025.
- Nhung, N.T., Chansiripornchai, N., Carrique-Mas, J.J., 2017. Antimicrobial Resistance in Bacterial Poultry Pathogens: A Review. Front. Vet. Sci. 4, 126. doi: 10.3389/fvets.2017.00126.
- Nowaczek, A., Dec, M., Stępień-Pyśniak, D., Urban-Chmiel, R., Marek, A., Różański, P., 2021. Anti-biotic resistance and virulence profiles of *Escherichia coli* strains isolated from wild birds in
- Poland. Pathogens 10, 1059. doi: 10.3390/pathogens10081059.
 Nwobodo, D.C., Ugwu, M.C., Anie, C.O., Al-Ouqaili, M.T.S., Ikem, J.C., Chigozie, U.V., Saki, M., 2022.
 Antibiotic resistance: The challenges and some emerging strategies for tackling a global menace. J. Clin. Lab. Anal. 36, e24655. doi: 10.1002/jcla.24655.
- Osman, K.M., Kappell, A.D., Elhadidy, M., ElMougy, F., El-Ghany, W.A.A., Orabi, A., Yousef, H.M.,
- Osman, K.M., Kappell, A.D., Elhadidy, M., ElMougy, F., El-Ghany, W.A.A., Orabi, A., Yousef, H.M., 2018. Poultry hatcheries as potential reservoirs for antimicrobial-resistant *Escherichia coli*: A risk to public health and food safety. Sci. Rep. 8, 5859. doi: 10.1038/s41598-018-23962-7.
 Paixao, A.C., Ferreira, A.C., Fevereiro, M., Fontes, M., Martins, L., Themudo, P., Albuquerque, T., de Sa, M.I.C., 2016. Detection of virulence-associated genes in pathogenic and commensal avian *Escherichia coli* isolates. Poult. Sci. 95, 1646–1652. doi: 10.3382/ps/pew087.
 Patel, S.S., Patel, A.C., Mohapatra, S.K., Chauhan, H.C., Sharma, K.K., Shrimali, M.D., Prajapati, B.I., 2024. Antibiotic resistance and virulence gene patterns associated with multi drug resistant avian pathogenic *Escherichia coli* (APEC) isolated from broiler chickens in India. Indian J. Microbiol. 64, 917–926. doi: 10.1007/s12088-023-01132-2.
- Peterson, E., Kaur, P., 2018. Antibiotic resistance mechanisms in bacteria: relationships between
- resistance determinants of antibiotic producers, environmental bacteria, and clinical pathogens. Front. Microbiol. 9, 2928. doi: 10.3389/fmicb.2018.02928.
 Pokharel, P., Dhakal, S., Dozois, C.M., 2023. The Diversity of *Escherichia coli* Pathotypes and Vaccination Strategies against. This Versatile Bacterial Pathogen. Microorganisms 11, 344. doi: 10.0302/fmicb.2018.02924.
- 10.3390/microorganisms11020344. Poulsen, L.L., Thøfner, I., Bisgaard, M., Christensen, J.P., Olsen, R.H., Christensen, H., 2017. Longitudinal study of transmission of *Escherichia coli* from broiler breeders to broilers. Vet. Microbi-ol. 207, 13–18. doi: 10.1016/j.vetmic.2017.05.029.
- Fradika, A.Y., Chusniati, S., Purnama, M.T.E., Effendi, M.H., Yudhana, A., Wibawati, P.A., 2019. Total Test of *Escherichia coli* on Fresh Cow Milk at Dairy Farmer Cooperative (*KpsP*) Karyo Ngr-emboko Purwoharjo Banyuwangi. J. Med. Vet. 2, 1–6. doi: 10.20473/jmv.vol2.iss1.2019.1-6.
- Putri, M.F.R., Kendek, I.A., Wibisono, F.J., Effendi, M.H., Rahardjo, D., Tyasningsih, W., Ugbo, E.N., 2023. Molecular detection of iron gene on multidrug resistant avian fecal *Escherichia coli* isolated from broiler on traditional markets, Surabaya, Indonesia. Biodiversitas 24, 6454-6460. doi: 10.13057/biodiv/d241207.
- Rahman, M.T., Sobur, M.A., Islam, M.S., Ievy, S., Hossain, M.J., El Zowalaty, M.E., Ashour, H.M., 2020. Zoonotic diseases: etiology, impact, and control. Microorganisms 8, 1405. doi: 10.3390/microorganisms8091405
- Ranabhat, G., Subedi, D., Karki, J., Paudel, R., Luitel, H., Bhattarai, R.K., 2024. Molecular detection of avian pathogenic Escherichia coli (APEC) in broiler meat from retail meat shop. Heliyon 10, e35661. doi: 10.1016/j.heliyon.2024.e35661.

- Rezatofighi, S.E., Najafifar, A., Badouei, M.A., Peighambari, S.M., Soltani, M., 2021. An integrated perspective on virulence-associated genes (VAGs), antimicrobial resistance (AMR), and phylogenetic clusters of pathogenic and non-pathogenic avian *Escherichia coli*. Front. Vet. Sci. 8, 758124. doi: 10.3389/fvets.2021.758124.
- Rodriguez-Siek, K.E., Giddings, C.W., Doetkott, C., Johnson, T.J., Nolan, L.K., 2005. Characterizing the APEC pathotype. Vet. Res. 36, 241–256. doi: 10.1051/vetres:2004057.Roess, A.A., Winch, P.J., Ali, N.A., Akhter, A., Afroz, D., El Arifeen, S., 2013. Animal husbandry practic-
- Koess, A.A., Winch, P.J., Ali, N.A., Akhter, A., Atroz, D., El Anteen, S., 2013. Animal husbandry practices in rural Bangladesh: potential risk factors for antimicrobial drug resistance and emerging diseases. Am. J. Trop. Med. Hyg. 89, 965–970. doi: 10.4269/ajtmh.12-0713.
- Sabar, M.A., Honda, R., Haramoto, E., 2022. CrAssphage as an indicator of human-fecal contamination in water environment and virus reduction in wastewater treatment. Water Res. 221, 118827. doi: 10.1016/j.watres.2022.118827.Salam, H.S., El-Ela, F.I.A., Hamra, S.A., Ismail, I.I., 2024. Antimicrobial resistance and virulence fac-
- Salam, H.S., El-Ela, F.I.A., Hamra, S.A., Ismail, I.I., 2024. Antimicrobial resistance and virulence factors in chicken-derived *E. coli* isolates. J. Adv. Vet. Res. 14, 48–54.
- Salam, M.A., Al-Amin, M.Y., Salam, M.T., Pawar, J.S., Akhter, N., Rabaan, A.A., Alqumber, M.A.A., 2023. Antimicrobial Resistance: A Growing Serious Threat for Global Public Healthcare (Basel) 11, 1946. doi: 10.3390/healthcare11131946.Santos, A.C.D.M., Santos, F.F., Silva, R.M., Gomes, T.A.T., 2020. Diversity of hybrid-and hete-
- Santos, A.C.D.M., Santos, F.F., Silva, R.M., Gomes, T.A.T., 2020. Diversity of hybrid-and hetero-pathogenic *Escherichia coli* and their potential implication in more severe diseases. Front. Cell. Infect. Microbiol. 10, 339. doi: 10.3389/fcimb.2020.00339.Saraiva, M.M.S., Lim, K., do Monte, D.F.M., Givisiez, P.E.N., Alves, L.B.R., Neto, O.C.F., Kariuki, S.,
- Saraiva, M.M.S., Lim, K., do Monte, D.F.M., Givisiez, P.E.N., Alves, L.B.R., Neto, O.C.F., Kariuki, S., Júnior, A.B., de Oliveira, C.J.B., Gebreyes, W.A., 2022. Antimicrobial resistance in the globalized food chain: a One Health perspective applied to the poultry industry. Braz. J. Microbiol. 53, 465–486. doi: 10.1007/s42770-021-00635-8.
- Sarowska, J., Futoma-Koloch, B., Jama-Kmiecik, A., Frej-Madrzak, M., Ksiazczyk, M., Bugla-Ploskonska, G., Choroszy-Krol, I., 2019. Virulence factors, prevalence and potential transmission of extraintestinal pathogenic *Escherichia coli* isolated from different sources: recent reports. Gut Pathog. 11, 10. doi: 10.1186/s13099-019-0290-0.
- Schwan, W.R., 2011. Regulation of *fim* genes in uropathogenic *Escherichia coli*. World J. Clin. Infect. Dis. 1, 17–25. doi: 10.5495/wjcid.v1.i1.17.
- Sgariglia, E., Mandolini, N.A., Napoleoni, M., Medici, L., Fraticelli, R., Conquista, M., Perugini, G., 2019. Antibiotic resistance pattern and virulence genes in avian pathogenic *Escherichia coli* (APEC) from different breeding systems. Vet. Ital. 55, 27–33. doi: 10.12834/Vetlt.1617.87011.
- Shahid, A., Ali, M.A., Muzammil, S., Aslam, B., Shahid, M., Saqalein, M., Khurshid, M., 2021. Antibiotic residues in food chains; impact on the environment and human health: a review. Appl. Ecol. Environ. Res. 19, 3959–3977. doi: 10.15666/aeer/1905_39593977.
- Sharif, H., Javed, M.T., Ghafoor, H., Younis, M., Khan, S.U., Rehman, A., Rafique, A., 2018. Association of pathogenicity genes (cvaC, iss, *iut*A, Stx1A, Stx2A and Vat) of *E. coli* with gross and histopathological lesions of colibacillosis in broilers. Biomed. Lett. 4, 40–46.
- Siahaan, S., Herman, M.J., Fitri, N., 2022. Antimicrobial Resistance Situation in Indonesia: A Challenge of Multisector and Global Coordination. J. Trop. Med. 2022, 2783300. doi: 10.1155/2022/2783300.

Soon-Gu, K., Se-Yeoun, C., Eun-Ju, C., Kim, B., Hee-Jong, S., Hyung-Kwan, J., 2008. Epidemiolog-

ical Prevalence of Avian Pathogenic *Escherichia coli* Differentiated by Multiplex PCR from Commercial Chickens and Hatchery in Korea. J. Bacteriol. Virol. 38, 179–188. doi: 10.4167/jbv.2008.38.4.179.

- Stordeur, P., Brée, A., Mainil, J., Moulin-Schouleur, M., 2004. Pathogenicity of *pap*-negative avian *Escherichia coli* isolated from septicaemic lesions. Microbes Infect. 6, 637–645. doi: 10.1016/j. micinf.2004.03.006.
- Su, Q., Guan, T., Lv, H., 2016. Siderophore biosynthesis coordinately modulated the virulence-associated interactive metabolome of uropathogenic *Escherichia coli* and human urine. Sci. Rep. 6, 24099. doi: 10.1038/srep24099.
- Subedi, M., Bhattarai, R.K., Devkota, B., Phuyal, S., Luitel, H., 2018. Antibiotic resistance pattern and virulence genes content in avian pathogenic *Escherichia coli* (APEC) from broiler chickens in Chitwan, Nepal. BMC Vet. 14, 166. doi: 10.1186/s12917-018-1442-z.
- Swelum, A.A., Elbestawy, A.R., El-Saadony, M.T., Hussein, E.O.S., Alhotan, R., Suliman, G.M., Taha, A.E., Ba-Awadh, H., El-Tarabily, K.A., Abd El-Hack, M.E., 2021. Ways to minimize bacterial infections, with special reference to *Escherichia coli*, to cope with the first-week mortality in chicks: an updated overview. Poult. Sci. 100, 101039. doi: 10.1016/j.psj.2021.101039.
- Thomrongsuwannakij, T., Blackall, P.J., Djordjevic, S.P., Cummins, M.L., Chansiripornchai, N., 2020. A comparison of virulence genes, antimicrobial resistance profiles and genetic diversity of avian pathogenic *Escherichia coli* (APEC) isolates from broilers and broiler breeders in Thailand and Australia. Avian Pathol. 49, 457–466. doi: 10.1080/03079457.2020.1764493.Van Boeckel, T.P., Brower, C., Gilbert, M., Grenfell, B.T., Levin, S.A., Robinson, T.P., Laxminarayan,
- Van Boeckel, T.P., Brower, C., Gilbert, M., Grenfell, B.T., Levin, S.A., Robinson, T.P., Laxminarayan, R., 2015. Global trends in antimicrobial use in food animals. Proc. Natl. Acad. Sci. U S A 112, 5649–5654. doi: 10.1073/pnas.1503141112.
- Vanstokstraeten, R., Crombé, F., Piérard, D., Moral, A.C., Wybo, I., De Geyter, D., Janssen, T., Caljon, B., Demuyser, T., 2022. Molecular characterization of extraintestinal and diarrheagenic *Escherichia coli* blood isolates. Virulence 13, 2032–2041. doi: 10.1080/21505594.2022.2147735
- erichia coli blood isolates. Virulence 13, 2032–2041. doi: 10.1080/21505594.2022.2147735. Watts, A., Wigley, P., 2024. Avian Pathogenic *Escherichia coli*: An Overview of Infection Biology, Antimicrobial Resistance and Vaccination. Antibiotics (Basel) 13, 809. doi: 10.3390/antibiotics13090809.
- Wibisono, F.J., Effendi, M.H., Wibisono, F.M., 2022. Occurrence, antimicrobial resistance, and potential zoonosis risk of avian pathogenic *Escherichia coli* in Indonesia: A review. Int. J. One Health 8, 76–85. doi: 10.14202/IJOH.2022.76-85.
- Widodo, A., Lamid, M., Effendi, M.H., Khairullah, A.R., Riwu, K.H.P., Yustinasari, L.R., Kurniawan, S.C., Ansori, A.N.M., Silaen, O.S.M., Dameanti, F.N.A.E.P., 2022. Antibiotic sensitivity profile of multidrug-resistant (MDR) *Escherichia coli* isolated from dairy cow's milk in Probolinggo, Indonesia. Biodiversitas 23, 4971–4976. doi: 10.13057/biodiv/d231002.
- Xu, C., Kong, L., Gao, H., Cheng, X., Wang, X., 202. A Review of Current Bacterial Resistance to Antibiotics in Food Animals. Front. Microbiol. 13, 822689. doi: 10.3389/fmicb.2022.822689. Yanestria, S.M., Dameanti, F.N.A.E.P., Musayannah, B.G., Pratama, J.W.A., Witaningrum, A.M., Effen-
- Yanestria, S.M., Dameanti, F.N.A.E.P., Musayannah, B.G., Pratama, J.W.A., Witaningrum, A.M., Effendi, M.H., Ugbo, E.N., 2022. Antibiotic resistance pattern of Extended-Spectrum β-Lactamase (ESBL) producing *Escherichia coli* isolated from broiler farm environment in Pasuruan district, Indonesia. Biodiversitas 22, 4460–4465. doi: 10.13057/biodiv/d230911.